201-1604A

IUCLID

Data Set

Existing Chemical

CAS No. **EINECS Name** EC No. **TSCA Name**

Molecular Formula

: ID: 67-68-5 67-68-5

dimethyl sulfoxide 200-664-3

: Methane, sulfinylbis-: C2H6OS

Producer related part

Company Creation date : ATOFINA Chemicals Inc. : 28.05.2003

Substance related part

Company Creation date : ATOFINA Chemicals Inc.

: 28.05.2003

Status Memo

Printing date Revision date : 17.05.2004 : 14.05.2004

Date of last update Number of pages

: 157

Chapter (profile) Reliability (profile)

: Chapter: 1, 2, 3, 4, 5, 6, 7, 8, 10 Reliability: without reliability, 1, 2, 3, 4 Flags (profile)

Material Safety Dataset, Risk Assessment, Directive 67/548/EEC, SIDS

Flags: without flag, confidential, non confidential, WGK (DE), TA-Luft (DE),

52

ld 67-68-5 Date 17.05.2004

1.0.1 APPLICANT AND COMPANY INFORMATION

lead organisation Type

Name Dimethyl Sulfoxide Producers Association

Contact person Betty Hunt Date

941 Rhonda Place SE 20175 Lessburg, Va Street Town **United States** Country Phone 703-669-5688 Telefax

Telex 703-669-5689

Cedex

ehunt@adelphia.com Email

Homepage

29.05.2003

Type cooperating company

Name

Contact person

Date 4-8, cours Michelet La Défense 10 Street

95091 Paris La Défense Cedex Town Country France

Phone

Telefax Telex Cedex

Email Homepage

Source : Atofina Paris La Défense Cedex

28.05.2003

Type cooperating company ATOFINA Chemicals Inc. Name

Contact person

Date Street

2000 Market Street PA 19103 Philadelphia

Town Country **United States**

Phone

Telefax Telex Cedex

Email Homepage

12.08.2003

Type cooperating company Gaylord Chemical Corporation Name

Contact person John Ferguson

Date Street

Town 70427 Bogalusa, LA Country United States

Phone

ld 67-68-5 **Date** 17.05.2004

Telefax Telex Cedex Email Homepage

12.08.2003

Type : cooperating company

Name : Toray Fine Chemicals Company, Ltd.

Contact person : Katsuhiro Shibayama

Date : Street : 8-1, Mihama 1-chome

Town : 279-8555 Urayasu, ChibaShiga

Country : Japan

Phone : +81 (47) 350-6174
Telefax : +81 (47) 350-6091

Telex Cedex

Email : katsuhiro_shibayama@tfc.toray.co.jp

Homepage

10.07.2003

1.0.2 LOCATION OF PRODUCTION SITE, IMPORTER OR FORMULATOR

1.0.3 IDENTITY OF RECIPIENTS

1.0.4 DETAILS ON CATEGORY/TEMPLATE

1.1.0 SUBSTANCE IDENTIFICATION

IUPAC Name

Smiles Code :

Molecular formula : C2H6OS Molecular weight : 78.13

Petrol class

: "

Source : Atofina, Paris-le-Défense, France.

Atofina Paris La Défense Cedex

27.12.2002

1.1.1 GENERAL SUBSTANCE INFORMATION

Purity type : typical for marketed substance

Substance type
Physical status
Purity
Colour

Substance type
Subst

Odour :

Jui

Source : Atofina, Paris-le-Défense, France.

Atofina Paris La Défense Cedex

27.12.2002

ld 67-68-5 **Date** 17.05.2004

1.1.2 SPECTRA

1.2 SYNONYMS AND TRADENAMES

dimethyl sulphoxide

22.07.2003

Sulfinylbis(methane); methyl sulfoxide; DMSO; SQ 9453; DMS-70; DMS-90; deltan; Demasorb; Demavet; Demeso; Dermasorb; Dolicur; Domoso; Dromisol; Gamasol 90; Hyadur; Infiltrina, Rimso-50; Somipront; Syntexan; Topsym (rescinded)

Source

: Atofina, Paris-le-Défense, France.

Atofina Paris La Défense Cedex

27.12.2002

(144)

1.3 IMPURITIES

1.4 ADDITIVES

1.5 TOTAL QUANTITY

1.6.1 LABELLING

Labelling Specific limits : no labelling required (no dangerous properties)

Jeonne minis

: Atofina, Paris-le-Défense, France. Atofina Paris La Défense Cedex

າດວ

09.12.2002

Source

1.6.2 CLASSIFICATION

Classified :

Class of danger R-Phrases

Specific limits

1st Concentration

2nd Concentration

3rd Concentration

4th Concentration 5th Concentration 6th Concentration 7th Concentration

8th Concentration 1st Classification 2nd Classification 3rd Classification

4th Classification 5th Classification : no classification required (no dangerous properties)

ld 67-68-5 Date 17.05.2004

6th Classification 7th Classification 8th Classification

:

Source

: Atofina, Paris-le-Défense, France. Atofina Paris La Défense Cedex

09.12.2002

- 1.6.3 PACKAGING
- 1.7 USE PATTERN
- 1.7.1 DETAILED USE PATTERN
- 1.7.2 METHODS OF MANUFACTURE
- 1.8 REGULATORY MEASURES
- 1.8.1 OCCUPATIONAL EXPOSURE LIMIT VALUES
- 1.8.2 ACCEPTABLE RESIDUES LEVELS
- 1.8.3 WATER POLLUTION
- 1.8.4 MAJOR ACCIDENT HAZARDS
- 1.8.5 AIR POLLUTION
- 1.8.6 LISTINGS E.G. CHEMICAL INVENTORIES
- 1.9.1 DEGRADATION/TRANSFORMATION PRODUCTS
- 1.9.2 COMPONENTS
- 1.10 SOURCE OF EXPOSURE

Remark

 Continuous process. Oxydation of Dimethylsulfide with N2O4/NO2 as catalyst. Purification by distillation after neutralisation (NaOH).

ld 67-68-5 Date 17.05.2004

Effluents: water treatment plant

Losses of product can only occur if problem during

maintenance

Atofina, Paris-le-Défense, France. Atofina Paris La Défense Cedex

27.12.2002

Source

1.11 ADDITIONAL REMARKS

1.12 LAST LITERATURE SEARCH

Type of search

: Internal and External

Chapters covered Date of search : 3, 4, 5 : 27.12.2002

Date of Search

: Atofina, Paris-le-Défense, France. Atofina Paris La Défense Cedex

Source 27.12.2002

1.13 REVIEWS

ld 67-68-5 Date 17.05.2004

(130)

(124) (133)

MELTING POINT

Value

: = 18.5 °C : no, at °C

Decomposition Sublimation Method

: other: no data

Year

GLP Test substance

: no data

Reliability

(2) valid with restrictions : Critical study for SIDS endpoint

Flag 10.12.2002

(56)

2.2 **BOILING POINT**

Value

: = 189 °C at 1013 hPa

Decomposition Method

other: no data

Year

GLP Test substance

: no data

Remark

Source

: Start of decomposition : T>190°C

Decomposition products: Methane thiol; formaldehyde;

dimethyl sulfide and dimethylsulfone.

: ATOFINA, Paris-La Défense, France.

Atofina Paris La Défense Cedex Reliability

: (2) valid with restrictions Data from handbook

: Critical study for SIDS endpoint 12.12.2002

DENSITY

Type

: density

Value = 1.1 g/cm3 at 20 °C

Method : other: no data

Year

Source

Test substance

: no data

: ATOFINA, Paris-La Défense, France. Atofina Paris La Défense Cedex

Reliability

: (2) valid with restrictions

12.12.2002

2.3.1 GRANULOMETRY

VAPOUR PRESSURE

Value

: = .55 hPa at 20 °C

Decomposition

ld 67-68-5 Date 17.05.2004

(145)

(72)

(26)(71)

Method

other (calculated): no data

Year

GLP

no data

Test substance Source

Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability

(2) valid with restrictions

Flag

Critical study for SIDS endpoint

12.12.2002

Value : = .81 hPa at 25 °C

Source ATOFINA, Paris-La Défense, France.

Atofina Paris La Défense Cedex

Reliability Flag

(2) valid with restrictions

Critical study for SIDS endpoint 07.05.2004

(38)

PARTITION COEFFICIENT

Partition coefficient

Log pow

= -1.35 at °C

pH value Method

other (measured)

Year

GLP Test substance

no data

Source : ATOFINA, Paris-La Défense, France. Atofina Paris La Défense Cedex

Reliability

(2) valid with restrictions Data from handbook

Flag : Critical study for SIDS endpoint 18.09.2000

2.6.1 SOLUBILITY IN DIFFERENT MEDIA

Solubility in Value

Water at °C

pH value

concentration

at °C

Temperature effects

Examine different pol.

Description

at 25 °C

Stable

Remark

Totally soluble in water.

Source Reliability ATOFINA, Paris-La Défense, France. Atofina Paris La Défense Cedex

(2) valid with restrictions Data from handbook

Flag

12.12.2002

Critical study for SIDS endpoint

Solubility in

Water

Value = 1000 g/l at °C

ld 67-68-5 Date 17.05.2004

(75)

pH value

at °C concentration

Temperature effects Examine different pol.

рКа Description at 25 °C

Stable

Reliability : (2) valid with restrictions

29.07.2003 (47)

2.6.2 SURFACE TENSION

2.7 **FLASH POINT**

Value : = 87 °C Type : closed cup Method other: no data Year

GLP

: no data

Test substance

Source : ATOFINA, Paris-La Défense, France. Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions

Data from handbook 17.12.2002

AUTO FLAMMABILITY 2.8

Value : 300 - 302 °C at

: ATOFINA, Paris-La Défense, France. Source Atofina Paris La Défense Cedex Reliability

: (2) valid with restrictions

Data from handbook

17.12.2002 (74)

FLAMMABILITY 2.9

Result : flammable

Source : Elf Aquitaine Lacq

Atofina Paris La Défense Cedex

23.10.1995 (128)

2.10 EXPLOSIVE PROPERTIES

Result explosive under influence of a flame

Method other Year

GLP no

Test substance as prescribed by 1.1 - 1.4

ld 67-68-5 Date 17.05.2004

Remark Source

Explosivity limits of vapours : lel:2.6 %; uel:28.5%
 ATOFINA, Paris-La Défense, France.
 Atofina Paris La Défense Cedex

Reliability

: (2) valid with restrictions

Data from handbook

12.12.2002

(131)

2.11 OXIDIZING PROPERTIES

2.12 DISSOCIATION CONSTANT

2.13 VISCOSITY

Value Result

Source

: 2.14 - mPa s (dynamic) at 20 °C

: Atofina, Paris-le-Défense, France.

Atofina Paris La Défense Cedex

Reliability

: (2) valid with restrictions (133)26.02.2003

2.14 ADDITIONAL REMARKS

ld 67-68-5 Date 17.05.2004

3.1.1 PHOTODEGRADATION

air

Light source Light spectrum

Relative intensity based on intensity of sunlight

INDIRECT PHOTOLYSIS

Sensitizer : OH

Conc. of sensitizer 1000000 molecule/cm3

Rate constant = .0000000000062 cm3/(molecule*sec)

Degradation = 50 % after 3 hour(s) Deg. product

Method

other (calculated) Year

GLP Test substance

Source

Remark Preliminary investigations indicated that dimethylsulfone is

the major reaction product (BARNES, I. et al, 1987.In: Phys. Chem. Behavior Atmos. Pollut., 327-37.)

Concentration of OH radicals = 1000000 OH/cm3 (Prin et al, 1995. Atmospheric trends and lifetime of CH3CCl3 and global

OH concentration, vol. 269.) Atofina, Paris-le-Défense, France.

Atofina Paris La Défense Cedex Reliability (1) valid without restriction

03.01.2003 (7)

air Light source

Light spectrum nm

Relative intensity based on intensity of sunlight

Remark : DMSO does not contain any chromophores that absorb UV

radiation above 290 nm and therefore, direct photolysis will

not be significant.

Source Atofina, Paris-le-Défense, France.

Atofina Paris La Défense Cedex

Reliability : (4) not assignable

27.12.2002 (78)

3.1.2 STABILITY IN WATER

Type abiotic t1/2 pH4 at °C t1/2 pH7 at °C t1/2 pH9 at °C

Result : In aqueous solution, oxidation rate by OH radical, at

neutral pH was 0.71E10 I/mol sec, with OH concentration in

water 1E10-17 mol/l.

Source Atofina, Paris-le-Défense, France.

Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions

12.08.2003 (46)

ld 67-68-5 Date 17.05.2004

Type t1/2 pH4 : abiotic : at °C : at °C

at °C

t1/2 pH7 t1/2 pH9

Result

: In aqueous solution, the rate of reaction of DMSO with hydroxy radicals was found to be 7.3E9 cm3/mol sec, which corresponds to a half-life range of 53 h to 732 d, assuming that OH concentration in natural waters is 0.15E-17 to

50E-17

Source

Atofina, Paris-le-Défense, France. Atofina Paris La Défense Cedex

27.12.2002

(110)

Remark

DMSO disproportionates in water to dimethyl sulfide and dimethyl sulfone. The redox reaction is catalyzed by light. DMSO is readily reduced to dimethyl sulfide by reducing

agents such as Sn(II), Ti(III), Cr(II) that may be present in the environment.

Source

Atofina, Paris-le-Défense, France. Atofina Paris La Défense Cedex

Reliability 29.07.2003 (4) not assignable

(78)

3.1.3 STABILITY IN SOIL

Result

The reduction of DMSO to dimethyl sulfide (DMS) in soil was measured in a range of 47 New Zealands topsoils and humus samples.

The relationships between the rate of DMSO reduction and soil type and land use, organic C and N, the soil microbial C estimated by the substrate-induced-respiration (SIR) method, mineralization of N under anaerobic incubation and the mineralization of organic S, were investigated.

DMSO was reduced rapidly in all soils and rates ranged between 180 and 8124 ng DMS/g soil/h, with the coefficient of variance being typically <10%.

Reduction was significantly correlated with the organic carbon and N content of the 44 mineral soils (r=0.61 and 0.62 respectively), anaerobically-mineralized N (r= 0.80), microbial biomass C $\,$ (r=0.81) and aerobically mineralized SO4-S (r=0.60).

The reduction of DMSO was shown to be more sensitive to the presence of Cr(VI) or As(V) than was the SIR response. The sensitivity and reproducibility of the assay make the technique potentially useful for the study of microbial activity in aggregates, rhizospheres samples and

contaminated soils.

Source

Atofina, Paris-le-Défense, France.
 Atofina Paris La Défense Cedex

Reliability 07.02.2003 : (4) not assignable

(140)

ld 67-68-5 Date 17.05.2004

(5)

(4)(73)

3.2.1 MONITORING DATA

Type of measurement

Media

background concentration

Concentration

Result

Method

surface water

DMSO is a common constituent of natural water.

A representative surface sample of seawater from the north

pacific contained 0.49 ppb.

Representative samples from 3 U.S. rivers contained 0.08 to

Atofina, Paris-le-Défense, France.

Atofina Paris La Défense Cedex

Source 07.02.2003

Type of measurement

Media

other: rain

Concentration

Method

Remark Samples of rainwater in US contained 0.14 to 0.19 ppb (ref

Samples of rain from 2 storms in the south Pacific contained

1300-2600 ng/l and 100 ng/l DMSO (ref 2).

Source Atofina, Paris-le-Défense, France.

Atofina Paris La Défense Cedex

07.02.2003

Type of measurement

Media Concentration

Method

air

Remark

Aerosol concentration of DMSO (and other compounds) was measured with land-based stations (principally Plymouth, Devon, UK) and various shipboard stations in the North sea

and North atlantic ocean.

Aerosol samples collected between July, 1985 and July, 1987 were analyzed both in terms of their back trajectories and variation with time DMSO shows seasonal cycles (abstract).

Source

Atofina, Paris-le-Défense, France.

07.02.2003

Atofina Paris La Défense Cedex (154)

3.2.2 FIELD STUDIES

3.3.1 TRANSPORT BETWEEN ENVIRONMENTAL COMPARTMENTS

Type

fugacity model level III

Media

% (Fugacity Model Level I)

Air Water Soil Biota

Soil

% (Fugacity Model Level I) % (Fugacity Model Level I)
% (Fugacity Model Level II/III)
% (Fugacity Model Level II/III)
% (Fugacity Model Level II/III)

id 67-68-5

Date 17.05.2004

(97)

Method Year

:

Result

: Level III Fugacity Model:

*Chem Name : Methane, sulfinylbis-

Molecular Wt: 78.13

Henry's LC: 1.51e-009 atm-m3/mole (Henry database) Vapor Press: 0.622 mm Hg (Mpbpwin program)

Log Kow : -1.35 (Kowwin program) Soil Koc : 0.0183 (calc by model)

Mass Amount Half-Life Emissions

 (percent)
 (hr)
 (kg/hr)

 Air
 0.0458
 4.14
 1000

 Water
 45.9
 360
 1000

 Soil
 53.9
 360
 1000

 Sediment
 0.0766
 1.44e+003
 0

 Fugacity Reaction Advection Reaction (atm) (kg/hr) (kg/hr) (percent)

Air 1.74e-012 93.3 5.57 3.11 Water 5.41e-014 1.08e+003 559 35.9 Soil 2.35e-012 1.26e+003 0 42.1 Sediment 4.5e-014 0.449 0.0186 0.015

Advection (percent) Air 0.186 Water 18.6 Soil 0 Sediment 0.000622

*Persistence Time: 406 hr Reaction Time: 500 hr Advection Time: 2.15e+003 hr Percent Reacted: 81.2 Percent Advected: 18.8

*Half-Lives (hr), (based upon Biowin (Ultimate) and Aopwin):

Air: 4.14 Water: 360 Soil: 360 Sediment: 1440

Biowin estimate: 3.027 (weeks)

*Advection Times (hr): Air: 100 Water: 1000 Sediment: 5e+004

Sediment: 5

: ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

Reliability : (2) valid with restrictions
Flag : Critical study for SIDS endpoint

03.06.2003

Source

Type : adsorption

Media :
Air : % (Fugacity Model Level I)

ld 67-68-5 Date 17.05.2004

(77)

Water % (Fugacity Model Level I) Soil % (Fugacity Model Level I) Biota % (Fugacity Model Level II/III) Soil % (Fugacity Model Level II/III) Method

Year

Remark DMSO adsorbs both chemically and physically to clay

It intercalates between the layers of smectite, kaolinite, montmorillonite, dictite and halloysite.

With smectite, crystallography studies indicate that a double layer of DMSO is intercalated between clay layers.

With kaolin, three hydrogen bonds are formed between the

kaolin hydroxyls and the DMSO oxygen.

Source Atofina, Paris-le-Défense, France.

Atofina Paris La Défense Cedex

29.07.2003 (6) (45) (98) (113)

volatility Media water - air

Air % (Fugacity Model Level I) % (Fugacity Model Level I)
% (Fugacity Model Level I)
% (Fugacity Model Level I) Water Soil Biota % (Fugacity Model Level II/III) Soil % (Fugacity Model Level II/III)

Method Year

Remark The Henry's law constant at 15°C is 7.77E-9 atm cm3/mol.

DMSO will therefore not volatilize from water. Its concentration would increase in time as water

At a wind speed of 1 m/sec, the volatilization rate was approximately 1.2 and 9 μ g/min at -17.5 and 0 degree C, respectively. The calculated volatilization rate at zero wind velocity is 0.8 and 0.07 $\mu g/min$ at 0 and -20 degree C,

respectively.

Source Atofina, Paris-le-Défense, France.

Atofina Paris La Défense Cedex

29.07.2003

3.3.2 DISTRIBUTION

3.4 MODE OF DEGRADATION IN ACTUAL USE

BIODEGRADATION 3.5

Type

Inoculum

Concentration

162 mg/l related to Test substance

related to Contact time

27 day(s)

= 99 (±) % after 27 day(s) Degradation Result readily biodegradable

ld 67-68-5 Date 17.05.2004

(51)

Control substance

Acetic acid, sodium salt

Deg. product Method

Kinetic

ISO 7827 "Evaluation in an aqueous medium of the 'ultimate' aerobic

biodegradability of organic compounds - method by anlaysis of dissolved organic carbon (DOC)*

Year

GLP no data Test substance other TS

Test substance

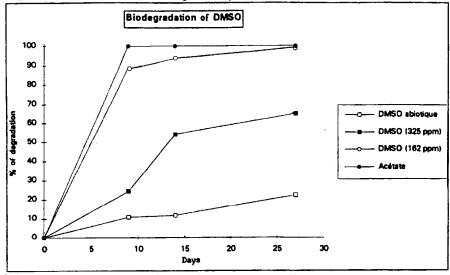
dimethyl sulfoxide (CAS 67-68-5), purity not specified

DMSO (CAS 67-68-5) purity >99%

Result TOC (ppm) day 0 day 9

day 14 day 27 DMSO 325 ppm 111 95 62 50 DMSO 162 ppm 62 15 18 8 DMSO abiotic 112 111 110 98

Attached document dmso biodeg curve.bmp



Source ATOFINA, PARIS-LA-DEFENSE, FRANCE Reliability

(2) valid with restrictions

Guideline study without detailed documentation and with acceptable

restrictions.

Flag Critical study for SIDS endpoint

14.05.2004

Type aerobic

Inoculum activated sludge

Concentration 100 mg/l related to Test substance

related to

Contact time

Degradation = 3.1 (±) % after 14 day(s)

Result

Deg. product

Method OECD Guide-line 301 C "Ready Biodegradability: Modified MITI Test (I)"

Year 1981 GLP no data Test substance no data

ld 67-68-5 Date 17.05.2004

(13)

: ATOFINA, PARIS-LA-DEFENSE, FRANCE. Source

Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions Critical study for SIDS endpoint Flag

03.06.2003

: aerobic Type

Inoculum domestic sewage, adapted Concentration 65 mg/l related to Test substance

20 mg/l related to DOC (Dissolved Organic Carbon)

68 day(s) Contact time

Degradation

: = 90.4 (±) % after 32 day(s) : other: Have a good potential of biodegradation but it's recommended to be Result

careful about its concentration in effluent because toxic effects can appear

Kinetic of testsubst. 14 day(s) = 18 % 25 day(s) = 79.5 %

28 day(s) = 87 % 32 day(s) = 98 % 68 day(s) = 54 %

Control substance

Kinetic 32 day(s) = 92.6 %

68 day(s) = 92.1 % Deg. product not measured

OECD Guide-line 303 A "Simulation Test - Aerobic Sewage Treatment: Method Coupled Unit Test"

1981 Year GLP no

Test substance as prescribed by 1.1 - 1.4

INOCULUM/TEST ORGANISM- Type of sludge: Domestic sewage, **Test condition**

- Sampling site: Versailles.

- Preparation of inoculum: Not available. - Initial cell concentration: 3.0 g/l.

TEST SYSTEM

- Culturing apparatus: 2 litre reactor.

- Aeration device: Bubbling ungreasy compressed air.

- Measuring equipment: DOHRMANN DC 80, DIONEX 4010 I, PERKIN

ELMER Lambda 5.

INITIAL TEST SUBSTANCE CONCENTRATION:

2 concentrations:

1- 65 mg/l.

2- 130 mg/l.

METHOD OF PREPARATION OF TEST SOLUTION: The substance was directly introduced in the test solution afetr 13 days of the beginning of the

DURATION OF THE TEST: 68 days with two phases:

1- From the 13th days until the 33th days.

2- from the 34th days until the 67th days.

ANALYTICAL PARAMETER: The evaluation of the biodegradation was done by measuring the difference of COD level between the effluent issued

from the 2 following units:

1- Activated sludge.

2- Activated sludge+DMSO.

SAMPLING: Every day until the end of the test (68 days).

ld 67-68-5 Date 17.05.2004

TEST CONDITIONS- Composition of medium: For one litre:
1- 10 ml of nutritive solution:
16g peptone
11g meat extract
3g urea
0.7g NaCl
0.4g CaCl2,2H2O
0.2g MgSo4,2H2O
q.s.p ultrapure water 1 l.

2- 10 ml of K2HPO4 : 2.8g K2HPO4 q.s.p 1 litre.3- q.s.p 1 liter of city water.

- Additional substrate: No.- pH value: Not available.

INTERMEDIATES / DEGRADATION PRODUCTS: Not identified.

NITRATE/NITRITE MEASUREMENT: Yes.

Conclusion: DMSO has a good potential of biodegradation but it's recommended to be careful about its concentration in effluent because toxic effects can appear.

1- First case: DMSO concentration = 65mg/l DR = 90.4%.

2- second case: DMSO concentration = 130mg/l DR = 65.1% toxic effects can appear.

Attached document

dmso biodeg day 33-68.bmp dmso biodeg days 1-32.bmp

ld 67-68-5 Date 17.05.2004

ATOCHEM Centre d'Application de Levallois

Tableau 1 (p. 2/2)

Biodégradation du Dimethylsulfoxyde

	UNIT	E 6 (temo	omi		(INITE 3 (essa))			
	COD	000	300D		(10D)	((0))	3000	
	entrée	sortje	elimination		entrée	sortie	elimination	
JOUR								9k
33	99,5	8,6	91.4		127,1	10,3	91,9	91,5
34	110,5	8,7	91.3	(40 ppm)	150.0	9,9	92.3	94,0
35	110,5	3.6	92,2	•	150.0	115	92.3	92.8
36	109.3	8.6	92,2		151.4	14,4	90,4	85.5
37	109,3				151.4			ļ
36	109.3				151.4			İ
39	106.7	8.3	92.4		146.4	20,6	36,4	69,3
40	106.7	9,9	90.7		146.4	29,2	80.1	51.0
41	0.801	10,2	90.4		145,€	17,1	88 3	32,8
42	111.0	7,2	93.3		151,5	15,3	89.5	79,8
43	111,0				151,5			
44	114,6	6,6	93 9		154,7	13,4	91.3	83,5
45	114,6				154,7			-
46	103,5	7.9	93.1		151.4	15,4	90.0	81,3
47	103,5	7.3	92,9		151.4	19,8	86,9	68.8
48	106.3	7.3	92,9		157,3	50.0	86.8	68,3
49	106,3	7.3	93,1		157,3	32.3	85.8	62.5
50	116.0	6.5	93,9		161,0	23.0	85.4	8.83
51	116.0				161.0			
52	116.0				161.0			
53	116.0				161.0			
54	84.9	7,0	94,6		129,8	20,3	87.4	66,6
55	84,9	7,8	9.09		129.8	18,5	85.7	73,3
56	85,0	7,8	90.8		134,0	14.8	88.6	82,5
57	85,0	7,2	91,5		124,0	18,2	85.4	72.6
58	109,4	6,1	92,8	1	151,2	28,5	77.0	44 ()
59	109,4			Ì	151,2			
60	109,4				151,2			
61	109,3	7.1	93,2		153.1	36,8	75.7	26.5
62	109,8	8.0	92,7		153,1	28,6	81.3	48.5
63	119.0	8.2	92,5		170,2	29,0	81,1	45.0
64	119.0	80	93.2		170,2	27,4	63.9	51.5
65	120,2	10.1	91,5	ļ	159.0	31,3	81,8	47.0
56	2,081				159,0			
67	120,2			I	[59,0			l
68		9,5	92,1	l		27.9	82,5	\$1.0

ld 67-68-5 Date 17.05.2004

ATOCHEM Centre d'Application de Levaliois

Tableau 1 (p. 1/2)

Biodegradation du Dimethylsulfoxyde

	UNITE 6 (térnoin)				UNITE 3 (essai)		
	COD	COD	#COD	COD		3000	
	entree	sortie	elimination	entré	e sortie	elimination	
OUR							
0	105.7				6,0		
1	107.6	35,5	75,9	- 11	3.0 26.1	77.0	
5	107,6			- 11	3,0		
4 5 6	107,6			- 11	3.0		
4	111.5	13,3	87.6	- 11	6,5 14,6	86.9	
Ō	111.5	14.8			6,5 14,5	87.6	
6	109.5	12,7	87.7	10	9,9 [4,0	88,9	
7	109.5	13.9		10	9,0 14,	86,8	
8	104.4	14.2	87.0	11:	2.4 16,5	84.7	
9	104,4				2,4		
10	104,4				2.4		
- 11	104,9	13.3	87.3		3,8 13,0		
12	104,9	10.1	90,4		3.8 16.0	85,9	
13	108.1	9,5	90,9	opm: 140),4 9.7	91,5	
14	108,1	10,0	90,7).4 26.4	81,2	
15	112,6	9,8	90,9	130	3.1 32.6	76,8	
16	112.6			136			
17	112,6			139			
181	106.3	9.2	8,19	13	0 30.1	77.9	
19	106.3	9.5	91,1	13			
50	104.8	9,6	91.0	127			
81	6.1401	9,4	91.0	127			
22	109.0	9.3	91,1	144		82.6	
23	109,0			144			
34	109,0			140			
25	105,0	8,8	91,9	139			
26	105,0	10,6	89.9	132			
37	105.5	7.5	92,9	130			
38	105.5	7.6	92,8	130			
39	99.3	7.8	92.6	120		92,0	
30	99.3			126			
31	99.3			126			
32	99.5	7.3	92,6	127	',4 9,5	92.5	

Source Reliability Flag 14.05.2004 ATOFINA, PARIS-LA-DEFENSE, FRANCE.
(2) valid with restrictions
Critical study for SIDS endpoint

Type : Aerobic

20 / 157

(53)

ld 67-68-5 Date 17.05.2004

Inoculum	: other: see remark	
Concentration	: 100 µg/l related to	
	related to	
Contact time	: 24 hour(s)	
Degradation	: (±) % after	
Result	: other: see remark	
Deg. product	:	
Method	: other: see remark	
Year	: 1978	
GLP	: no data	
Test substance	: as prescribed by 1.1 - 1.4	
Method	 To 10 ml TYEG medium, sterilized in a 40 ml serum vial, sterile DMSO was added to a final concentration of 100 μg/ml. 	
	After the medium had been inoculated, the vials were sealed with teflon-coated rubber stoppers.	
	For anaerobic incubations, the headspace was gassed with sterile nitrogen. Cultures were incubated at 30°C for 24 h. The headspace was then sampled and analyzed for DMS by GC.	
Remark	: DMSO was reduced to DMS by 15 different microorganisms	
	including prokaryotes, eukaryotes, aerobes and anaerobes. Micoorganisms % DMSO reduced to DMS	
	- Escherichia coli 18.0	
	- Salmonella thyphimurium 10.5	
	- Klebsiella pneumoniae 37.0	
	- Proteus vulgaris 24.0	
	- Providencia alcalifaciens 26.0	
	- Pseudomonas aeruginosa 27.0	
	- Staphylococcus aureus 0.5	
	- Streptococcus faecalis 0.2	
	- Bacillus subtilis 17.0	
	- Bacillus cereus 0.5	
	- Clostridium butyricum 0.1	
	- Arthrobacter sp. 5.0 - Desulfovibrio sp. 1.1	
	- Desulfovibrio sp. 1.1 - Aspergillus niger 3.0	
	- Saccharomyces cerevisiae 4.5	
	Dimethyl sulfone was not reduced by any of the microorganisms.	
	Activity was greater in anaerobically grown cells than in	
	those grown aerobically.	
Source	: Elf Aquitaine Lacq	
Course	ECB - Existing Chemicals Ispra (VA)	
Godiec		
Journe	Maslansky GeoEnvironmental, Inc. Prescott, AZ	
	Maslansky GeoEnvironmental, Inc. Prescott, AZ Atofina Paris La Défense Cedex	
Reliability	Maslansky GeoEnvironmental, Inc. Prescott, AZ	
	Maslansky GeoEnvironmental, Inc. Prescott, AZ Atofina Paris La Défense Cedex	(164)
Reliability 12.08.2003	Maslansky GeoEnvironmental, Inc. Prescott, AZ Atofina Paris La Défense Cedex : (2) valid with restrictions	(164)
Reliability 12.08.2003 Type	Maslansky GeoEnvironmental, Inc. Prescott, AZ Atofina Paris La Défense Cedex : (2) valid with restrictions : aerobic	(164)
Reliability 12.08.2003 Type Inoculum	Maslansky GeoEnvironmental, Inc. Prescott, AZ Atofina Paris La Défense Cedex : (2) valid with restrictions : aerobic : activated sludge, adapted	(164)
Reliability 12.08.2003 Type Inoculum Concentration	Maslansky GeoEnvironmental, Inc. Prescott, AZ Atofina Paris La Défense Cedex : (2) valid with restrictions : aerobic	(164)
Reliability 12.08.2003 Type Inoculum Concentration Contact time	Maslansky GeoEnvironmental, Inc. Prescott, AZ Atofina Paris La Défense Cedex : (2) valid with restrictions : aerobic : activated sludge, adapted : 200 mg/l related to Test substance related to :	(164)
Reliability 12.08.2003 Type Inoculum Concentration Contact time Degradation	Maslansky GeoEnvironmental, Inc. Prescott, AZ Atofina Paris La Défense Cedex : (2) valid with restrictions : aerobic : activated sludge, adapted : 200 mg/l related to Test substance related to : : (±) % after	(164)
Reliability 12.08.2003 Type Inoculum Concentration Contact time Degradation Result	Maslansky GeoEnvironmental, Inc. Prescott, AZ Atofina Paris La Défense Cedex : (2) valid with restrictions : aerobic : activated sludge, adapted : 200 mg/l related to Test substance related to :	(164)
Reliability 12.08.2003 Type Inoculum Concentration Contact time Degradation	Maslansky GeoEnvironmental, Inc. Prescott, AZ Atofina Paris La Défense Cedex : (2) valid with restrictions : aerobic : activated sludge, adapted : 200 mg/l related to Test substance related to : : (±) % after	(164)
Reliability 12.08.2003 Type Inoculum Concentration Contact time Degradation Result Deg. product	Maslansky GeoEnvironmental, Inc. Prescott, AZ Atofina Paris La Défense Cedex : (2) valid with restrictions : aerobic : activated sludge, adapted : 200 mg/l related to Test substance related to : (±) % after : under test conditions no biodegradation observed	(164)

ld 67-68-5 Date 17.05.2004

Year

no data

GLP Test substance

no data

Method

The test apparatus was a 3.0 I cylindrical acrylic container, operated in a sequential batch manner.

The air flow rate and water temperature could be controlled. The water samples were adjusted so that their concentration was 100-200 mg/l in terms of COD(Mn) or about 100 mg/l in

term of concentration of test substance.

The temperature was 25-30°C and pH was adjusted to

neutral.

The operational procedure was as follows: 2.0 l of the sample water was added to 0.5 I of activated sludge and the aeration of the mixed liquor was started. After 23h aeration, and 1 hour of sedimentation, 2.0 l of the supernatant solution was replaced by the sample water

(fill and draw method).

After one or several days of fill and draw operation (depending on the substance) to acclimatize the sludge to the test water, the water in the container was sampled during aeration at 0 hours (the initial concentration) and

24 h later (the final concentration) for analysis. ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

Reliability 12.08.2003

Source

(2) valid with restrictions

(106)

Type

aerobic :

Inoculum

Remark

In batch system, with activated sludge as inoculum,

degradation of DMSO was < 20%

Source

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Atofina Paris La Défense Cedex

Reliability

(4) not assignable 07.02.2003

(162)

BOD5, COD OR BOD5/COD RATIO 3.6

3.7 **BIOACCUMULATION**

Species

Cyprinus carpio (Fish, fresh water)

Exposure period

42 day(s) at 25 °C

Concentration

1 mg/l

< .4

Elimination

no data

Method

OECD Guide-line 305 C "Bioaccumulation: Test for the Degree of

Bioconcentration in Fish*

Year GLP 1981

Test substance

no data no data

Remark

: With 0.1 mg/l DMSO, BCF < 4.

Exposure method: Continuous flow system

ld 67-68-5 Date 17.05.2004

Source

Analytical method: Gas chromatography ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Reliability

27.12.2002

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(2) valid with restrictions

(14)

ADDITIONAL REMARKS 3.8

Remark

: Formaldehyde is produced when DMSO reacts with OH radicals. OH radicals are produced by oxidation of xanthine by

xanthine oxydase.

With 3.3 mM DMSO, there was 10.88+-4.25 mmol/30 min formaldehyde produced. With 33 mM DMSO, there was 12.12+-3.64 mmol/30 min

Source

formaldehyde produced.

: ATOFINA, PARIS-LA-DEFENSE, FRANCE.

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03.06.2003

(91)

ld 67-68-5 Date 17.05.2004

ACUTE/PROLONGED TOXICITY TO FISH

: flow through Type

Species Pimephales promelas (Fish, fresh water)

yes

other

Exposure period 96 hour(s) Unit g/l LC50 = 34 Limit test

Analytical monitoring

Method Year GLP

no data

Test substance other TS: > 99%

Test substance : DMSO purity >99%

Adult fatheads minnows were held at 25°C in flowing water Test condition

with a controlled photoperiod of 16 h light.

They were fed frozen adult brine shrimp (Artemia sp.).

Age: 31 days, , Mean length: 15.8 mm (SD : 3.259) Mean weigh: 0.062 g (SD: 0.0493)

Temperature: 24.9°C Dissolved oxygen: 7.0 mg/l Hardness: 44.3 mg/l CaCO3 Alkalinity: 46.2 mg/l CaCO3 Tank volume: 0.25 l

pH: 7

Result Toxicant stock: 59.1 g/l

Fish were exposed to 0%, 20%, 40%, 60%, 80% and 100% of the

stock solution.

DM	ISO	Concer	ntrations	(g/l)
		_		_

	Α	в с	D	Ε		
Nominal	0 11.4	22.8	34.1	45.5	56.9	
Measured		8.33	13.8	23.6	26.9	55.6
		9.58	21.4	28.5	46.1	56.3
		7.30	22.7	28.3	44.6	56.7
		9.12	23.4	27.8	46.5	60.1
		10.2	20.3	27.8	45.2	54.3

Average 8.91 20.3 27.2 41.9 56.6

% recovery 99.3 (Standard deviation: 7.3) (number of samples: N=8)

Method of chemical analysis: Gas-liquid chromatography

Number of fish: 10

MORTALITIES

C	Α	В	С	D	E
T0			0	0	0
24			0	10	10
48			0	10	10
72			0	10	10

ld 67-68-5 Date 17.05.2004

(69)

(15)

Source

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Reliability Flag

Type

Year

(1) valid without restriction

12.08.2003

Material Safety Dataset, Critical study for SIDS endpoint

other: static (Renewal of test water at 24 hrs) or semi-static (Renewal

of test water at every 8-16hrs) Oryzias latipes (Fish, fresh water)

Species **Exposure period**

48 hour(s) g/l

Unit LC50

Limit test

Analytical monitoring

Method

no data

= 33

other: in accordance with Japanese Industrial Standard (JIS K 0102-1986-

71) titled "Testing methods for industrial wastewater"

GLP no data

Test substance as prescribed by 1.1 - 1.4

Test condition

- Conditions for fish keeping :

At reception, fishes showing abnormal signs were removed.

Then fishes were reared according to flow through system for 2-3 weeks after external disinfection.

- Condition of acclimatation :

Fishes were reared in an acclimatization tank according to flow through system at temperature of 25+-2 °C for about 28

During the period, abnormal fishes were removed.

- Dilution water for the test Origin: underground water

Water temperature, pH and dissolved oxygen were continuously

measured.

Total hardness, evaporated residue, chemical oxygen demand, chloride ion, ammoniacal nitrogen, harmful

substances such as organic phosphorus compounds, cyanide ion

and heavy metal etc.., were analyzed regularly once a six

months.

The quality of the dilution water used for the test was confirmed to meet the ministerial ordinance of the Ministry of Health and Welfare (August 31, 1978) in total hardness

and evaporated residue.

- Test conditions :

Test tank: round glass vessel Volume of test water: 4l/level Temperature of test water: 25+-2°C Number of fish: 10 fishes/level

-The 48h LC50 value was estimated by Doudoroff method or

Probit method

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Reliability

Source

(2) valid with restrictions

Flag 29.07.2003 Critical study for SIDS endpoint

Type Species

Oncorhynchus mykiss (Fish, fresh water)

Id 67-68-5 Date 17.05.2004

(87)

Exposure period 96 hour(s) Unit g/l LC50 = 33 - 37

no

other

Limit test

Analytical monitoring Method Year

1975 **GLP** no data Test substance no data

Method

: Committee on methods for toxicity tests with aquatic organisms.1975.Methods for acute toxicity tests with fish, macroinvertebrates and amphibians.US Environmental Protection Agency, Ecol. Res. Serv., EPA-660/3-75-009.61 pp.

Test condition

- Life stage: 0.7 g - Temperature: 12°C

- Hardness: 40 to 50 mg/l CaCO3 - Alkalinity: 30 to 35 mg/l CaCO3

- pH: 7.2 to 7.5

Test water (dilution water) was reconstituted from deionized water of at least 10E6 ohms resistivity by the addition of the appropriate reagent grade chemicals.

Fish were acclimated to dilution water by gradually changing the water in acclimatation tanks from 100% well water to 100% reconstituted water over a 1 to 3 day period at the

Temperature of test solutions was maintained within +/-1°C. Fingerling fish weighing 0.2 to 1.5 g were tested at each

concentration.

At least 10 organisms were exposed to each concentration in definitive test. At least 6 concentrations were used per

toxicity test.

Source ATOFINA, PARIS-LA-DEFENSE, FRANCE.

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desired testing temperature.

Reliability

(2) valid with restrictions Critical study for SIDS endpoint Flag

03.06.2003

Type static

Species Lepomis macrochirus (Fish, fresh water)

96 hour(s) **Exposure** period Unit g/l

LC50 > 40 Limit test

Analytical monitoring no Method other Year 1975 GLP no data Test substance no data

Method

Committee on methods for toxicity tests with aquatic organisms.1975.Methods for acute toxicity tests with fish, macroinvertebrates and amphibians. US Environmental

Protection Agency, Ecol. Res. Serv., EPA-660/3-75-009.61 pp. - Life stage: 1.0 g

Test condition

- Temperature: 24°C

- Hardness: 40 to 50 mg/l CaCO3 - Alkalinity: 30 to 35 mg/l CaCO3

- pH: 7.2 to 7.5

ld 67-68-5 Date 17.05.2004

(86)

Test water (dilution water) was reconstituted from deionized water of at least 10E6 ohms resistivity by the addition of the appropriate reagent grade chemicals.

Fish were acclimated to dilution water by gradually changing the water in acclimatation tanks from 100% well water to 100% reconstituted water over a 1 to 3 day period at the desired testing temperature.

Temperature of test solutions was maintained within +-1°C. Fingerling fish weighing 0.2 to 1.5 g were tested at each

concentration. At least 10 organisms were exposed to each concentration in definitive test. At least 6 concentrations were used per

toxicity test.

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Atofina Paris La Défense Cedex

Reliability Flag 29.07.2003

Source

(2) valid with restrictions Critical study for SIDS endpoint

Method Year **GLP**

other: calculated

Test substance

Result

ECOSAR Program (v0.99g) Results:

SMILES: O=S(C)C CHEM: Methane, sulfinylbis-

CAS Num: ChemID1: ChemID2: ChemID3:

MOL FOR: C2 H6 O1 S1

MOL WT: 78.13

Log Kow: -1.22 (KowWin estimate)

Melt Pt:

Wat Sol: 6.687E+005 mg/L (calculated)

ECOSAR v0.99g Class(es) Found

Neutral Organics

Predicted

ECOSAR Class Organism Duration End Pt mg/L (ppm)

Neutral Organic SAR

: Fish

14-day LC50 66901.469

(Baseline Toxicity)

Neutral Organics

: Fish

96-hr LC50 61604.953

Neutral Organics **Neutral Organics Neutral Organics**

: Fish : Fish : Fish (SW) 14-day LC50 66901.461 30-day ChV 4722.975 4722.975 96-hr LC50 2974.511

Note: * = asterick designates: Chemical may not be soluble enough to measure this predicted effect.

ld 67-68-5 Date 17.05.2004

Fish and daphnid acute toxicity log Kow cutoff: 5.0 Green algal EC50 toxicity log Kow cutoff: 6.4

Chronic toxicity log Kow cutoff: 8.0 MW cutoff: 1000

Remark

: ECOSAR data not sufficient by itself to assess aquatic toxicity. However, these data are similar to those reported in the literature and support the conclusion that DMSO is of low toxicity to aquatic organisms.

Reliability 07.05.2004 : (4) not assignable

(49)

Type Species

: other: several species

Exposure period

g/l

: static

Limit test Analytical monitoring Method

other: no data

Year

LC50 (g/l)

Brook trout

95% CI

GLP

Test substance

: other TS: 90% in water

Result

: Toxicity of 90% DMSO to nine species of fish at 12°C

48h

- (Salvenilus fontinales)

(50.9-58.3) (42.2-50.1) (33.2-40.2)

24h

54.5 46.0

36.5

96h

- (Salvenilus namaycush) Lake trout 47.8 38.2 37.3

95% CI

(42.3-54.0) (35.4-41.3) (35.2-39.5)

- (Oncorhynchus mykis)

Rainbow trout 53.0 41.7 32.3 (48.6-57-8) (39.3-44.2) (30.2-34.6) 95% CI

- (Cyprinus carpio) Carp 44.0 44.0 41.7

95% CI (39.3-49.3) (39.3-49.3) (36.3-48.0)

- (Ictalurus melas) Black bullhead 42.5 39.2 36.5

(37.9-47.6) (35.3-43.5) (33.8-39.4) 95% CI

- (Ictalurus punctatus)

Channel catfish 39.0 34.5

95% CI (36.1-42.1) (31.7-37.6) (29.8-35.4)

- (Lepomis cyanellus) Green sunfish 65.0

52.5 43.0 95% CI (61.3-68.9) (47.7-57.8) (35.8-51.6)

- (Lepomis macrochirus)
72 0 56.0 33.5 (63.2-82.1) (51.9-60.5) (29.9-37.5) 95% CI

- (Perca flavascens)

Yellow perch 65.0 57.0

95% CI (61.3-68.9) (52.3-62.1) (33.9-40.3)

Changes in water quality at 12° C had no or little effect

Source

Source

ld 67-68-5 Date 17.05.2004

(156)

upon the toxicity of DMSO

Water quality LC50, 96 h (g/l) Soft 33.5

(30.7-36.5) Medium 32.3

(30.2-34.6) Hard 38.0 (35.0-40.3)

Increases in temperature at medium hardness cause a definitive increase in the toxicity of rainbow trout

°C LC50, 96 h (g/l) 7 41.5 (37.7-45.6) 12 32.3 (30.2-34.6)

17 27.7 (25.0-30.7)

Remark : After preliminary assay carried out with yellow perch,

bioassays were conducted in a 5-gallon glass jars each containing 15 l of reconstituted deionized water and ten fish.

Each test included 5 to 9 concentrations of chemical and 50 to 90 test fish plus 10 fish for control.

Various water qualities were obtained by adding selected concentrations were obtained by adding selected concentrations of reconstituting salts to deionized water:

pH total hardness total alkalinity as ppm CaCO3 as ppm CaCO3 - Soft 6.4-6.8 10-13 10-13

- Soft 6.4-6.8 10-13 10-13 - Medium 7.2-7.6 40-48 30-35 - Hard 7.6-8.0 160-180 110-120

Survival and mortality were recorded at 24, 48 and 96 h.The data were analyzed by plotting concentration versus

mortality on log (probability) graph pa

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Reliability : (2) valid with restrictions
Flag : Critical study for SIDS endpoint

29.07.2003

Remark : The toxic and cryoprotective effects of 10% DMSO

concentration were studied in early stages of loach embryo development (stage 2(8 blastomeres) to stages 27-33

(organogenesis)).

Embryos at stages 22-28 of development were most resistant

to the toxic effects of DMSO.

Maximum cryoprotection was observed at stages 25-33. The number of embryos which did not survive increased with

decreasing temperature from 0 to -10 degree C.

The number of embryos with developmental defects was

greatest at -2 degree C.

At lower temperatures, embryos died rather than developed

abnormally.

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ld 67-68-5 Date 17.05.2004

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Reliability : (3) invalid 07.02.2003

Type

other: see remark Oncorhynchus sp.

Species Exposure period Unit

sure period :

Limit test Analytical monitoring Method

ytical monitoring : no data
od : other
: 1968
: no

GLP : no Test substance : as prescribed by 1.1 - 1.4

Remark

Year

: ACUTE AND CHRONIC TOXICITY

- Intraperitoneal injection studies

The acute toxicity (g DMSO/kg body weight) of DMSO to chinook salmon (Oncorhynchus tshawytscha): LD50 = 12.0, sockeye salmon (O. nerka): LD50 = 13.0, coho salmon (O. kisutch): LD50 = 16.0 and rainbow trout (Salmo gardneri): LD50 = 17.0 was determinated by intraperitoneal injections of aqueous solutions of DMSO.

Fish usually died within 24 h; however, a few died between 24 and 48 hours. Symptoms of toxicity were intermittent spasms and disorientation of swimming shortly before expiring.

In the preliminary chronic study, moderate hematologic changes were observed in fish receiving daily injections of 9.0 - 31 g per kg of body weight (30-100% DMSO) and consisted in slight pycnosis of red blood cell nuclei and moderate degeneration of white blood cells.

Average death rates for this group was: 31 g/kg (100%), 4 h; 12 g/kg (50%), 33 h; 10 g/kg (40%), 90h and 9.0 g/kg (30%), 130 h

No mortality was observed in fish injected with 0.9 g/kg. Histologic alterations noted in fish injected with 31 g/kg were confined to liver, kidney, spleen and pancreas.

- Immersion study

Median tolerance limit (TLm) in yearling coho salmon was:

TLm, 24h = 7.2% TLm, 48h = 5.5% TLm, 72h = 4.9% TLm, 96h = 4.6%

LET50 for coho salmon was determined over a range of 2.0-16.0%.

Groups of fish exposed to DMSO concentrations of 0.01, 0.1 and 2.0% for a period of 100 days; there was no mortality and all groups except fish exposed to 2.0% level gained weight.

Morphologic changes were observed in blood cells from fish exposed to 8, 12 and 16% DMSO and consisted of slight to moderate degeneration of erythrocytes and leukocytes. Tissues of fish subjected to 4% DMSO revealed moderate to severe damage to gills and kidney.

Exposure to 8 and 16% produced similar changes in gills and kidney.

Fish immerged in 16% DMSO revealed marked engorgement and dilation of brain and menigeal blood vessels, engorgement of cerebral capillaries, and cerebral edema.

ld 67-68-5 Date 17.05.2004

(12)

(11)

- Oral studies

No mortality was observed when DMSO (0.01% to 20% v/w) was

fed as a dietary ingredient.

Fish ingesting DMSO displayed a dose-dependent decrease in

diet consumption and body weight gain.

Hematologic differences were noted in fish receiving

15 and 18% DMSO in their diets.

Histopatholic examinations were conducted on fish fed 6, 12,

and 18% DMSO.

Lesions observed in the 12% group were confined to gill lamellae and consisted of moderate edema and moderate to

severe epithelial cell hypertrophy.

Similar, more pronounced changes were observed fish

receiving 18% DMSO.

The injection, immersion and ingestion experiments showed a safe level for yearling coho salmon of 2.8 g of DMSO /kg body weight /day for 28 days, 1% v/v per 100 days and 1%

v/w for 16 weeks, respectively.

These fish would tolerate higher concentrations of DMSO for a shorter period of time and recover rapidly when removed

from DMSO.

Source ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

Reliability

(3) invalid 12.08.2003

> other Oncorhynchus mykiss (Fish, fresh water)

Type Species

Exposure period 96 hour(s)

Unit g/l LC0 = 30.8 LC50 = 38

Limit test

Analytical monitoring

no data Method other: no data

Year GLP

no data Test substance no data

Test condition

Temperature: 12°C Life stage: fingerling

Remark Source

ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex : (4) not assignable

Reliability

27.12.2002

ACUTE TOXICITY TO AQUATIC INVERTEBRATES

Type

Species Exposure period Daphnia magna (Crustacea) 48 hour(s)

g/l

Analytical monitoring

Method

yes

OECD Guide-line 202

Year

no data

GLP **Test substance**

other TS

ld 67-68-5 Date 17.05.2004

(44)

Test substance

dimethyl sulfoxide, Merk, purity >99%

TEST ORGANISMS-Test condition

Strain: Daphnia magna Straus

- Age: Less than 24 hours. From a single clone derived from a healthy

- Feeding: Microscopic algae Selenastrum capricomutum.

- Feeding during test: No. - Control group: Yes.

During the test,

Temperature, O2 dissolved, pH total hardness and conductivity were

monitored weekly.

Ten animals were used per treatment and control.

ASTM hard water (ASTM, 1994), enriched with the organic additive Marinure "25", an extract from the algae Ascophyllum nododum.

Total hardness: 160-180 mg/l CaCO3

pH range : 7.5-8.0

Conductivity of 580 µScm-1

Result The EC50-48h was calculated to be 24.6 mg/l with 95 % confident interval

ranging from 19.1 to 31.7 g/l.

Reliability (2) valid with restrictions Published in peer reviewed journal

Published in peer reviewed journal

Critical study for SIDS endpoint Flag

06.05.2004 (8)

Type

Species Daphnia sp. (Crustacea)

Exposure period 24 hour(s) Unit mg/l EC50 = 7000

Analytical monitoring

Method ISO 6341 15 "Water quality - Determination of the inhibition of the mobility

of Daphnia magna Straus (Cladocera, Crustacea)*

Year

GLP no **Test substance** no data

Remark : Control performed on potassium bichromate (EC50, 24h = 1.2

ATÓFINA, PARIS-LA-DEFENSE, FRANCE. Source

Atofina Paris La Défense Cedex

Reliability (4) not assignable

Critical study for SIDS endpoint Flag

07.05.2004 (127)

Type

Species Daphnia magna (Crustacea)

Exposure period

Unit g/l EC50 = 58.2

ATOFINA, PARIS-LA-DEFENSE, FRANCE. Source

Atofina Paris La Défense Cedex

Reliability

(4) not assignable 27.12.2002

Type other: calculated

Species

ld 67-68-5 Date 17.05.2004

Exposure period Unit

:

Result

ECOSAR Program (v0.99g) Results:

SMILES: O=S(C)C

CHEM : Methane, sulfinylbis-

CAS Num: ChemID1: ChemID2: ChemID3:

MOL FOR: C2 H6 O1 S1

MOL WT: 78.13 Log Kow: -1.22 (KowWin estimate) Melt Pt:

Wat Sol: 6.687E+005 mg/L (calculated)

ECOSAR v0.99g Class(es) Found

Neutral Organics

Predicted

ECOSAR Class

Organism

Duration End Pt mg/L (ppm)

Neutral Organics

: Daphnid

48-hr LC50 52846.434

Neutral Organics

: Daphnid

16-day EC50 662.545

Neutral Organics

: Mysid Shrimp

96-hr LC50 1.77e+005

Note: * = asterick designates: Chemical may not be soluble enough to measure this predicted effect.

Fish and daphnid acute toxicity log Kow cutoff: 5.0 Green algal EC50 toxicity log Kow cutoff: 6.4

Chronic toxicity log Kow cutoff: 8.0

MW cutoff: 1000

Remark

ECOSAR data not sufficient by itself to assess aquatic toxicity. However, these data are similar to those reported in the literature and support the

conclusion that DMSO is of low toxicity to aquatic organisms.

Reliability (4) not assignable

07.05.2004

Type

Daphnia magna (Crustacea)

Species

Exposure period Unit

24 hour(s) g/l = 19.25

Analytical monitoring

EC50

Method Year GLP

1974 no data

no data

Test substance

no data

Test condition Source

Temperature : 20+-1°C

ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

other: Norme AFNOR T 90-301

Reliability 27.12.2002 (4) not assignable

33 / 157

(58)

(49)

ld 67-68-5 Date 17.05.2004

Type

Species

Artemia salina (Crustacea)

Exposure period

24 hour(s) mg/l

Unit EC

= 1300 - 13000

Analytical monitoring

no

Remark

: Life stage of tested organism: nauplii I

The assay is based on disturbance of elongation development (relative to controls raised at the same time) from 24 h to 48 h in animals cultured in medium containing a presumptive

teratogen.

A sample of 50 insulted animals is measured and their average length is compared with that of control animals. A statistically significant difference between the length averages is taken as indicative of teratogenesis. It was considered a significant difference to represent 20% or more of the expected growth increment of the controls

from 24 to 48 h.

DMSO presented no teratogenicity in concentrations ranging

from 0.13% to 1.3%.

Source

ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

07.02.2003

Type Species

Artemia salina (Crustacea)

Exposure period

Unit

g/l

EC50 = 68.6

Source

ATOFINA, PARIS-LA-DEFENSE, FRANÇE.

Atofina Paris La Défense Cedex

03.06.2003

(43)

Type

Species

Remark

Artemia salina (Crustacea)

Exposure period

Unit

Brine shrimp was exposed to DMSO at various stages of its

life cycle. Toxicity was not related to developmental stage for DMSO.

(abstract).

Source ATOFINA, PARIS-LA-DEFENSE, FRANCE.

03.06.2003

Atofina Paris La Défense Cedex

Type Species

other aquatic arthropod: Culex pipiens molestus (larvae)

Exposure period

Unit

g/l

EC50

= 23.2

Source : ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

(42)

(9)

(90)

03.06.2003 Type

ld 67-68-5 Date 17.05.2004

(16)

Species : other aquatic arthropod: Culex restuans

Exposure period : 18 hour(s)
Unit : a/l

 Unit
 : g/l

 EC50
 : = 25.9 - 30.7

 Analytical monitoring
 : no

 Method
 : other

Year : 1975 GLP : no Test substance :

Test condition : Method reported by:

STEPHAN, C.E.: Methods for acute toxicity tests with fish, macroinvertebrates and amphibians.U.S. Environmental Protection Agency, Report N° EPA-660/3-75-009, Covallis, OR,1975. Bioassy had a duration of 18 h, which allowed ample time for the

preparation of samples and calculation and reporting of results within a 24 b period

h period.

Source : ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

12.08.2003 (19)

Туре

Species : other aquatic crustacea: Hyalella azteca

Exposure period : 18 hour(s)
Unit : mg/l

EC50 : = 31900 - 58000 Analytical monitoring : no

Method : other
Year : 1975
GLP : no
Test substance :

Test condition : Method reported by:

STEPHAN, C.E.: Methods for acute toxicity tests with fish, macroinvertebrates and amphibians.U.S. Environmental Protection Agency, Report N° EPA-660/3-75-009, Covallis, OR,1975. Bioassy had a duration of 18 h, which allowed ample time for the

preparation of samples and calculation and reporting of results within a 24

h period.

Source : ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

12.08.2003

Туре

other aquatic crustacea: Palaemonetes kadiakensis

Species : other aquation
Exposure period : 18 hour(s)
Unit : mg/l

EC50 : = 22100 - 45000

Analytical monitoring : no Method : other Year : 1975 GLP : no Test substance : ro

Test condition : Method reported by:

STEPHAN, C.E.: Methods for acute toxicity tests with fish, macroinvertebrates and amphibians.U.S. Environmental Protection

Agency, Report N° PA-660/3-75-009, Covallis, OR,1975.
Bioassy had a duration of 18 h, which allowed ample time for the

preparation of samples and calculation and reporting of results within a 24

h period.

Source : ATOFINA, PARIS-LA-DEFENSE, FRANCE.

ld 67-68-5 Date 17.05.2004

(18)

(17)

12.08.2003

Atofina Paris La Défense Cedex

Type

Year

Species

Daphnia pulex (Crustacea) 18 hour(s)

Exposure period Unit

mg/l

EC50 Analytical monitoring = 22300 - 27100

Method

no other 1975 no

GLP Test substance

no data

Test condition

Method reported by:

STEPHAN, C.E.: Methods for acute toxicity tests with fish, macroinvertebrates and amphibians.U.S. Environmental Protection Agency, Report N° EPA-660/3-75-009, Covallis, OR,

Bioassay had a duration of 18 h, which allowed ample time for the preparation of samples and calculation and reporting

of results within a 24 h period.

Temperature: 23+-1°C Number of organisms: 10

Source

ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

Reliability 07.05.2004

(4) not assignable

TOXICITY TO AQUATIC PLANTS E.G. ALGAE 4.3

Species

Chlorella pyrenoidosa (Algae)

Endpoint Exposure period growth rate 14 day(s)

Unit Limit test

Analytical monitoring Method

other

Year GLP

no data Test substance no data

Remark

Flasks were incubated at 25+-1°C and a light intensity of 7

klux on a 12 hour light-dark cycle.

Growth was monitored by following the increase of optical density over time for 10 to 14 days (precise duration not

specified).

The solvent was assayed at 10 concentrations ranging from

0.1% to 4.0% (v/v) or 1 to 40 g/l.

EC50 value (% v/v), calculated using linear regression analysis (percent inhibition versus solvent concentration),

was 2.01 (95% CI : 1.76-2.26).

Source

ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

Reliability

(2) valid with restrictions

ld 67-68-5 4. Ecotoxicity Date 17.05.2004 Flag : Material Safety Dataset 03.06.2003 (141)Method other:calculated Year GLP Test substance ECOSAR Program (v0.99g) Results: Result SMILES : O=S(C)C CHEM: Methane, sulfinylbis-CAS Num: ChemID1: ChemID2: ChemID3: MOL FOR: C2 H6 O1 S1 MOL WT: 78.13 Log Kow: -1.22 (KowWin estimate) Wat Sol: 6.687E+005 mg/L (calculated) ECOSAR v0.99g Class(es) Found **Neutral Organics** Predicted **ECOSAR Class** Organism Duration End Pt mg/L (ppm) **Neutral Organics** : Green Algae 96-hr EC50 27448.309 Neutral Organics : Green Algae 96-hr ChV 426.870 Note: * = asterick designates: Chemical may not be soluble enough to measure this predicted effect. Fish and daphnid acute toxicity log Kow cutoff: 5.0 Green algal EC50 toxicity log Kow cutoff: 6.4 Chronic toxicity log Kow cutoff: 8.0 MW cutoff: 1000 Remark ECOSAR data not sufficient by itself to assess aquatic toxicity. However, these data are similar to those reported in the literature and support the conclusion that DMSO is of low toxicity to aquatic organisms. Reliability (4) not assignable 07.05.2004 (49)**Species** other algae: Anabaena sp. and Nostoc sp. Endpoint growth rate Exposure period Unit mg/l Limit test **Analytical monitoring** no Method other Year

37 / 157

Five species of blue-green algae were used as test cultures

no data

no data

GLP

Remark

Test substance

ld 67-68-5 Date 17.05.2004

- Anabaena sp.
- Anabaena cylindrica .
- Anabaena variabilis
- Anabaena inaequalis
- Nostoc sp.

Flasks were incubated at 25+-1°C and a light intensity of 7 klux on a 12 hour light-dark cycle.

Growth was monitored by following the increase of optical density over time for 10 to 14 days.

The solvent was assayed at 10 concentrations ranging from 0.1% to 6.0% (1 to 60 g/l).

EC50 value was calculated using linear regression analysis (percent inhibition versus solvent concentration).

EC50 %(v/v) 95% CI - Anabaena variabilis: 3.57 (2.32-4.82)inaequalis: 1.71 (1.24-2.18) " cylindrica : 0.84 - Anabaena sp. (0.12 - 0.66)- Nostoc sp. : 4.02 (3.64-4.40) ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

(2) valid with restrictions

Material Safety Dataset

(142)

Species

Source

Flag 03.06.2003

Reliability

Endpoint Exposure period Unit

EC0 EC12 = 10 Limit test

Analytical monitoring

Method Year GLP

Test substance

Dunaliella bioculata (Algae)

growth rate 48 hour(s) g/l = .5

other

no

Test condition

Bacteria-free algae were maintained on agar plates.

200 ml medium were inoculated with D. bioculata from agar plates and incubated at 24°C under continuous light. Air containing 5% CO2 was bubbled through. At an optical density of 0.6 at 600 nm (about 4 days), 60 - 100 ml of the algae could be transferred to 600 ml of the main culture.

The main culture was diluted to an optical density of 0.6+-0.05 (at 600 nm) with fresh medium, when it was in the logarithmic phase (optical density below 2.0).

Shaking flasks were used in screening assays.

19 ml of a culture with an optical density of 0.6+-0.05 and 1 ml of a solution containing a chemical were shaken at 120 rpm at 24°C under continuous light.

ld 67-68-5 Date 17.05.2004

(57)

Effects of chemical on growth are compared with the control, where the optical density increased from 0.6 to 0.85 within

48 h.

Remark EC4, 48h = 1 g/l (96% growth)

This indicates that there is 4% of growth inhibition.

Source ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

Reliability (3) invalid 10.02.2003

Species other algae: Chiamydomonas eugametos

Endpoint growth rate **Exposure period** 48 hour(s) Unit g/l

EC63 = 25 Limit test

Analytical monitoring no Method other Year GI P

Test substance no data

Test condition Temperature: 25°C

Remark Effect concentration :calculated by Aquire staff based on

data in paper.

Source ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

Reliability (3) invalid

27.12.2002

(76)

Species

Endpoint growth rate Exposure period

Unit

Limit test **Analytical monitoring** no Method other

Year GLP no Test substance no data

Flasks were rotated at 130 rpm on a rotary shaker at 24°C Method

under fluorescent lighting of 310 fc.

Incubation period varied from 2 to 10 days.

Growth was determined in a Bausch and Lomb Spectronic 20

spectrophotometer at a % transmittance of 450 nm.

Type of water not reported.

Remark -Coelastrum microporum Naeg. (algae)

-Bracteacoccus cinnibarinus -Anacystis nidulans (bacteria) -Serratia marcescens (bacteia)

ATOFINA, PARIS-LA-DEFENSE, FRANCE. Source

Atofina Paris La Défense Cedex

Reliability

(4) not assignable 07.02.2003

Species : Skeletonema costatum (Algae)

ld 67-68-5 Date 17.05.2004

(34)

Endpoint growth rate Exposure period 96 hour(s) Unit mg/l

EC50 = 12350 - 25500

Limit test **Analytical monitoring** no data Method other

GLP no Test substance no data

Remark

Saltwater algae. ATOFINA, PARIS-LA-DEFENSE, FRANCE. Source

Atofina Paris La Défense Cedex Reliability (4) not assignable

07.05.2004

TOXICITY TO MICROORGANISMS E.G. BACTERIA

Type aquatic

Species Photobacterium phosphoreum (Bacteria)

Exposure period 5 minute(s) Unit mg/l EC50 **Analytical monitoring** no Method other

Year GLP no Test substance

Remark Test performed at 15°C with a photometer MICROTOX.

The EC50 value was compared with EC50, 24h to Daphnia magna,

a test performed by : IRCHA, les produits chimiques dans l'environnement, Paris, 1981. It was found for Daphnia:

Microtox test was more sensible than acute toxicity test to

daphnia.

ATOFINA, PARIS-LA-DEFENSE, FRANCE. Source

Atofina Paris La Défense Cedex

07.02.2003 (59)

Type aquatic

Species Tetrahymena pyriformis (Protozoa)

Exposure period 24 hour(s) Unit g/l EC50 = 32 **Analytical monitoring** no

Method other Year GLP no data Test substance

Test condition The method was carried out under sterile conditions

Tetrahymena pyriformis was precultured at 30°C for 24h. Two different counting methods were used. One used a

microscope, the other a Coulter counter.

ATOFINA, PARIS-LA-DEFENSE, FRANCE, Source

Atofina Paris La Défense Cedex

07.02.2003 (161)

ld 67-68-5 Date 17.05.2004

Type

: aquatic

Species

other protozoa: Vorticella nebulifera

Exposure period Unit 12 hour(s) g/l

Unit

= 35

Remark

Immersion of healthy populations of V. nebulifera resulted in lethal damage to the outermost pellicular membrane and electron microscopy revealed electron-donor in the

pellicle.

similar exposures to 10-32.5 g/l DMSO were not fatal if the organisms were washed with isotonic salt solution at the

end of the immersion period.

Treatment with these lower concentrations did not interfere

with subsequent reproduction.

RANGANATHAN, V.S., 1976. Effects of dimethyl sulfoxide on the

pellicle of the peritrich ciliate Vorticella

nebulifera.

Trans. Am. Microsc. Soc., 95(3), 394-399. ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

Source 07.02.2003

4.5.1 CHRONIC TOXICITY TO FISH

4.5.2 CHRONIC TOXICITY TO AQUATIC INVERTEBRATES

4.6.1 TOXICITY TO SEDIMENT DWELLING ORGANISMS

4.6.2 TOXICITY TO TERRESTRIAL PLANTS

4.6.3 TOXICITY TO SOIL DWELLING ORGANISMS

4.6.4 TOX. TO OTHER NON MAMM. TERR. SPECIES

Species

other avian

Endpoint

mortality 18 hour(s)

Exposure period Unit

mg/kg bw

LD50

= 100

Test condition

: Method: wild-trapped birds were preconditionned to captivity for 2 to 6 weeks and were usually dosed by gavage with solutions or suspensions of the test chemical in propylene glycol, according to methods described by DeCino et al (1966), Schafer (1972) and Schafer et al (1967).

LD50 values were calculated by the method of Thompson (1948), Thompson and Weil (1952) and Weil (1952). Repellency tests were conducted by the methods of starr et

Remark

ld 67-68-5 Date 17.05.2004

(132)

(148)

al (1964) and Schafer and Brunton (1971), and R50's were calculated either by the method of Litchfield and Wilconxin

(1949) or Thompson and Weil (1952).

Bird species: Agelaius phoeniceus (Red-winged blackbird).

Repellency value (R50)= 1.0 %

Repellency-toxicity index (hazard factor)=0.769, indicating that DMSO has a possible potential for acute oral poisoning.

In the same studies, for starling (Sturnus vulgaris),

LD50= 100 mg/kg. Source

ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

22.07.2003

BIOLOGICAL EFFECTS MONITORING

BIOTRANSFORMATION AND KINETICS

aguatic

Deg. product

Remark

The toxic and cryoprotective effects of 10% DMSO concentration were studied in early stages of loach embryo development (stage 2(8 blastomeres) to stages 27-33 (organogenesis)). Embryos at stages 22-28 of development were

most resistant to the toxic effects of DMSO.

Maximum cryoprotection was observed at stages 25-33. The number of embryos which did not survive increased with decreasing temperature from 0 to -10 degree C.The number of embryos with developmental defects was greatest at -2 degree C.At lower temperatures, embryos died rather than developed

abnormally.

Source

ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

07.02.2003

Type Deg. product aquatic

Remark

Intact developing embryos of the zebrafish Brachydanio rerio were exposed to (14C)DMSO (1 M in fish ringer's solution) to assess the degree of permeation of this

cryoprotectant.

DMSO entered the embryo, reaching only

approx. 2.5% of the expected equilibrium level after 2h

at room temperature.

To identify the barrier to permeation, dechorionated embryos were similarly exposed to isotopic DMSQ.Permeation increased several fold, indicating that the chorion

retards the free exchange of solute.

Embryos were unaffected by exposure to 1M DMSO at 23 degree

The number of embryos hatching after 1h exposure to DMSO at varying concentrations was decreased at 1.5 and 2 M.

ld 67-68-5 Date 17.05.2004

Source

: ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

07.02.2003

(121)

4.9 ADDITIONAL REMARKS

Remark

Species: amphibians

Newts were collected during the summer and spring, acclimated for 10 days at 10°C, weighed, treated with DMSO and maintained in an unfed state which simulated winter conditions.

Intraperitoneal injections of 15 g DMSO/kg or immersion in 2% DMSO were determined to be lethal threshold doses for newts. Death was rapid.

In addition to a transient increase in the body weight of the DMSO-treated animals, other changes were observed:

- a reduction in respiration to 70% that of the controls

- an increase in the weights of spleen (by day 1, the average spleen weight was over twice that of the controls), liver (33% increase after 24 h), and

abdominal fat (150% increase after 20 days).

Lethality was attributed primarily to irreversible changes in the permeability of the skin, causing a water imbalance, hyperhydration and death. The

reduced respiratory rate increased the mortality rate.

Source

: ATOFINA, PARIS-LA-DEFENSE, FRANCE.

Atofina Paris La Défense Cedex

03.06.2003

(93)

ld 67-68-5 Date 17.05.2004

gavage

5.0 TOXICOKINETICS, METABOLISM AND DISTRIBUTION

In Vitro/in vivo In vivo Toxicokinetics Type **Species** monkey

Number of animals

Males **Females**

Doses

Males Females

Vehicle water

Route of administration

Exposure time

Product type guidance

Decision on results on acute tox. tests Adverse effects on prolonged exposure

Half-lives

2nd. 3rd:

Toxic behaviour Deg. product

Method other Year GLP

Test substance as prescribed by 1.1 - 1.4

Method

The absorption and excretion of DMSO were studied in Rhesus monkeys during and after 14 days oral administration of 3 g DMSO/kg. DMSO and its major metabolite, dimethylsulfone (DMSO2), were measured in serum, urine and feces by

gas-liquid chromatography.

Result Serum Concentrations of DMSO and DMSO2

To determine its absorption and maximal blood level, DMSO was measured in serum of Rhesus monkeys at 1, 2, 4, 6, 8 and 24 hrs after the initial oral dose. An average peak serum concentration of 2.3 mg/ml was observed after about 4 hrs which declined relatively rapidly to about 0.95 mg/ml after 24 hrs (Fig. 1). The decline in serum DMSO was linear on semilogarithmic coordinates, i.e. a constant fraction was eliminated in each interval time. Its half-life of 16 hrs was found by measuring the time required for a given serum concentration to decline by one-half. Its elimination rate

constant Ke=0.693/16, or about 4% per hr.

With continued daily oral administration, serum DMSO rose slightly from 0.95 to 1.1 mg/ml on day 2 and then reached a steady state concentration of about 0.9 mg/ml after 4 days (Figure 2). The increase of serum DMSO on day 2 was not statistically significant from the steady state

concentration reached on day 4. After oral DMSO was stopped, serum DMSO declined rapidly and was not detected after 72

DMSO2 became detectable in serum after about 2 hrs, rose slowly and reached about 0.18 mg/ml at 24 hrs (Figure 1). With continued DMSO treatment, DMSO2 attained a steady state concentration of 0.34 mpg/ml after 4 days (Figure 2). When

ld 67-68-5

Date 17.05.2004

oral DMSO was stopped after 14 days, the mean DMSO2 serum concentration declined slowly over the next 96 hrs and trace amounts were detected at 120 hrs. The decline in serum DMSO2 was approximately linear on semilogarithmic coordinates. Its half-life was calculated to be about 38 hrs and its elimination rate constant equaled 0.018, or about 2% per hr.

Urine Concentrations of DMSO and DMSO2
The average total urinary excretion of DMSO and DMSO2 in the monkeys is shown in Figure 3. Urinary excretion of DMSO increased rapidly, reached a steady state level of approximately 9 gms/day after 2 days. The increase in DMSO excretion at 5 days reflected an increased urine volume on that day. DMSO disappeared rapidly from urine after treatment ended and only trace amounts were detected after 72 hrs. About 128 gms, or 60% of the ingested DMSO was excreted in the urine unchanged.

Urinary excretion of DMSO2 increased slowly and reached a maximum of about 3 gms/day after 5 days of DMSO administration (Figure 3). Excretion remained between 2-3 gms/day during the remainder of oral DMSO. Once DMSO treatment stopped, urinary DMSO2 declined slowly over the next 5 days.

Approximately 33 gms, or about 16% of the ingested DMSO was excreted in urine as DMSO2.

Elimination of DMSO in Feces

All fecal samples were collected and stored at 4°C for several weeks prior to analysis by gas-liquid chromatography. Although freshly collected fecal samples smelled of DMSO, no DMSO or DMSO2 was detected when they were analyzed two weeks later. Because of this, we considered the possibility that gut bacteria had degraded the compounds during storage. In an attempt to determine this, DMSO was added to a 5 ml suspension of 1.89 gms of control monkey feces at a final concentration of 1000 uµ/ml. The mixture was incubated at 37°C and at 1, 2, 4, 6 and 8 hrs small aliquots were taken for analysis. The amount of DMSO in the incubation mixture decreased with time. After 8 hrs none could be detected. DMSO2 was not detected at any time during the incubations. The slope of the line representing the loss of DMSO with time was calculated to be 0.64 for DMSO degraded per hr. Since the incubation mixture contained 1.89 gms of feces, approximately 0.34 mg of DMSO was degraded per hr per gm of feces. Based upon an average excretion of about 60 gms of feces per day, we estimated that gut bacteria may be responsible for degrading 450-500 mg of DMSO per day, or about 7 gms of DMSO over the treatment period. This would represent about 3% of the total DMSO ingested.

Attached document

Layman figure 1.bmp Layman figure 2.bmp Layman figure 3.bmp



ld 67-68-5 Date 17.05.2004

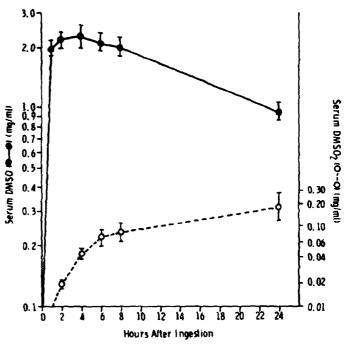


FIG. 1

Serum DMSO and DMSO₂ in Monkeys After a Single Oral Dose of 3 gms DMSO Per kg Body Weight.



ld 67-68-5 Date 17.05.2004

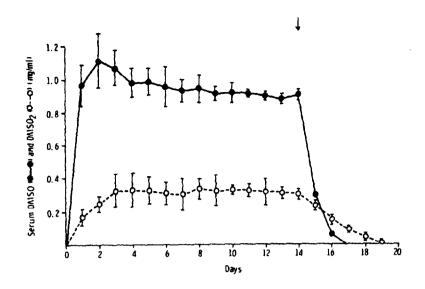


FIG. 2

Serum DMSO and DMSO₂ in Monkeys Given Paily Oral Doses of 3 gms DMSO Per kg Body Weight for 14 Days. Arrow Indicates Last Dose.

ld 67-68-5 Date 17.05.2004

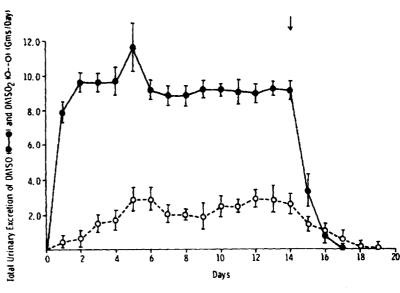


FIG. 3

Urinary Excretion of DMSO and DMSO2 in Monkeys Given Daily 3 gms DMSO Per kg Rody Weight for 14 Days. Arrow Indicates Last Dose.

Source

Atofina, Paris-la-Défense, France. Atofina Paris La Défense Cedex

Reliability 12.08.2003

(2) valid with restrictions

(95)

In Vitro/in vivo

Type

In vivo

Species Number of animals **Toxicokinetics**

Males

Females

Males

Females

Vehicle

Method Year

Doses

GLP

Test substance

as prescribed by 1.1 - 1.4

Method

Absorption, distribution and metabolism of DMSO have been studied in rats, rabbits and guinea pigs using S35-labeled DMSO. Male Sprague-Dawley rats (160-210 g) received 0.55 g/kg (0.5-11 µC) of S35-DMSO orally, dermally or intraperitoneally. The animals were sacrificed at various times, and as much blood as possible was removed from the heart by intracardiac puncture. Tissues were removed, blotted, weighed and homogenized.

ld 67-68-5

Date 17.05.2004

Result

Male New Zealand white rabbits kept in stocks received 0.55 g/kg (approximately 7 μ C) of S35-DMSO dermally. The animals were bled from an ear vein at various times, killed by cervical dislocation and bled. Tissues were removed and homogenized. Aqueous humor was removed. Urine and feces were collected separately from rats and rabbits and guinea pigs receiving DMSO. Rats were placed in all-glass metabolism cages to collect expired air. Biological samples were analysed by paper, thin layer or gas chromatography. Absorption of DMSO and its half-life in plasma:

Rats received S35-DMSO (0.55 g/kg, approximately $0.5 \,\mu\text{C}$) dermally, orally and intraperitoneally and were killed at various times and the plasma assayed for total radioactivity. There were appreciable concentrations of radioactivity in the plasma 0.5 hr after dosing by all routes. The concentration was highest at 2 hr after dermal administration, at which time the level was nearly the same as the maximum attained at 0.5 to 1 hr after either oral or intraperitoneal administration. The levels thereafter declined, with an average half-life of approximately 6 hr. After 24 hr, the levels had declined to about 5 to 10% of the peak concentration.

The rate of passage of dermally applied S35-DMSO through the skin was also estimated in rats and rabbits. Eight rats were dosed dermally with 0.55 g/kg of S35-DMSO. Two animals each were sacrificed after 30 min, 1 hr, 2 hr and 24 hr respectively.

In rats, after 30 min, 63% (average) of the dose remained at the site of application; after 1 hr, 19% (average) of the dose remained; and, after 2 hr, 14% (average) was left. After 24 hr, the radioactivity at the site of application was the same as that of the surrounding skin. Six rabbits were similarly treated. In 2 animals, sacrificed after 30 min, 85% (average) of the dose remained at the site of application. After 4 hr, 11% (average) remained, and, after 24 hr, the radioactivity was essentially equal to that of the surrounding skin.

Disposition of S35-DMSO in rat tissues:

S35-DMSO was administered to rats orally and dermally, the animals were killed at various times and the tissues were assayed for total radioactivity (tables 1 and 2). There were appreciable concentrations of radioactivity in all tissues 0.5 hr after an oral dose. Plasma, kidney, spleen, lung, heart and testes appeared to have somewhat higher levels than liver, fat, small intestine, brain, skeletal muscle and red cells.

Concentrations in the testes, brain skeletal muscle and heart increased after 0.5 hr, but remained virtually constant in other tissues. Levels had declined to minimal values in all tissues after 24 hr.

Concentrations of DMSO and DMSO2 in selected tissues were estimated by extraction and thin layer chromatography. The ratio of DMSO2 to DMSO in rats 4 hr after oral administration of S35-DMSO was found to be virtually constant in liver, testes, kidney, spleen, small intestine, heart and plasma, averaging about 6.5% (range of 4.1-10.6% for tissues of 2 rats). The recovery of radioactivity from these tissues was, in all cases, virtually identical with that obtained when S35-DMSO and S35-DMSO2 were added to tissues in amounts equivalent to those present in rats given S35-DMSO. Thus, the major part, at least, of radioactivity

ld 67-68-5 Date 17.05.2004

present in tissues seems to be represented by DMSO. After dermal administration, tissue concentrations of radioactivity were also appreciable after 0.5 hr, but were somewhat lower than after an oral dose. In this case, levels in the plasma, spleen, liver and lungs were higher than the other tissues. Concentrations in the liver, testes, kidney, spleen, brain, lungs, skeletal muscle, heart, plasma and red cells increased after 4 hr to values comparable to those after an oral dose. Levels in the fat and small intestine remained virtually constant. All tissue concentrations had declined to minimal values after 24 hr.

Disposition of S35-DMSO in rabbit tissues: S35-DMSO was administered dermally to rabbits. The animals were killed at various times and the tissues were assayed for total radioactivity (table 3). Concentrations of radioactivity were appreciable after 0.5 hr in all tissues except the lens. The levels were lower than those seen in the rat, except for the testes which were equivalent. Concentrations in all tissues increased sharply after 4 hr to values that were 3 to 60 times higher than at 0.5 hr. Levels in the lungs, heart, plasma, bile; aqueous humor, vitreous humor and cornea were higher than those in other tissues. Concentrations in the fat tended to remain lower than in other tissues. After 24 hr, the level had declined, but remained somewhat higher than corresponding levels in the

Concentrations of DMSO and DMSO2, in selected tissues were also estimated by extraction and thin layer chromatography. The ratio of DMSO2 to DMSO in rabbits 4 hr after dermal administration of S35-DMSO appeared to be slightly higher than in rats, averaging 11.6% (range of 0-18%) for testes, brain plasma, bile, aqueous humor, lens, vitreous humor and the area of skin at the site of application. The ratio in liver and kidney was slightly higher, averaging 21% (range of 14-26%) in 2 animals.

Excretion of radioactivity following S35-DMSO administration:

Rats received S35-DMSO dermally, orally and intraperitoneally. Rabbits and guinea pigs were given S35-DMSO dermally and intraperitoneally respectively. Urine and feces were collected for 24 hr and assayed for total radioactivity (table 4). Approximately 67 % of the dose was excreted in rat urine within 24 hr and 4 to 10% in the feces. Rabbits excreted 30% of the dose in the urine in 24 hr. Guinea pigs excreted an average of 52% of the dose in the urine in 24 hr and 4% of the dose in the feces. An additional 16% of the dose was excreted by guinea pigs in 24 to 48 hr.

Radioactivity in respired air was measured from 2 rats given 0.55 g/kg of S35-DMSO (approximately 8 μ C) dermally. An average of 6.0% of the dose was found in the respired air assayed over a 24-hr period following dosing. Less than 1% of the dose was found in the respired air of a rabbit monitored over a 3-hr period following an intraperitoneal dose of 0.55 g/kg (4 μ C).

Identification of DMSO and DMSO2 in urine The amount of DMSO2 in rat urine was estimated quantitatively. S35-DMSO (0.55 g/kg, 2.5 μ C) was given to each of 3 rats intraperitoneally, and the 24-hr urine was collected and used for determination of DMSO2.

ld 67-68-5 Date 17.05.2004

DMSO2 was found in an amount which represented an average of 12.8% of the administered DMSO. Total radioactivity in the urine represented an average of 75.8% of the dose.

Urine from rabbits and guinea-pigs given 0.55 g/kg of S35-DMSO dermally or intraperitoneally, respectively, was collected for 24 hr. DMSO and DMSO2 were identified in the

urine.

Attached document

Hucker figures 1 and 2.bmp Hucker figure 3.bmp

Hucker figure 4.bmp

	1			Teni Re	directority			
Timbus			1 0	-	25 hr			
	,	,	,					•
Planna	284	585	691	821	452	623	1 81	67
layer	0.65	0.46	9.53	0.65	0.62	0.56	0 94	, .
Tentra	0.35	0.56				0.87	1.06	. 0
Fat	0.51	0.59	0.50			0.43	0 47	a
Kidney	p. 58	0.76	0.70			0 46	1 14	
Spiren	0.86	0.66	. 8 65	0.71		0.67	1.14	. 0
Small intentine	0.45	0.55	0.34	0.48		0.38	1.06	6
Brane	0.47	0.32	0.56			0.70	1.12	0
Lunes	0.71	0.72	0.81	0.75		0.76	1 02	1 0
Skeletal march	0.50	0.55				0.57	1.19	
Heart	0.64	0.71	0.60	0.73			1.12	
Red celle	0.52	0.49	0.56	0 62	0.44	0.54	0.86	

Eight max received 0.50 g/kg (approximately 7 ac) of 59-DMSO orally. Four animals were killed after 1 ft in and 2 such after 4 and 2 ft in Testines were assayed for total radioactivity. Results for plasma, are expressed as microgram equivalents of DMSO per gram of tissue, and the remaining tissue concentions of the concentration of the extra 6 ft in testing 1 ft in terms and the remaining tissue concentration of the concentration of the extra 6 ft in testing 1 ft in terms are finely assayed as a few field of the first field of the first field of the first field of the field of th

				Total Res	lieuctwity			
Tieter	• 3			20.30				
		,		•	,	•	,	•
Plaima	220	302	393	565	417	445	65	70
Liver	0.62	0.78	0.63	0.60	0.71	9.66	0.62	0.5
Testes	0.24	0.43	0.86	0.86	0.89	9.94	0.50	0.8
FAL	0.50	0.40	0.14	0.32	0.57	0.36	9.84	0.5
Kidney	0.59	0.64	0.06	0.74	0.92	0.77	0 80	
Solven	0.76	0.66	0.71	9.76	0.77		0.66	0.0
Small intertine	0.00	0.50	0.46	0.40	0 45	0 42	0.36	. 0
Brain	0.53	0.54	0.72	0.73	0.71	0.76	0.74	0
Larence	0.63	9.75	9.66	D.70	0.92	0 82	0.78	0.
Skeletal poterie	9 83	9.36	0.50	0.74	0.80	9.77	9.72	. 0
Heart	0.68		0.06	0.66	0.87	0.89	0.68	l e
Red cells	0.54	0.56	0.56	0 56	0.61	0.75	0.63	

Eigh, rate received 9.8 at kt (approximately 11 sc) of 89-1MsO dermilly. Two animals were known steer by ht, 4 animals after 4 ht and 2 minuta after 44 ht. Theses were averaged for rotar admonstrately. Remitts for plasma are expressed as microgram entireless of DMSO per gram of tissue, and the remaining tunner concentrations are expressed as the ratio of the trons level to the corresponding plasma concentration.

TABLE 3

Total radioactivity in rabbit tissues after dermal administration of S²⁵-DMSO

	Total Radioactivity											
Tissue	0.	5 hr	4	hr	24 hr							
	1	2	3	4	S	6						
Plasma	93	124	472	530	117	213						
Liver	0.32	0.66	0.52	0.38	0.69	0.63						
Testes	1.81	0.57	0.61	0.55	0.84	0.82						
Fat	0.27	0.12	0.15	0.13	0.20	0.10						
Kidney	0.27	0.56	0.69	0.46	0.89	0.83						
Spleen	1.25	0.62	0.65	0.61	0.90	0.95						
Small in- testine	0.30	0.54	0.65	0.60	0.85	0.89						
Brain	0.40	0.48	0.65	0.61	0.63	0.66						
Lungs	0.66	0.75	0.73	0.58	0.72	0.72						
Skeletal muscle	0.46	0.38	0.63	0.59	0.75	0.74						
Heart	0.76	1.02	0.69	0.63	0.77	0.77						
Red cells	0.22	0.57	0.55	0.59	0.61	0.56						
Bile	0.57	0.06	1.77	0.88	1.40	0.84						
Aqueous humor	0.47	0.56	0.92	0.91	1.06	1.14						
Vitreous humor	0.29	0.43	0.96	0.80	0.88	0.98						
Lens	0.00	0.05	0.22	0.29	0.53	0.54						
Cornea	0.84	1	ì	1	1	l .						
Skin	1.30	1	}	ľ	1.12	į.						

Six rabbits received 0.55 g/kg (approximately 7 µc) of S³⁵-DMSO dermally. Two animals were

TABLE 4

Excretion of total radioactivity after administration of S²⁵-DMSO by various routes to rats and rabbits

Species	Route of		Total Radioactivity						
	Administration		Urine	Feces					
		-	%	-	%				
Rat	Oral	67	(50-77)	10	(1-14)				
Rat	Dermal	66	(64-67)	4	(2-7)				
Rat	Intraperi- toneal	68	(60-73)	4	(1-9)				
Rabbit	Dermal	30	(20-40)	1					
Guinea pig	Intraperi- toneal	52	(49-55)	4	(2-5)				

Three rats received 0.55 g/kg (approximately 0.9 μ c) of S²⁶-DMSO orally, 3 received the same dose intraperitoneally and 3 received 0.55 g/kg (approximately 5 μ c) dermally. Two rabbits received 0.55 g/kg (approximately 7 μ c) of S²⁶-DMSO dermally. Two guinea pigs were given 0.55 g/kg (approximately 5 μ c) of S²⁶-DMSO intraperitoneally. Urine and feces were collected for 24 hr and assayed for total radioactivity. Results are expressed as percentage of the radioactivity administered. Average values are given with the range shown in brackets.

Conclusion

: DMSO is rapidly and well absorbed when administered orally or dermally to rats or rabbits. Plasma levels of radioactivity following a dermal dose of S35-labeled DMSO to rats reach a maximum value after 2 hr, comparable to the level reached approximately 1 hr after either oral or intraperitoneal administration. Approximately 67 % of the administered radioactivity is excreted in the

Id 67-68-5 Date 17.05.2004

urine of rats within 24 hr, somewhat less than this in the rabbit and guinea pig. The rate of dermal absorption is apparently somewhat faster in the rat than in the rabbit. Radioactivity is rapidly and widely distributed in the tissues of rats and rabbits following administration of labelled DMSO. The levels of radioactivity declined to relatively low values after 24 hr in the rat but remained somewhat higher in the rabbit, suggesting possible accumulation in the latter species. Of particular interest is the finding in the rabbit of relatively high concentrations of radioactive material in the various ocular

The tissue levels of DMSO are roughly equivalent in blood, testis, spleen, liver, kidney and brain. About 10 to 15% of administered DMSO is metabolized to DMSO2 by the rat. Both the DMSO2 and unchanged DMSO are excreted chiefly in the urine. Quantitative gas chromatographic analysis (unpublished data, Hucker et al.) of the urine of rats given S35-labeled DMSO indicates that unchanged DMSO and DMSO2 account for all theradioactivity present in the urine. DMSO was also shown converted to the DMSO2 by rabbits and guinea pigs. Excretion of DMS0 is less rapid in the rabbit than in the rat, suggesting that accumulation of both DMSO and DMSO2 may occur in the rabbit. Quantitative gas chromatographic studies (unpublished data, Hucker et al.) show that the urinary excretion pattern of DMSO and DMSO2 in rabbits differs from that in rats. In rabbits given DMSO dermally (0.55 g/kg), excretion of DMSO was complete within 24 hr and represented 30% of the dose. Excretion of DMSO2, however, continued over a period of 5 days and amounted to 23% of the dose. Excretion of DMS0 and DMSO2 in the guinea pig was net as extensively studied, but preliminary results (unpublished) suggest that it resembles the rabbit in this respect. Ratios of DMSO2 to DMSO excreted in 24 hr ranged from 0.2 to 0.5.

Source

Atofina, Paris-la-Défense, France. Atofina Paris La Défense Cedex

Reliability 12.08.2003

(2) valid with restrictions

(79)

5.1.1 ACUTE ORAL TOXICITY

Type

= 28300 mg/kg bw Value

Species

rat

Strain Sex

male/female 30

Number of animals Vehicle

other: none 10, 20, and 40 g/kg

Doses Method

other 1965

Year **GLP**

Test substance

as prescribed by 1.1 - 1.4

Method

Single oral doses of undiluted DMSO were administered by gavage to groups of 5 male and 5 female Carworth CFN rats. Dose levels were 10, 20, and 40 g/kg. Animals were fasted for 16-18 hr prior to DMSO administration. Animals were observed for 14 days following administration

Id 67-68-5 Date 17.05.2004

(159)

(159)

of DMSO. Body weight was not monitored.

Median lethal dose (LD50), 95% confidence limit, and probit slopes were determined by the Cornfield-Mantel modifications of Karber's method.

Result

With one exception, all deaths occurred within the first 24 hours. Lethal doses caused ataxia, myasthenia, decreased motor activity, and bradypnea shortly after administration. Non-lethal doses of DMSO produced decreased motor activity, although polydipsia and polyuria were noted in rats

following doses of 20 g/kg. The LD50 was determined to be

28.3 g/kg.

Atofina, Paris-la-Défense, France Source

Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions

Flag Critical study for SIDS endpoint

06.05.2004

Туре

Value = 17100 - 26900 mg/kg bw

Species mouse

Strain

male/female Number of animals

30 Vehicle other: none 10, 20, and 40 g/kg Doses

Method other Year 1965 GLP

Test substance as prescribed by 1.1 - 1.4

Method Single oral doses of undiluted DMSO was administered by gavage to

> groups of 5 male and 5 female albino mice. Dose levels were 10, 20, and 40 g/kg. Animals were fasted for 16-18 hr prior to DMSO administration. Animals were observed for 14 days following administration of DMSO.

Bodyweight was not monitored.

Median lethal dose (LD50), 95% confidence limit, probit slopes, and maximum tolerated dose (LD0.1) were determined by the Cornfield-Mantel

modification of Karbers method.

Result With one exception, all deaths occurred within the first 24 hours. Death was

preceded by ataxia, myasthenia, decreased motor activity, and bradypnea.

Non-lethal doses of DMSO produced decreased motor activity.

The LD50 was determined to be 21.4 g/kg; the 95% confidence limits were 17.1 - 26.9 g/kg. The LD0.1 was calculated to be 9.42 g/kg. No more detail

available.

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex (2) valid with restrictions

Reliability

Critical study for SIDS endpoint Flag 06.05.2004

Type LD50

Value > 22000 mg/kg bw

Species rat

Strain

Sex male/female

Number of animals

Vehicle no data

Doses

Method other: no data

Year 1963

ld 67-68-5 Date 17.05.2004

(24)

GLP

: no

Test substance as prescribed by 1.1 - 1.4

Result : 20 ml/kg killed one of four animals in a group of male and

also in a group of female rats. Below this dose there were

no deaths.

Atofina, Paris-la-Défense, France Source

Atofina Paris La Défense Cedex

Reliability (3) invalid

Documentation insufficient for assessment

24.12.2002

Reliability

: LD50 Type

= 13400 - 15700 mg/kg bw Value

Species Strain

Sex no data Number of animals 72 Vehicle other

Doses

Method other: no data Year 1969 GLP no

Test substance as prescribed by 1.1 - 1.4

LD50 was calculated 14.5 g/kg. Hyperemia and inflammation of Remark

the eyes were noted in animals receiving 13 g/kg or more

Source Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

(3) invalid

Documentation insufficient for assessment

24.12.2002 (63)

LD50 Type

Value = 19250 - 22000 mg/kg bw

Species mouse Strain

Sex no data Number of animals

Vehicle other: undiluted Doses Method other: no data

Year 1963 GLP no

as prescribed by 1.1 - 1.4 Test substance

This small experiment indicated that the acute oral toxicity Remark

of DMSO to mice was similar to that of rats. 1/4 mice died at a dose of 17.5 ml/kg, and 2/4 died at 20.0 ml/kg.

Atofina, Paris-la-Défense, France Source Atofina Paris La Défense Cedex

Reliability (3) invalid

Documentation insufficient for assessment

24.12.2002 (24)

5.1.2 ACUTE INHALATION TOXICITY

LC0 Type Value > 1.6 mg/l Species rat

ld 67-68-5 Date 17.05.2004

Strain : Sprague-Dawley

Sex : male Number of animals : 8

Vehicle : other: none

Doses : 1.6 mg/l

Exposure time : 4 hour(s)

Method : other: no data

Year : 1969

Year : 19
GLP : no

Test substance : as prescribed by 1.1 - 1.4

Method : Atmosphere generation and monitoring:

A 30-liter exposure chamber was used in the inhalation experiments. Compressed air at a controlled rate was passed through a modified medicinal nebulizer to produce an aerosol of DMSO which was further passed through an impinger prior to entering the chamber. Cascade impactor samples and subsequent chemical analyses of the material deposited on the slides indicated that 98% of the particles in the chamber were less than 8μ in mean diameter, 72% were less than 4μ , and 15% were less than 2μ , and could therefore be considered respirable.

Atmospheric samples were taken from the chamber for chemical analysis throughout all exposures. Chamber air was passed through a midget impinger containing absolute ethanol. An aliquot of this was further diluted with ethanol and the absorbance determined at 220 mg with a spectrophotometer. The concentration was then calculated from a linear curve of absorbance versus concentration.

Three groups of 8 male Sprague-Dawley rats were exposed to an aerosol of 1600 mg DMSO per cubic meter of air for 4 hr. Control rats were exposed to a normal chamber environment.

One groups were sacrificed immediately after exposure, another 24 hr after exposure, and the third group was observed for 2 weeks after exposure before sacrifice.

All animals were examined for signs of toxicity before, during, and after exposure. Blood was obtained prior to and after exposure. Animals were sacrificed and organs examined; and blood and tissues obtained for biochemical analyses and histologic examination. These signs included diarrhea, lacrimation, dyspnea, ataxia, anorexia, and unusual behavior. All animals were allowed food and water ad libitum when not being exposed. Body weight was not monitored. Blood was obtained prior to the first exposure and after the final exposure for hematologic evaluation. Hemoglobin concentration, packed erythrocye volume as expressed by the microhematocrit, and total leukocyte and reticulocyte counts were done on all animals using standard techniques. At the termination of the experiments, all animals were sacrificed with an overdose of a barbiturate. Blood and tissue specimens were obtained for biochemical analysis (Serum urea nitrogen concentration, serum glutamic-oxaloacetic and glutamic-pyruvic transaminases activity, liver lactate concentration, and the activity of alkaline phosphatase in liver tissue), and gross observations of the organs were made. Sections of heart, lung, liver, spleen, and kidney were taken for histologic examination.

Result

There was no mortality and none of the animals displayed outward signs of toxicity during and after exposure to DMSO. After exposure, the hair was damp and slightly yellow, and the animals had a characteristic garlic-like odor.

Organs appeared normal at necropsy. Histopathologic examination revealed areas of hemorrhage in lung sections of

ld 67-68-5

Date 17.05.2004

(63)

control and DMSO treated animals. Focal and diffuse collections of clear pneumocytes were noted within lung alveoli in DMSO treated rats; similar edematous changes were not seen in control animals.

Biochemical and hematological analyses did not reveal any deviations that could be ascribed to DMSO exposure. acute inhalation data.bmp

Attached document

Cross A.—Expend to 1713 a 178 major for 8 hours, successed 20 hours too Group De Gossand to 100 a 19 major for 6 hours, interpret immediately Group C.—Equand to 100 a 19 major for 6 hours, interpret immediately Group C.—Expend to 2015 a 170 major for 6 hours interpret 2 works for Group D.—Expend to 2015 a 170 major for 6 hours interpret appropriately Group E.—Expends to 2015 a 170 major for 60 hours interpret assemblately All bules for expendit to the mean a 1 intended designs of 5 rets.

Source : Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability : (2) valid with restrictions
Flag : Critical study for SIDS endpoint
06.05.2004

Type : LC0

 Type
 : LC0

 Value
 : > 2.9 mg/l

 Species
 : rat

Strain : Sprague-Dawley

Sex : male Number of animals : 8

 Vehicle
 : other: no data

 Doses
 : 2.9 mg/l

 Exposure time
 : 24 hour(s)

 Method
 : other

 Year
 : 1969

 GLP
 : no

Test substance : as prescribed by 1.1 - 1.4

Method

Atmosphere generation and monitoring:A 30-liter exposure chamber was used in the inhalation experiments. Compressed air at a controlled rate was passed through a modified medicinal nebulizer to produce an aerosol of DMSO which was further passed through an impinger prior to entering the chamber. Cascade impactor samples and subsequent chemical analyses of the material deposited on the slides indicated that 98% of the particles in the chamber were less than 8μ in mean diameter, 72% were less than 4μ , and 15% were less than 2μ , and could therefore be considered respirable. Atmospheric samples were taken from the chamber for chemical analysis throughout all exposures. Chamber air was passed through a midget impinger containing absolute ethanol. An aliquot of this was further diluted with ethanol and the absorbance determined at 220 m μ with a spectrophotometer. The concentration was then calculated from a linear curve of absorbance versus concentration.

A group of 8 male Sprague-Dawley rats was exposed to 2900 mg/m3 for 24 hours and sacrificed immediately after exposure. Control rats were exposed to a normal chamber environment.

All animals were examined for signs of toxicity before, during, and after exposure. Blood was obtained prior to and after exposure. Animals were sacrificed and organs examined; and blood and tissues obtained for

ld 67-68-5

Date 17.05.2004

biochemical analyses and histologic examinatio These signs included diarrhea, lacrimation, dyspnea, ataxia, anorexia, and unusual behavior. All animals were allowed food and water ad libitum when not being exposed. Body weigth was not monitored. Blood was obtained prior to the first exposure and after the final exposure for hematologic evaluation. Hemoglobin concentration, packed erythrocye volume as expressed by the microhematocrit, and total leukocyte and reticulocyte counts were done on all animals using standard techniques. At the termination of the experiments, all animals were sacrificed with an overdose of a barbiturate. Blood and tissue specimens were obtained for biochemical analysis (Serum urea nitrogen concentration, serum glutamic-oxaloacetic and glutamic-pyruvic transaminases activity, liver lactate concentration, and the activity of alkaline phosphatase in liver tissue), and gross observations of the organs were made. Sections of heart, lung, liver, spleen, and kidney were taken for histologic examination.

Result

There was no mortality and none of the animals displayed outward signs of toxicity during and after exposure to DMSO. After exposure, the hair was damp and slightly yellow, and the animals had a characteristic garlic-like odor.

Organs appeared normal at necropsy. Histopathologic examination revealed areas of hemorrhage in lung sections of control and DMSO treated animals. Areas of pulmonary edema were seen in some treated animals but not in control

Biochemical and hematological analyses did not reveal any deviations that could be ascribed to DMSO exposure, with the possible exception of elevated serum urea nitrogen in rats exposed to 2900 mg/m3.

Source

Atofina, Paris-le-Défense, France.
 Atofina Paris La Défense Cedex
 (2) valid with restrictions

Reliability Flag

Critical study for SIDS endpoint

06.05.2004

(63)

 Type
 : LC0

 Value
 : > 2 mg/l

 Species
 : rat

Strain Sex

Number of animals : 8
Vehicle : other: none
Doses : 2.0 mg/l
Exposure time : 40 hour(s)
Method : other

: male

Test substance

as prescribed by 1.1 - 1.4

Method

Atmosphere generation and monitoring: A 30-liter exposure chamber was used in the inhalation experiments. Compressed air at a controlled rate was passed through a modified medicinal nebulizer to produce an aerosol of DMSO which was further passed through an impinger prior to entering the chamber. Cascade impactor samples and subsequent chemical analyses of the material deposited on the slides indicated that 98% of the particles in the chamber were less than 8μ in mean diameter, 72% were less than 4μ , and 15% were less than 2μ , and could therefore be considered respirable. Atmospheric samples were taken from the chamber for chemical analysis throughout all exposures. Chamber air was passed through a midget impinger containing absolute ethanol. An aliquot of this was further diluted with ethanol and the absorbance determined at 220 m_{μ}

ld 67-68-5 Date 17.05.2004

(63)

with a spectrophotometer. The concentration was then calculated from a linear curve of absorbance versus concentration.

A group of 8 male Sprague-Dawley rats was exposed to 2000mg/m3 for 40 hours and sacrificed immediately after exposure. Control rats were exposed to a normal chamber environment.

All animals were examined for signs of toxicity before, during, and after exposure. These signs included diarrhea, lacrimation, dyspnea, ataxia, anorexia, and unusual behavior. All animals were allowed food and water ad libitum when not being exposed. Body weigth was not monitored. Blood was obtained prior to the first exposure and after the final exposure for hematologic evaluation. Hemoglobin concentration, packed erythrocye volume as expressed by the microhematocrit, and total leukocyte and reticulocyte counts were done on all animals using standard techniques. At the termination of the experiments, all animals were sacrificed with an overdose of a barbiturate. Blood and tissue specimens were obtained for biochemical analysis (Serum urea nitrogen concentration, serum glutamicoxaloacetic and glutamic-pyruvic transaminases activity, liver lactate concentration, and the activity of alkaline phosphatase in liver tissue), and gross observations of the organs were made. Sections of heart, lung, liver, spleen, and kidney were taken for histologic examination.

Result

There was no mortality and none of the animals displayed outward signs of toxicity during and after exposure to DMSO. After exposure, the hair was damp and slightly yellow, and the animals had a characteristic garlic-like odor.

Organs appeared normal at necropsy. Histopathologic examination revealed areas of hemorrhage in lung sections of control and DMSO treated animals. Focal and diffuse collections of clear pneumocytes were noted within lung alveoli in DMSO treated rats; similar changes were seen in control animals, but less frequently. Areas of pulmonary edema were seen in some exposed animals, similar edematous changes were not noted in control animals.

Biochemical and hematological analyses did not reveal any deviations that could be ascribed to DMSO exposure.

Source

Atofina, Paris-le-Défense, France. Atofina Paris La Défense Cedex

Reliability

(2) valid with restrictions

Flag

Critical study for SIDS endpoint

06.05.2004

5.1.3 ACUTE DERMAL TOXICITY

LD50 Type

Value ca. 40000 mg/kg bw rat

Species

Strain

Sex

male/female

Number of animals

water

Vehicle **Doses**

other: 1968

Method Year

no

GLP Test substance

as prescribed by 1.1 - 1.4

Method

: Groups of 4 to 14 unshaven rats (108-182 g) were immersed in

Result

Source

ld 67-68-5 Date 17.05.2004

a DMSO solution (40, 60, 80 or 100%) until the fur and the skin were thoroughly wetted. The animals were then withdrawn from the solution and allow most of the excess solution to run off. From the weights before and after dipping, it was possible to calculate the amount of DMSO.

There was no immediate response, but within 24 hours 13/14

rats dipped into 100% DMSO were dead. A complete microscopic

examination of tissues revealed no changes.

Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (3) invalid

Significant methodological deficiences

(138)22.07.2003

Type

ca. 50000 mg/kg bw Value

Species mouse

Strain

Sex male/female Number of animals

Vehicle water

Doses Method other Year 1968

GLP Test substance as prescribed by 1.1 - 1.4

Method Groups of 4 to 6 unshaven mice (13-28 g) were immersed in a

DMSO solution (40, 60, 80 or 100%) until the fur and the skin were thoroughly wetted. The animals were then withdrawn from the solution and allow most of the excess solution to run off. From the weights before and after dipping, it was

possible to calculate the amount of DMSO.

There was no immediate response, but within 24 hours all

Result mice dipped into 100% DMSO were dead.

Atofina, Paris-la-Défense, France Source Atofina Paris La Défense Cedex

Reliability (3) invalid

Significant methodological deficiences

24.12.2002 (138)

5.1.4 ACUTE TOXICITY, OTHER ROUTES

Type LD50

Value = 5360 mg/kg bw

Species rat

Strain

Sex male/female Number of animals Vehicle other: none

Doses

Route of admin. iv. 1 minute(s) Exposure time Method other

1965 GLP

Test substance as prescribed by 1.1 - 1.4

Single i.v. injections of undiluted DMSO were administered Method

to groups of 5 male and 5 female Carworth CFN rats. Dose

ld 67-68-5 Date 17.05.2004

(159)

(159)

levels were 2.5, 5.0, and 10 g/kg. Each dose was

administered over a 1-minute interval. Animals were observed

for 14 days following DMSO administration.

Median lethal dose (LD50), 95% confidence limit, probit slopes and maximum tolerated dose (LD0.1) were determined.

Result With one exception, deaths occurred within the first 24

hours. Death was preceded by tremors, myasthenia, dyspnea, and occassionally, convulsions. Non-lethal doses of DMSO produced decreased motor activity and myasthenia. The LD50 was determined to be 5.36 g/kg; the LD0.1 was

calculated to be 2.35 g/kg.

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions 24.12.2002

Type LD50

Value = 5750 mg/kg bw

Species mouse

Strain

Sex male/female Number of animals 30 Vehicle other: none

Doses Route of admin. i.v. **Exposure time** 1 minute(s)

Method other Year 1965 **GLP**

Test substance as prescribed by 1.1 - 1.4

Single i.v. injections of undiluted DMSO were administered Method

to groups of 5 male and 5 female albino mice. Dose levels were 2.5, 5.0, and 10 g/kg. Each dose was administered over a 1-minute interval. Animals were observed for 14 days

following DMSO administration.

Median lethal dose (LD50), 95% confidence limit, probit slopes and maximum tolerated dose (LD0.1) were determined.

Result With one exception, deaths occurred within the first 24

hours. Death was preceded by tremors, myasthenia, dyspnea, and occassionally, convulsions. Non-lethal doses of DMSO produced decreased motor activity and myasthenia The LD50 was determined to be 5.75 g/kg; the LD0.1 was

calculated to be 2.74 g/kg.

Atofina, Paris-la-Défense, France Source

Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions

24.12.2002

Type LD50

Value = 3100 mg/kg bw

Species mouse Strain Sex

no data Number of animals

Vehicle other: Hank's balanced salt solution

Doses Route of admin.

i.v.

Exposure time

Method other : no data 1969

GLP no

ld 67-68-5 Date 17.05.2004

Test substance : as prescribed by 1.1 - 1.4

Range of values = 2700 - 3500 mg/kg. Remark Atofina, Paris-la-Défense, France Source

Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions

24.12.2002 (63)

5.2.1 SKIN IRRITATION

Species rabbit Concentration undiluted Exposure Occlusive Exposure time 24 hour(s)

Number of animals

Vehicle PDII

Result not irritating Classification not irritating Method Draize Test Year 1969

GLP no

Test substance as prescribed by 1.1 - 1.4

Result Essentially no effect was observed when DMSO was applied

topically other than a slight erythema which faded quickly

after removal of the taped patch. Atofina, Paris-la-Défense, France

Source Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions

26.02.2003 (64)

guinea pig Species undiluted Concentration Exposure Occlusive **Exposure time** 4 hour(s) Number of animals

Vehicle PDII

Result slightly irritating Classification not irritating

OECD Guide-line 404 "Acute Dermal Irritation/Corrosion" Method

Year GLP no data

as prescribed by 1.1 - 1.4 Test substance

Atofina, Paris-la-Défense, France Source

Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions

24.12.2002 (143)

Species mouse Concentration undiluted Exposure Open Exposure time Number of animals 5

Vehicle

PDII Result

not irritating Classification not irritating

ld 67-68-5 Date 17.05.2004

(25)

(63)

Method Year

other 1963

GLP

no

Test substance

as prescribed by 1.1 - 1.4

Method

Undiluted DMSO was painted on to the dorsal scapula region

of 5 male AH mice twice a week for 30 weeks.

Result

At the end of the exposure period, no discernable effect on

the skin was observed.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability

22.07.2003

(2) valid with restrictions

5.2.2 EYE IRRITATION

Species

rabbit Concentration undiluted Dose .1 ml Exposure time 24 hour(s)

Comment

Number of animals

Vehicle

Result Classification Method Year

slightly irritating not irritating **Draize Test** 1969

GLP no

Test substance as prescribed by 1.1 - 1.4

Remark

Slight conjunctivitis in the eyes of rabbits were noted at the 24 hr observation period in the animals undergoing

ocular tests. This had disappeared by 48 hrs.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability

(2) valid with restrictions

24.12.2002

Species rabbit Concentration undiluted Dose .1 mi

Exposure time

Comment 6

Number of animals

Vehicle

Result slightly irritating Classification not irritating

Directive 84/449/EEC, B.5 "Acute toxicity (eye irritation)" Method

24 hour(s)

Year GLP no data

Test substance as prescribed by 1.1 - 1.4

Result

DMSO produced slight erythema of the conjunctiva over the first three days of the study, and a low level of key scoring was also recorded for chemosis, iritis and corneal opacity. The degree of eye injury described by these key scores would not result in DMSO being labelled as an eye

irritant according EEC classification.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

ld 67-68-5 5. Toxicity Date 17.05.2004 Reliability : (2) valid with restrictions 24.12.2002 (84) **Species** rabbit Concentration undiluted Dose .1 ml Exposure time 24 hour(s) Comment Number of animals 4 Vehicle Result slightly irritating Classification not irritating Method Draize Test 1977 Year GLP no data as prescribed by 1.1 - 1.4 Test substance Pure DMSO is only slightly irritant and no irritation was Result observed after 1 day. Atofina, Paris-la-Défense, France Source Atofina Paris La Défense Cedex Reliability (2) valid with restrictions 24.12.2002 (37)rabbit Species Concentration undiluted Dose .1 ml Exposure time 24 hour(s) Comment Number of animals 3 Vehicle Result slightly irritating Classification not irritating Method OECD Guide-line 405 "Acute Eye Irritation/Corrosion" Year GLP no data Test substance as prescribed by 1.1 - 1.4 Result DMSO induced a very slight conjunctival irritation which cleared in 3 days. Atofina, Paris-la-Défense, France Source Atofina Paris La Défense Cedex Reliability (2) valid with restrictions 24.12.2002 (101) 5.3 **SENSITIZATION**

Type :	Guinea pig maximization test
--------	------------------------------

Species guinea pig

Concentration Induction undiluted intracutaneous

Induction undiluted open epicutaneous

3rd: Challenge undiluted open epicutaneous 10

Number of animals

Vehicle other: none Result not sensitizing Classification not sensitizing

Method OECD Guide-line 406 "Skin Sensitization"

Year 1994 no data GLP

Test substance as prescribed by 1.1 - 1.4

ld 67-68-5

Date 17.05.2004

Test condition

- : TEST ORGANISMS:
 - Strain: Hartley strain albino
 - Sex: female (nulliparous and non-gravid)
 - Source, age, weight at study initiation: no data
 - preliminary study: no data
 - Number of animals for final study:
 - . Test substance: 10 females per dosed groups (30 groups). Negative controls: yes
 - . Positive control (dinitrochlorobenzene tested at different concentrations): ves

PRELIMINARY TEST: no data

ADMINISTRATION:

- Vehicles:
- . for intradermal injection: none
- . for the 1st topical application (induction): none,
- . for the last topical application (challenge): none.
- Tested concentrations:
- . For the induction: undiluted DMSO was tested by intradermal injection.
- . For challenge: undiluted DMSO was tested by topical application.
- Test procedure: performed in accordance with the original procedure of Magnusson and Kligman with some modifications. 21 days after the initial intradermal injection (induction), 0.1 mL aliquots of various concentrations of test substance were applied on the flank of each animal for challenge.
- Challenge exposure: open patch held in place for 24h
- Rechallenge: no

EXAMINATIONS:

- Dermal response of each challenge site was evaluated 48 h after the challenge application.
- Grading system:
- . Erythema formation
- 0: no erythema
- 1: very slight erythema (barely perceptible)
- 2: well-defined erythema
- 3: moderate to severe erythema
- 4: severe erythema (beet redness) with eschar formation.
- . Edema formation
- 0: no oedema
- 1: very slight edema (barely perceptible)
- 2: slight edema (visible swelling with well-defined edges)
- 3: moderate edema (visible swelling raised more than 1 mm)
- 4: severe edema (visible swelling raised more than 1 mm and extending beyond the area of exposure)

CALCULATED PARAMETERS:

- SR = sensitization rate = number of animals showing a positive reaction / number of animals tested
- MR = mean response = Total erythema and edema scores / Number of animals tested.
- : Atofina, Paris-la-Défense, France
- Atofina Paris La Défense Cedex

: (2) valid with restrictions

(115)

Source Reliability

14.05.2004

.__.

Type Species Concentration

Buehler Test

guinea pig 1st: Induc

1st: Induction 10 % intracutaneous 2nd: Challenge 10 % intracutaneous

3rd:

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(25)

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7 Number of animals Vehicle water Result not sensitizing Classification not sensitizing Method other: no data

Year 1963 GLP

Test substance as prescribed by 1.1 - 1.4

Source Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions

24.12.2002

Type Species Mouse ear swelling test

mouse Concentration Induction undiluted open epicutaneous

Challenge undiluted open epicutaneous

Number of animals

Vehicle other: none Result not sensitizing Classification not sensitizing Method other

Year 1986 GLP no data

as prescribed by 1.1 - 1.4 Test substance

Atofina, Paris-la-Défense, France Source

Atofina Paris La Défense Cedex Reliability (2) valid with restrictions

24.12.2002

Mouse local lymphnode assay Type

Species mouse

1st: Induction 100 % active substance open epicutaneous 2nd: Concentration

3rd:

Number of animals

Vehicle other: none

Result Classification

Method other Year 1993 GLP no data

Test substance as prescribed by 1.1 - 1.4

Result

Exposure of groups of 3 mice to pure DMSO (concentration 20 and 50 % being ineffective) induced a small increase (approximately 2-fold) in LNC proliferation compared with

the water-solution treated group.

Atofina, Paris-la-Défense, France Source Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions

24.12.2002 (82)

REPEATED DOSE TOXICITY

Type **Species** rat

ld 67-68-5 Date 17.05.2004

Sex Strain male/female Sprague-Dawley

Route of admin. Exposure period

inhalation 13 weeks 6 hours a day, 7 days a week

Frequency of treatm. Post exposure period

4 weeks (control and high dose group)

Control group Method

0, 0.310, 0.964, 2.783 mg/l yes, concurrent no treatment

Year

OECD Guide-line 413 "Subchronic Inhalation Toxicity: 90-day Study"

2000 GI P Test substance

yes other TS

Test substance

dimethyl sulfoxide, CAS# 67-68-5,

Source: Sigma Aldrich Batch: 29356-089 Purity: > 99%

Method

EXPOSURE: The study mean analysed chamber concentrations of DMSO were 0.310 mg/l, 0.964 mg/l and 2.783 mg/l for Groups 2 (Low dose), 3 (Intermediate dose) and 4 (High dose) respectively.

CLINICAL INVESTIGATIONS: Throughout the study, all cages were checked in the morning and again at the end of the normal working day for dead or moribund animals. Clinical signs both during exposures and at other times were monitored and recorded.

BODYWEIGHT: Each rat was weighed for allocation to groups and weekly thereafter commencing one week prior to the start of exposures. Bodyweights were recorded before dosing on the day.

FOOD CONSUMPTION: The quantity of food consumed by each cage of Main and Withdrawal group rats was recorded weekly commencing 1 week prior to the start of exposures until the end of the study.

WATER CONSUMPTION: The amount of water consumed by each cage of Main and Withdrawal group rats was recorded daily beginning one week before the start of dosing.

OPHTHALMIC EXAMINATION: All Main and Withdrawal group rats were subjected to ophthalmoscopic examination prior to the start of exposures. rats from the control and High dose group were examined during Week 13.

FUNCTIONAL OBSERVATION BATTERY: A functional observation battery was conducted on rats from all groups. Comprehensive observations were conducted prior to the start of exposure, during Week 12 and during the recovery period. A shortened battery of observations was conducted during each of Weeks 1-11 of the study.

LABORATORY INVESTIGATIONS: Laboratory investigations comprising analysis of haematological and blood chemistry parameters together with urinary analysis (all according to OECD guideline) were conducted during Week 13 in rats form all Main groups.

OESTRUS CYCLE: The oestrus cycle of female rats was monitored. Vaginal smears were prepared daily from all female rats during Weeks 8 and 9 of the study.

MACROSCOPIC EXAMINATION AND ORGAN WEIGHTS: All rats were subjected to a detailed macroscopic examination . The following organs from all Main and Withdrawal animals killed at the scheduled sacrifice were dissected free of fat and weighed:

Adrenals

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Lungs(all lobes and mainstem bronci)
Brain (medulla, cerebellar and cortical sections)
Ovaries
Epididymides*
Spleen@
Heart
Testes
Kidneys
Thymus (where present)@

- * Right epididymis only. The left epididymis was used for sperm count analysis.
- @ Also recorded for Satellite rats.

Bilateral organs were weighed together

SEMINOLOGY: Male rats were subjected to seminological investigations. Immediately following sacrifice, samples from all males were taken for: - Sperm analysis: Sperm samples taken from vas deferens (from left side) from rats from all groups were assessed for motility using a computer assisted sperm analyser (CASA).

 Morphology: A manual assessment of sperm morphology was performed
 Cauda epididymis (from left side): The cauda epididymis was weighed and homogenised and the number of sperm was counted using a computer assisted sperm analyser (CASA).

HISTOPATHOLOGY: Histopathological examinations were performed on all scheduled tissues (marked with *) for Groups 1 and 4, and on tissues from all groups (marked with X). These tissues were embedded in paraffin wax and sections 4 - $5\,\mu m$ thick were cut, processed and stained with haematoxylin and eosin for examination by light microscopy. Sections, approximately $2\,\mu m$, were cut from the testes (transverse sections) and epididymides (longitudinal sections) and stained with PAS-haematoxylin. Adrenals*

Heart*

Liver

Sciatic nerve*

Alimentary tract

Kidneys'

Seminal vesicles*

Oesophagus*

Larynx *

Skeletal muscle*

Stomach (antrum, glandular and non glandular)*

Duodenum*

Skin

Lungs (all lobes and mainstern bronchi)X

Spinal column -Spinal cord (cervical, thoracic and lumbar)*

Lymph nodes (cervical, mesenteric and tracheobronchial)*

Jejunum*

lleum*

Spleen*

Caecum*

Sternum*

Colon*

Mammary gland

Testes* Rectum*

Nasal passages (head for rostral and caudal nasal cavities)*

Thymus (where present)*

Animal identification markThyroids (with parathyroids)*

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Aorta* Optic nerve Tongue Brain (4 levels)* Ovaries' Trachea (including bifurcation)* Epididymides(a) Pancreas* Eyes* Pharynx* Ureter Femur with joint (for bone marrow in situ) Pituitary' Urinary bladder* Prostate Uterus (corpus and cervix)* Gross abnormalitiesX Salivary gland* Vagina(a) Right epididymis only.

The left epididymls was used for sperm count analysis.

X All rats from all groups.

The lymph nodes were identified separately. The remaining head was retained for paranasal sinuses, oral cavity, nasopharynx, middle ear, teeth, evelids, lacrymal gland, Harderian gland and Zymbal's gland

STATISTICAL ANALYSIS: All statistical analyses were carried out separately for males and females. Food consumption was analysed using cage mean values. For all other parameters the analyses were carried out using the individual animal as the experimental unit. Bodyweight data were analysed using weight gains. The following sequence of statistical tests was used for bodyweight, organ weight and clinical pathology data. If the data consist predominantly of one particular value (relative frequency of the mode exceeded 75%), the proportion of animals with values different from the mode was analysed by appropriate methods. Otherwise:Bartlett's test was applied to test for heterogeneity of variance between treatments; where significant (at the 1% level) heterogeneity was found, a logarithmic transformation was tried to see if a more stable variance structure could be obtained. If no significant heterogeneity was detected (or if a satisfactory transformation was found), and more than two groups were being compared, group means were compared using Williams' test for a doserelated response (Williams, 1971-72), or if there was evidence for a nonmonotonic response, Dunnett's test (Dunnett, 1955, 1964). For separate two-group comparisons, a Student's t test was used. If significant heterogeneity of variance was present (and could not be removed by a logarithmic transformation), groups were compared using Shirley's nonparametric test for a dose-related response (Shirley, 1977), or if there was evidence for a non-monotonic response, Dunn's test (Dunn 1964). For separate two-group comparisons, a Wilcoxon rank sum test (Wilcoxon 1945) was used.

Result

CHAMBER ATMOSPHERE CONDITIONS: Chamber analysed

concentration of DMSO

Group target (mg/l) Actual (mg/l) 2 (Low dose) 0.3 0.31 3 (Inter dose) 1.0 0.964 4 (High dose) 3.0 2.793

The analysed concentrations were in agreement with the target concentrations.

MORTALITY: A Group 4 male was sacrificed on humane grounds in Week

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13 due to the condition of the upper incisors and subsequent weight loss.

CLINICAL SIGNS: Treatment-related clinical signs consisted of red staining around the nose, observed post-exposure in a proportion of Group 3 (Intermediate dose) from Week 5. All Group 4 (High dose) rats were observed to have red staining around thenose pre and post exposure from Week 4 of the study period, continuing throughout the exposure period.

BODYWEIGHT: Rats exposed to DMSO gained less weight over the 13 weeks of exposure compared with controls. The trend was reversed during the recovery phase of the study. However, the differences were small and in the absence of any other indications of systemic toxicity probably reflect a low degree of inappetance caused by the mildly irritant nature of the test atmosphere.

FOOD CONSUMPTION: Differences between Control and Treated groups were minimal and of no toxicological importance.

WATER CONSUMPTION: Differences between Control and Treated groups were minimal and of no toxicological importance.

OPHTHALMIC EXAMINATION: There were no treatment-related differences between the groups.

FUNCTIONAL OBSERVATION BATTERY: Treatment with (DMSO) for 13 weeks followed by a four week recovery period was not associated with any behavioural changes that were considered indicative of neurotoxicity.

OESTRUS CYCLE: There were no differences between control and test groups considered to be attributable to exposure to DMSO.

HAEMATOLOGY: There were no differences between control and test groups considered to be attributable to exposure to DMSO.

BLOOD CHEMISTRY: There were no differences between control and test groups considered to be attributable to exposure to DMSO.

URINALYSIS: There were no differences between control and test groups considered to be attributable to exposure to DMSO.

MACROSCOPIC EXAMINATION: There were no findings that were considered to be attributable to exposure to DMSO.

ORGAN WEIGHTS: The lung weights of male rats exposed to DMSO were significantly greater than Control weights however, the difference was small, not dose related and not seen in females. The difference is considered not to be attributable to exposure to DMSO. There were no other differences between the groups that were considered to be attributable to exposure to DMSO.

SEMINOLOGY: There were no differences between control and test groups considered to be attributable to exposure to DMSO.

HISTOPATHOLOGY: Treatment related changes were found in the nasal passages and pharynx of High dose females which were killed after treatment for 90 days. These changes were not found in rats from the Low and Intermediate dose groups killed at this time. Treatment-related changes in the nasal passages of High dose rats comprised lesions in the inferior ventral medial meatus (pseudogland formation in the respiratory epithelium and epithelial hyperplasia with or without inflammation in the squamous epithelium), and an increased degree ofeosinophilic inclusions in the olfactory epithelium. In the pharynx, prominent goblet cells were

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present in the majority of High dose rats. In rats killed after the recovery period, changes were still evident in the nasal passages of High dose male and females and also in the pharynx of the females.

Attached document Microscopic pathology

13 wk patho.bmp

Terminal kill

		Males					Pemales .				
Group		1	2	3	4	1	2	3	4		
Respiratory epithelium – pseudogland											
formation	Slight	0	0	0	76	0	C.	0	9с		
Squamous epithelium - hyperplasia	Minimal	0	0	0	9c	0	0	0	бa		
Squamous epithelium - inflammation	Minimal	0	0	0	4	0	0	0	3		
Olfactory epithelium – eosinophilic	Minimal	6	9	6	2	5	7	7	0		
inclusions	Slight	0	0	0	4	0	0	0	8		
	Moderate	Ō	Ō	0	4	Ō	0	Ō	2		
	Total	6	9	6	10	5	7	7	10a		
Number of nasal passages examined		10	10	10	10	10	10	10	10		

a = p< 0.05, b = p< 0.01, c = p< 0.001 with Fisher's Exact Test

Pharyan. Prominent goblet cells were present in the epithelium of the majority of rats from the high dose group. This change was not found in rats from the low or intermediate dose groups.

		Males				Females				
Croup		1	2	3	4	1	2	3	4	
Prominent goblet cells	Total	0	0	0	8c	0	0	0	9с	
Number of pharmes examined		10	10	10	10	10	10	10	10	

c = p< 0.001 with Fisher's Exact Test

Recovery kill

		Ma	1	Fen	عطم
Croup		1	4	1	4
Respiratory epithelium - pseudogland formation	Slight	0	3	0	10c
Squamous epithelium - hyperplasia	Minimal	0	0	0	7ь
Squamous epithelium - inflammation	Minimal	D	0	0	2
Olfactory epithelium - eosinophilic inclusions	Minimal	10	1	4	1
	Slight	0	7	0	5
	Moderate	0	2	0	4
	Total	10	10	4	10a
Number of nasal passages examined		10	10	10	10

a = p < 0.05, b = p < 0.01, c = p < 0.001 with Fisher's Exact Test

Conclusion : The no adverse effects concentration could be established at 0.964 mg/l for

Source

respiratory tract irritation and 2.783 mg/l for systemic toxicity.
Atofina, Paris-le-Défense, France.

(1) valid without restriction Reliability : Critical study for SIDS endpoint Flag

14.05.2004 (118)

Type

Species : monkey Sex : male/female

other: Macaca mulatta (rhesus) Strain

Route of admin. gavage Exposure period : 18 months Frequency of treatm. daily Post exposure period None

Doses 1 - 3 - 9 ml of 90% DMSO solution/kg (990 - 2970 - 8910 mg/kg)

Control group other: water, 9 ml/kg NOAEL = 2970 mg/kg bw LOAEL = 8910 mg/kg bw

ld 67-68-5 Date 17.05.2004

Method

other: no data

Year

1970 no

GLP Test substance

other TS

Test substance

dimethyl sulfoxide CAS no.: 67-68-5

Purity: Pharmaceutical-grade

Method

Pharmaceutical-grade DMSO was administered as a 90% solution to 4 groups of rhesus monkeys by gastric intubation, 7 days a week for up to 87 weeks. One half the dose was administered in the morning, the remainder was given in late afternoon. Groups of 2 animals of each sex were treated with 1 or 3 ml/kg. Three animals of each sex received 9 ml/kg. Dosages administered were equivalent to 990, 2970, and 8910 mg/kg/day.

Examinations included water consumption, clinical chemistry, hematology, urinalysis, EKG, reflexes, and body weight. All animals were subjected to a detailed necropsy. Organs were weighed and tissues prepared for histological examination.

Doses of 0, 1, 3, and 9 ml/kg of a 90 % (v/v) aqueous solution of pharmaceutical grade DMSO (these dosages were equivalent to 0, 0.99, 2.97, and 8.91 g/kg per day, respectively) were administered to 4 groups of rhesus monkeys by gastric intubation 7 days per week. One half of the indicated dose was given in the morning and the other half in the late afternoon. Control animais in each group were given water, 9.0 ml/kg body weight.

There were 2 females and 1 male in the oral control group, 2 animals of each sex in the groups treated with 1 and 3 ml/kg, and 3 animals of each sex in the groups receiving 9 ml/kg per day of DMSO. One monkey was kept in separate rooms to preclude the possibility of inhaling DMSO or its metabolites from treated animals. Physical signs, behavior, and survival time were recorded daily. The monkeys were given a nutritionally adequate laboratory ration (Purina Monkey Chow) supplemented with fresh fruit daily and water ad libitum.

Examinations included water consumption, electrocardiogram, neurologic (reflexes), heart rate, body weight, blood pressure, body temperature, respiratory rate, and ophthalmologic. Complete blood counts, serum glutamic-pyruvic transaminase (SGPT), serum alkaline phosphatase (SAP), blood urea nitrogen (BUN), blood glucose, 45-minute sulfobromophthalein (BSP) retention, and endogenous creatinine clearance were measured in all animals. Urinalysis consisted of specific gravity, pH, albumin, glucose, occult blood, ketone bodies, and microscopic examination of the sediment. All of these determinations were performed in accordance with standard procedures.

Because of the need to schedule the broad spectrum of clinical determinations in each animal, starting dates for dosing were staggered over a 6-week period. Surviving monkeys were treated for 74-87 weeks.

All animals that died or were sacrificed were submitted for a detailed necropsy. The following organs were weighed: liver, kidneys, heart, brain, gonads, prostate or uterus, adrenals, thyroid, pituitary, and lungs.

Histomorphologic examinations were performed on the

ld 67-68-5

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following hematoxylineosin-stained sections of formalin-fixed tissues: liver, spleen, stomach (including fundus and pyloric regions), small intestine (including duodénum, jejunum, and ileum), large intestine (including colon and cecum), pancreas, kidneys, bladder, adrenals, gonads, thyroid, pituitary, thymus, salivary glands, lymph nodes (including cervical and mesenteric), heart, lungs, femoral bone marrow, skin, skeletal muscle, spinal cord, brain, gallbladder, epididymis, seminal vesicles, prostate, uterus, aorta, larynx, trachea, peripheral nerve, diaphragm, and lacrimal glands. The eyes were fixed in formalin or Zenker's fixative. Bone marrow smears were stained with Wright's stain.

Result

All the results reported in the publication are listed below.

The highest oral dose, 9 ml/kg per day, was not well tolerated by 6 monkeys in this group; 1 died accidently and 5 succumbed as a result of treatment with DMSO. The 1 and 3 ml/kg doses were well tolerated, and the animais were sacrificed with the control monkeys at the end of the study.

The principal physical signs seen in the animals given DMSO orally included ptyalism and emesis. These signs occurred sporadically and did not appear to be related to the dose except in the group receiving 9 ml/kg. Also, they were observed in the control group, although less frequently. Anorexia occurred at high oral doses but was not evident at the 2 lower dose levels. Some monkeys in all treated groups had erythema of the skin.

Monkeys given 1 and 3 ml/kg orally, showed slightly less gain in mean body weight compared to the respective control animals during the study. However, no biologic significance is attached to these differences because of the limited number of animals per group and the wide range of initial weights. Marked losses in body weight occurred in animals given 9 ml/kg DMSO orally. Most severe losses were during the first 6 weeks of study when emesis and anorexia occurred. One monkey lost 1 kg of weight during week 1 and 1.6 kg by week 12. The remaining monkeys generally showed slight gains in body weight after week 6, but all weighed less at death than when the study began.

No DMSO-related changes were found in the treated monkeys during physical examinations. These tests included mean systolic blood pressure, heart rate, respiratory rate, body temperature, 48-hour water consumption, neurological reflexes, and electrocardiograms, performed during weeks 1, 4, 7, 12, 24, 37, 51, and 73 of study. No evidence of refractoriness to Tropicamide mydriasis was seen in any of these monkeys. The typical DMSO lenticular changes described in other species were not visible in any monkey during the course of the experiment.

The mean hematologic results at approximately 6-month intervals are presented for the control and 3 ml/kg groups in Table 1. Table 2 shows the biochemical data for the same groups. No significant différences were found between the DMSO-treated and control monkeys in any of the hematologic or biochemical parameters evaluated. Animals given 1 ml/kg orally responded in a similar manner. Although values for the animals receiving 9 ml/kg orally are not shown, the data agree with those reported for the control and other test groups. At no time were any abnormal hematologic or biochemical results seen in the animais given the high oral dosage of DMSO.

Further, no significant différences were seen in erythrocyte sedimentation

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rate (ESR), BSP retention, creatinine clearance, urinalysis, and absolute or relative organ weights between the treated and control animals.

No significant lesions attributable to DMSO were found upon gross examination at necropsy.

No histologic changes were visible in the lenses of treated animals.

Attached document

Vogin Table 1.bmp Vogin Table 2.bmp

Different (%)										
DMSO (mi/kg)	Room	Week of	Total WBC	-	L	Other	(\$4,000 mm) 140	(%)	Prothrombic tion (no:)	(~30 ³ /mm²
•	Deposi		9.4	42	*	2	129	42	13.3	130
		*	17	23	71	,	12.1	40	12.2	110
		51	16.2	33	*	•	11.0	>>	12.5	130
		73-74	7,0	-	52	•	11.9	-	26.7	130
,	Derent	•	12.1	43	57		13.5	44	14.5	135
		26	4.5	34	-	4	14.3	41	127	145
		51	**	29	64	2	12.4	*	12.3	139
		13-78	4.1	40	59	ï	12.0	41	14.2	150
	Oral		9.7	36	63		14.2	45	143	120
		200	7.1	30		,	13.0		12.4	230
		52	6.7	4)	34	,	11.9		12.6	170
		78	7.5	47	77	- 1	(3.1	43	14.9	130
	One		11.2	47	13		14.9	42	HAZ	120
		×	33	27	77	1	12.9	40	13.0	150
		52	E1	20	45	,	12.5	44	13.0	150
		79	7.2	33		i	13.3	42	14.9	130

TABLE 2 Mean Biochemical Findings in Monkeys Receiving DMSO (90% v/v)

DMSO (ml/kg)	Route	Week of study	BUN (mg/100 ml)	Glucose (mg/100 ml)	SGPT (units)	SAP (units)
0	Dermal	0	16	73	17	11.9
		26	39	87	20	27.1
		51	25	85	39	10.8
		73-78	24	68	25	6.1
9	Dermal	0	14	73	17	12.7
		26	32	83	21	19.1
		51	27	70	35	14.2
		<i>1</i> 3–78	25	72	18	6.4
. 0	Oral	0	16	67	21	12.0
		26 :	23	105	19	18.7
		52 ⁱ	14	72	37	12.5
		78	18	74	19	8.8
3	Oral	0 -	17	104	18	16.3
		26	17	100	29	20.5
		52 ;	20	102	27	10.8
		78	22	90	17	5.2

Source Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions Flag 06.05.2004 Critical study for SIDS endpoint

(150)

Type

Species monkey Sex male/female

Strain other: Macaca mulatta (rhesus)

Route of admin. dermal Exposure period 18 months Frequency of treatm. daily Post exposure period none

Doses

1 - 3 - 9 ml of 90% DMSO solution/kg (990 - 2970 - 8910 mg/kg)

Control group other: water, 9 ml/kg

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NOAEL

: = 8910 mg/kg

Method Year

: other : 1970 : no

GLP Test substance

: as prescribed by 1.1 - 1.4

Method

Doses (0, 1, 3, and 9 ml/kg) of a 90 % (v/v) aqueous solution of pharmaceutical grade DMSO were administered dermally to 4 groups of rhesus monkeys 7 days per week. Topical administration was by direct application to the entire abdominal skin. Animals were restrained in a supine position for 1 hour after drug administration to prevent ingestion of the applied solution. Control animals in each group were given water, 9.0 ml/kg body weight. Dosages administered were equivalent to 990, 2970, and 8910 mg/kg/day.

There were 2 males and 1 female in the control group, 2 animals of each sex in the groups treated with 1 and 3 ml/kg, and 3 animals of each sex in the groups receiving 9 ml/kg per day of DMSO. One monkey was kept in separate rooms to preclude the possibility of inhaling DMSO or its metabolites from treated animals.

Physical signs, behavior, and survival time were recorded daily. The monkeys were given a nutritionally adequate laboratory ration (Purina Monkey Chow) supplemented with fresh fruit daily and water ad libitum. Examinations included water consumption, electrocardiogram, neurologic (reflexes), heart rate, body weight, blood pressure, body temperature, respiratory rate, and ophthalmologic. Complete blood counts, serum glutamic-pyruvic transaminase (SGPT), serum alkaline phosphatase (SAP), blood urea nitrogen (BUN), blood glucose, 45-minute sulfobromophthalein (BSP) retention, and endogenous creatinine clearance were measured in all animals. Urinalysis consisted of specific gravity, pH, albumin, glucose, occult blood, ketone bodies, and microscopic examination of the sediment. All of these determinations were performed in accordance with standard procedures.

Because of the need to schedule the broad spectrum of clinical determinations in each animal, starting dates for dosing were staggered over a 6-week period. Surviving monkeys were treated for 74-87 weeks.

All animals that died or were sacrificed were submitted to a detailed necropsy. The following organs were weighed: liver, kidneys, heart, brain, gonads, prostate or uterus, adrenals, thyroid, pituitary, and lungs.

Histomorphologic examinations were performed on the following hematoxylineosin-stained sections of formalin-fixed tissues: liver, spleen, stomach (including fundus and pyloric regions), small intestine (including duodénum, jejunum, and ileum), large intestine (including colon and cecum), pancreas, kidneys, bladder, adrenals, gonads, thyroid, pituitary, thymus, salivary glands, lymph nodes (including cervical and mesenteric), heart, lungs, femoral bone marrow, skin, skeletal muscle, spinal cord, brain, gallbladder, epididymis, seminal vesicles, prostate, uterus, aorta, larynx, trachea, peripheral nerve, diaphragm,

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Result

and lacrimal glands. The eyes were fixed in formalin or Zenker's fixative. Bone marrow smears were stained with Wright's stain.

Several accidental deaths (self-strangulation) occurred as a result of vigorous attempts to escape by monkeys restrained in a supine position. Two monkeys that died early in the study were replaced. Two in the treated and 2 in the control groups that died or were sacrificed later were not replaced. In no instance was death attributed to the dermal treatment of these animals with DMSO.

All animals treated topically with DMSO exhibited scaling and flaking of the skin in the area of drug application during the initial phases of the study. There were no apparent differences among the various treatment groups. Although several animals had erythema of the skin it did not appear to be related to the dose, and erythema did not occur at regular internals in any animal. No other adverse behavioral or physical signs were seen that could be attributed to topical application of DMSO.

Monkeys given 1-9 ml/kg DMSO topically showed slightly less gain in mean body weight compared to the control animals during the study. However, no biologic significance is attached to these differences because of the limited number of animals per group and the wide range of initial weights.

No DMSO-related changes were found in the treated monkeys during physical examinations. These tests included mean systolic blood pressure, heart rate, respiratory rate, body temperature, 48-hour water consumption, neurological reflexes, and electrocardiograms, performed during weeks 1, 4, 7, 12, 24, 37, 51, and 73 of study. No evidence of refractoriness to Tropicamide mydriasis was seen in any of these monkeys. The typical DMSO lenticular changes described in other species were not visible in any monkey during the course of the experiment. The only ocular abnormality observed was in one animal which had been given 1 ml of DMSO/kg per day dermally for 82 weeks. In the final ocular examination, this animal had a unilateral complete retinal detachment and syneresis of the vitreous humor. There were no biomicroscopically visible changes in the vitreous humor of the remaining animals.

The mean hematologic results at approximately 6-month intervals are presented for the topically treated control and 9 ml/kg groups in Table 1. Table 2 shows the biochemical data for the same groups. No significant differences were found between the DMSO-treated and control monkeys in any of the hematologic or biochemical parameters evaluated. Animals given 1 or 3 ml/kg responded in a similar manner.

Further, no significant differences were seen in erythrocyte sedimentation rate (ESR), BSP retention, creatinine clearance, urinalysis, and absolute or relative organ weights between the treated and control animals.

No significant lesions attributable to DMSO were found upon gross examination at necropsy. Microscopic examination of the tissues showed tuberculosis in a control monkey in the dermally dosed group. This animal was sacrificed in week 41 because of a positive tuberculin reaction. Epidermal

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thickening and focal chronic hyperkeratosis were seen in skin sections of the abdomen from monkeys which had DMSO or water applied. This is often seen in animais treated dermally with aqueous solutions, and it is a result of constant application of fluid to the epidermis. No histologie changes were visible in the lenses of treated animals with the exception of the monkey with retinal detachment. In this animal, there appeared to be several swollen lenticular fibers in the equatorial portion of the lens in the affected eyes. The retina was undergoing cystic changes and the outer limbs of the neuroepithelial cells were sticky. There was no indication of inflammatory activity within the eye although there was an accumulation of inflammatory cells in the periphery of the optic nerve near the globe.

Attached document

Vogin Table 1.bmp Vogin Table 2.bmp

			-	De	Differential (%)					
DMSO (mi/kg)	Rouse	-	(× 20°/mm²)	•	L	Other	(g/100 me)	Het (%)	Heat (sec)	(· 301/mm)
	Opens		9.4	42	34	2	13.9	42	15.1	190
		-	7.2	23	75	3	12.3	-	12.2	110
		31	14.2	31	*	•	11.9	39	12.1	130
		73-70	7.0	•	2	•	91.0	•	20.7	136
9 December	Dermani		12.1	43	57	•	13.5	**	16.5	135
		26	4.8	×	42	4	14.3	44	12.7	143
		55	**	29	•	1	12.4	39	13.3	155
		73-79	4.5	•	*		12.8	41	14.1	150
	Oral	•	9.7	36	•	1	14.2	45	143	120
		26	7.1	*	64	,	13.4	**	12.4	200
		22	67	45	30	7	11.9	4	13.6	170
		78	7.5	49	57		23.1	43	14.9	120
5 Onsi	Onsi		11.2	47	13		14.9	42	14.2	130
		24	5.3	77	73	2	12.3	40	13.0	130
		24 52 76	8.1	77 28 39	45	7	12.5	40	13,6	150
		76	7.2	'n	-	,	13.5	42	14.9	120

TABLE 2 MEAN BIOCHEMICAL FINDINGS IN MONKEYS RECEIVING DMSO (90% v/v)

DMSO (ml/kg)	Route	Week of study	BUN (mg/100 ml)	Glucose (mg/100 ml)	SGPT (units)	SAP (units)
0	Dermal	0	16	73	17	11.9
		26	39	87	20	27.1
		51	25	85	39	10.8
		73-78	24	68	25	6.1
9	Dermal	0	14	73	17	12.7
		26	32	83	21	19.1
		51	27	70	35	14.2
		73-78	25	72	18	6.4
0	Oral	0	16	67	21	12.0
		26	23	105	19	18.7
		52 '	14	72	37	12.5
		78 :	18	74	19	8.8
3	Oral	0 -	17	104	18	16.3
		26	17	100	29	20.5
		52)	20	102	27	10.8
		78	22	90	17	5.2

Source

: Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability Flag

(2) valid with restrictions Critical study for SIDS endpoint

12.08.2003

Type **Species**

monkey

78 / 157

(150)

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Sex

Sex Strain : male

other: Macaca mulatta (rhesus)

Route of admin. : i.v.

Exposure period Frequency of treatm. Post exposure period

m. : 1 x per day iod : 120 days : 3 and 2 g/kg

Control group Method yes, concurrent vehicle other: no data

9 days consecutive

Year : 1981 GLP : no data

Test substance : as prescribed by 1.1 - 1.4

Method

Doses

Daily i.v. doses of 3 or 2 g/kg DMSO in a 40% solution were given respectively to 4 or 1 male rhesus monkeys for 9 consecutive days. The monkeys were monitored before and after treatment for 4 months for changes in blood chemistry, hematology, urine, and ocular, neurological, and cardiovascular systems. At the end of the study all animals were sacrificed and gross and microscopic pathological

examinations were performed.

Result

The blood chemistries, urinalysis, hematology, and ophtalmologic and gross and histological examinations of tissues in the DMSO animals were not significantly different

from those in the control monkey.

The rapid fourfold increase in diuresis over the control values in the monkeys receiving 2 or 3 g/kg DMSO supports previous results. This urine output indicates that DMSO is one of the strongest diuretics known. The absence of any microscopic or gross structural damage to the kidneys suggests that the coffee-colored urine seen after DMSO administration represents a transient erythrocytic hemolysis, possibly caused by an osmotic gradient due to DMSO in the vascular system. Since this effect was short lived and did not significantly alter the red cell count or

DMSO in the vascular system. Since this effect was short lived and did not significantly alter the red cell count or the hemoglobin and hematocrit values, we believe the coffee-colored urine represented a transient and reversible condition. It does point out, however, the need to

routinely monitor the CBC and hematocrit and hemoglobin whenever DMSO is used in patients at the doses indicated

in this study.

The partial thromboplastin time in DMSO treated monkeys decreased from a normal mean of 46 s to a mean of 18.7 s. These values returned to normal when DMSO administration was stopped. Although the PT values in both saline and DMSO

treated monkeys fluctuated slightly before and after treatment, the values did not significantly differ from the control standard range. By contrast, the PTT values in both DMSO and saline treated animals were abnormal even before treatments began and remained that way after

treatment.

An increase in respiratory rate after DMSO was previously reported. It appears from these studies that DMSO acts as a central respiratory stimulant when given iv in bolus form. A rise in minute respiratory volume concomitant with

the rise in respiratory rate was also reported.

Remark

These data indicate that monkeys receiving a total of 108-127g 40% DMSO or equal saline volumes iv during a 9-d study showed no significant or adverse changes in the chemical and physiological parameters studied. No gross or microscopic pathology was found in any monkey at the end of the 4-mo observation period.

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(40)

Source

Sex

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability

03.06.2003

(2) valid with restrictions

Type Species

rat male

Strain Route of admin. Exposure period Sprague-Dawley inhalation

Frequency of treatm.

6 weeks 7 hr/day, 5 days a week for a total 30 exp.

Post exposure period Doses

none 0.2 mg/l

Control group NOAFL

yes, concurrent no treatment = .2 mg/l

Method Year GLP

other 1969 no

Test substance

as prescribed by 1.1 - 1.4

Method

A total of 32 male Sprague-Dawley rats were exposed to 200 mg DMSO per cubic meter of air for 7 hr/day, 5 days a week, for 6 weeks for 30 exposures. Control rats were exposed to a

normal chamber environment.

All animals were examined for toxic signs prior to, during, and subsequent to each exposure. These signs included diarrhea, lacrimation, dyspnea, ataxia, anorexia, and unusual behavior. All animals were allowed food and water ad libitum when not being exposed. Body weight was obtained from each animal prior to the first exposure and after the final exposure. Blood was obtained prior to the first exposure and after the final exposure for hernatologic evaluation. Hemoglobin concentration, packed erythrocye volume as expressed by the microhematocrit, and total leukocyte and reticulocyte counts were done on all animals using standard techniques. At the termination of the experiments, all animals were sacrificed with an overdose of a barbiturate. Blood and tissue specimens were obtained for biochemical analysis, and gross observations of the organs were made. Sections of the heart, lung, liver, spleen, and

Result

Reliability

kidney were taken for histologic examination. There were no outward toxic signs noted in any of the exposed animals throughout the experimental period of 6 weeks. The characteristic garlic-like odor was detected in the breath of each of the rats after the first day of exposure, and the hair began to appear slightly yellow after the first week. All animals gained weight normally, showing a mean gain of 95.8 % for the test animals and 92.5 % for the controls. No significant alterations were noted in hemoglobin concentrations, microhematocrit, total leukocyte counts, reticulocyte counts, serum glutamic pyruvic and glutamic-oxalo-acetic transaminase activities, liver

Gross and histopathologic examinations of organs and tissue were unremarkable except for nonspecific inflammatory

alkaline phosphatase activity, or liver lactate concentrations.

changes in the lungs and livers of nearly all animals, including controls.

Source Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

(3) invalid

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(63)

Limited hematological, biochemical and histopathological

22.07.2003

investigations

Type

rat

Species Sex Strain

male/female Sprague-Dawley

Route of admin. Exposure period Frequency of treatm. gavage 18 months daily, 5 days/wk

Post exposure period

none 1 - 3 - 9 ml/kg/d (1100 - 3300 - 9900 mg/kg/d)

Doses Control group NOAEL

yes, concurrent vehicle = 1100 mg/kg bw = 9900 mg/kg bw other: no data

LOAEL Method Year GLP

1975 no

Test substance

as prescribed by 1.1 - 1.4

Method

A 50% aqueous solution of DMSO was administered by oral gavage daily, 5 days a week for a total of 18 months (78 weeks). Groups of 50 male and 50 female Sprague Dawley rats received daily doses of 1, 3 or 9 ml/kg DMSO; a control group received 9 ml distilled water/kg/day. After 52 weeks, 10 males and 10 females from each were sacrificed. Animals were observed daily, weighed weekly and food intake was calculated at weekly intervals. Ophthalmoscopic examination of the eyes of all animals was made before dosing and then at regular intervals throughout the study. Hematology studies (PCV, hemoglobin, total and differential white cell count and prothrombin index), together with urinalysis and measurement of urine concentration were performed on sample animals from each group after 4, 12, 20, 32, 51, 60 and 72 weeks. After 78 weeks remaining animals were sacrificed and their tissues preserved.

Result

Oral dosing with a 50% aqueous solution of DMSO was continued for a period of 18 months (with an interim sacrifice of some members of each group after one year). Mortalities were few and could not be related to DMSO treatment. Occasional behavioral changes, persisting for about 5 min after dosing, were observed. These consisted of stretching and arching of the back, accompanied by an in-drawing of flanks and abdomen, and were attributed to

abdominal discomfort.

Bodyweight records indicated a dose-related depression of weight gain in both sexes, with the exception of males receiving 1 ml/kg (Figures 7 and 8). There was no accompanying reduction of food intake. Laboratory investigations were limited to hematological tests, the only abnormality being a slight reduction of haemoglobin and

PCV in male rats receiving 9 ml/kg.

Examination of the eye revealed no changes in the retira or vitreous. No peripheral (equatorial) opacities were seen and there was no difference in incidence of polar opacities between test and control animals. Prominent nuclear annuli were seen in a small number of animals towards the end of the study, as expected, but there was no dose-relationship to suggest any increase resulting from administration of DMSO. The only relevant finding was some degree of change in

ld 67-68-5 Date 17.05.2004

the refractive index of the nuclear region in 3 rats

Attached document

receiving 9 ml/kg. Noel Fig 7.bmp Noel Fig 8.bmp

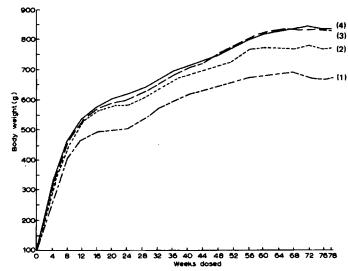


Fig. 7. Weekly group mean bodyweights of male rats. — · · · · · · Group 1, 9 ml DMSO/kg; — · · · · , Group 2, 3 ml DMSO/kg; — · · · , Group 3, 1 ml DMSO/kg; — · · · , Group 4, Control.

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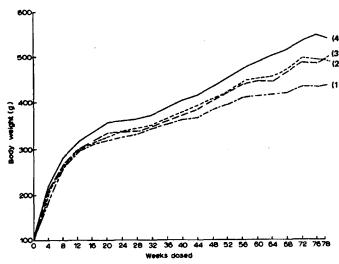


Fig. 8. Weekly group mean bodyweights of female rats. — · — , Group 1, 9 ml DMSO/kg; · · · · · , Group 2, 3 ml DMSO/kg; — — , Group 3, 1 ml DMSO/kg; — — , Group 4, Control.

Source

: Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability

(3) invalid

Limited investigation: no blood chemistry and

histopathology.

22.07.2003

(119)

(30)

Туре

Species Sex Strain

mouse male/female Swiss

Route of admin. Exposure period Frequency of treatm.

gavage 3 weeks (5 g/kg), 5 weeks (2.5 g/kg) or 10 weeks (2 g/kg)

Post exposure period

none

Doses Control group 5, 25.0 2.0 g/kg/day yes, concurrent no treatment

LOAEL Method = 2000 mg/kg other: no data 1969

Year GLP

no

Test substance

as prescribed by 1.1 - 1.4

Result Source Two mice died at 2 g/kg. Body weight gain was reduced by 20% at 2.5 g/kg and by 21-27% at 5 g/kg.
Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability 24.12.2002 (3) invalid

Type Species

mouse

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Sex : male/female Strain : Swiss Route of admin. : i.p. Exposure period : 5 weeks

Exposure period : 5 weeks
Frequency of treatm. : 6d/week
Post exposure period : none
Doses : 2.5 g/kg/day

Control group : yes, concurrent no treatment NOAEL : e2500 mg/kg bw

Method : = 2500 mg/kg l

Year : 1969 GLP : no

Test substance : as prescribed by 1.1 - 1.4

Result : Microscopic examination of the tissues in mice revealed that

repeated doses of 2.5 g/kg given i.p. causes tubulo nephritis and a more or less diffuse necrosis of the intraperitoneal organs in the area of injection.

Source : Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability : (3) invalid

24.12.2002 (30) (32)

Type

Species : rabbit Sex : no data

Strain : New Zealand white

Route of admin. : dermal
Exposure period : 30 days
Frequency of treatm. : daily
Post exposure period : none
Doses : 1.0, 5.0 g/kg

Control group : other: saline

NOAEL := 1000 mg/kg

LOAEL := 5000 mg/kg

Method : other

 Method
 : other

 Year
 : 1971

 GLP
 : no

Test substance : as prescribed by 1.1 - 1.4

Method : Medical grade DMSO was applied daily to the shaved backs of

rabbits for 30 days at a dose of 1.0 or 5.0 g/kg/day. Control animals received saline. Blood was drawn by cardiac puncture from each rabbit 1 and 7 days prior to treatment, and on treatment day 1, 7, and 30. Serum chemistry and serum enzyme levels were evaluated. Eye lenses of each rabbit were examined with a biomicroscope before and during

treatment. At the end of the study, all rabbits were autopsied and examined for gross pathology.

Result : There was no mortality reported for rabbits receiving daily

dermal exposures to DMSO. Application of DMSO at 1 g/kg/day did not induce significant changes in serum chemistry or enzyme levels. Serum LDH levels were elevated in rabbits receiving 5 g/kg/day. There were no gross lesions in DMSO or

control animals examined at the end of the study.

Eye lenses from all rabbits were examined. None of the animals treated with 1 g/kg/day displayed lenticular changes. All of the rabbits treated with 5 g/kg/day displayed microscopic changes, which were first detected after 10-15 days treatment. Results were the same in groups

of rabbits that received the same doses of DMSO by

Source

ld 67-68-5 Date 17.05.2004

(160)

intraperitoneal injection. Similar lens changes were observed in rabbits which received 5 g/kg/day; no lens changes were seen in the 1 g/kg/day group. Lenticular effects resulting from DMSO treatment appears to be the same regardless of the route of administration, however, the

effects are dose dependent.

Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (3) invalid

Histopathology limited to the eye.

12.08.2003

Type

Species rabbit Sex male/female Strain New Zealand white

Route of admin. dermal **Exposure** period 90 days not reported Frequency of treatm.

Post exposure period none

Doses 2.2, 4.4, and 8.8 g/kg (as 100 or 50 % solutions)

Control group yes, concurrent vehicle NOAEL = 2200 mg/kg bw

Method other Year 1967 **GLP** no

Test substance as prescribed by 1.1 - 1.4

Method Six groups of 2 male and 2 female rabbits received dermal

application of 100% DMSO solution at dose levels of 2, 4, and 8 ml/kg or 50% DMSO solution at 4, 8 and 16 ml/kg for 90 days. The rabbits were examined ophthalmoscopically prior to

onset of treatment and after 90 days of treatment.

Result Bilateral changes in the ophthalmoscopic appearance of the

lens were seen in the two higher dose level groups receiving the 100% DMSO solution and the 50% DMSO solution.

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (3) invalid

24.12.2002 (126)

Type

Species rabbit Sex male/female Strain New Zealand white

Route of admin. other: dermal to normal and abraded skin 22 weeks 5 day/week

Exposure period Frequency of treatm. Post exposure period

none 1.65 and 3 g/kg/d (as 50% or 90% DMSO solutions)

Control group yes, concurrent vehicle = 1650 mg/kg bw LOAEL

Method other Year 1967

GLP Test substance as prescribed by 1.1 - 1.4

Method 50% and 90% solution of DMSO in water were applied 5 days

> weekly to the skin of 2 series of 5 groups of 6 male and 6 female rabbits (3 months old) at the dose levels of 0, 3 and 9 ml/kg/d. In one series solutions were applied on the intact skin and the other on the abraded skin. The rabbits

ld 67-68-5 Date 17.05.2004

were examined opthalmoscopically in the 22nd dose-week, slit

lamp biomicroscopy was done as well.

Result : Lenticular changes were observed in rabbits receiving daily

dermal applications of 9 ml/kg, 50% and 90% DMSO, after 15-20 weeks. After 22 weeks these changes were observed in all rabbits receiving 9 ml/kg 90% DMSO and some receiving

the smallest dose administered, 3 ml/kg DMSO.

Source : Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability : (3) invalid

24.12.2002

(126)

Туре

 Species
 : rabbit

 Sex
 : male/temale

 Strain
 : New Zealand white

Route of admin. : dermal Exposure period : 6 months

Frequency of treatm. : daily, 5 days a week

Post exposure period : 12 weeks

Doses : 1.5, 2.7, 4.5, 8.1 ml/kg/day (1650, 2970, 4950, 8910 mg/kg/day)

Control group : other: distilled water LOAEL : = 1650 mg/kg

Method: otherYear: 1975GLP: no data

Test substance : as prescribed by 1.1 - 1.4

Method : Rabbits received dermal application of DMSO once a day, 5

days a week, to a closely shaven area of approximately 150 sq cm. DMSO was applied as 50% or 90% aqueous solution; volumes applied were equivalent to 1.5, 2,7, 4.5 or 8.1 ml undiluted DMSO per kg. Groups of males and females received daily applications of DMSO to normal and abraded skin. Control animals received distilled water. Treatments

continued for 6 months; animals were kept under observation for an additional 12 weeks after treatment was terminated.

Each rabbit was observed daily, weighed weekly and its water consumption was recorded during week 26. Ophthalmoscopic examination was performed on all animals before dosing commenced and then after 5, 8, 14, 20, 22, 28 and 33 weeks. Haematological investigations comprising PCV, haemoglobin, total and differential white cell count, and ESR, were performed initially and at 4, 12, 26 and 32 weeks. At termination, each animal was subjected to a post mortem examination, with subsequent organ weight analysis and

histopathology

Result : Rabbits received dermal applications of DMSO to normal and

abraded skin for a period of 23 weeks, when ocular changes were observed. Treatment was withheld from animals showing ocular changes; the remaining animals continued to receive DMSO applications for the scheduled 26 weeks (6 months).

Mortality was high in all groups, however there was no significant differences in mortality between groups. There were no clinical signs to suggest systemic toxicity. Local dermal reactions of slight erythema and mild edema were observed following each application. The degree of reaction was similar in all groups. Abrasion of the skin did not increase the reaction. Hair growth was normal and there were no dermal reactions observed during the post-treatment

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observation interval

Water consumption, measured during week 26, was markedly increased in animals receiving 8.1 m/kg; a lesser increase in water consumption was noted for other groups receiving DMSO.

Macroscopic and hematological examinations, organ weight analysis, and histopathology did not reveal any adverse effects. There were no changes in dermal morphology except for random occurrences of inflammatory reaction

Adverse ocular (lenticular) effects were observed. These were restricted to the lens, and consisted of nuclear refractive changes. There was no effect on the peripheral lens, vitreous humor, or retina. Abrasion of the skin had no

effect on the incidence of lenticular effects.

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

(3) invalid

No blood chemistry analysis. Excepted the eyes, the organs

subject to the histopatological examination are not

reported.

12.08.2003

Type

Source

Reliability

Species dog

Sex male/female

Strain other: Pembrokeshire Corgis

Route of admin. gavage **Exposure period** 2 years Frequency of treatm. daily, 5 days/wk

Post exposure period

none

1; 3; 9 ml/kg (1100 - 3300 - 9900 mg/kg/day) **Doses** Control group other: 1 ml/kg water LOAEL = 1100 mg/kg bw Method other: no data

Year 1975 GLP no data

Test substance as prescribed by 1.1 - 1.4

Method

Dogs were pure-bred Pembrokeshire Corgis obtained at the age of 4-5 months and were assigned to groups; 5 males and 5 females per group. Dogs were dosed orally, once per day for 5 days a week, by gastric intubation with a 50% aqueous solution of DMSO. Groups recieved 9, 3, or 1 ml/kg/day. A control group received 1 ml distilled water/kg/day.

After 18 weeks, unexpected eye changes were observed. Dosing was continued for half the dogs in each group for the remaining 86 weeks; the other half was not treated but observed for signs of recovery. The total interval of dosing was 2 years. Clinical signs were recorded daily; food intake was measured twice daily. All animals were subject to regular examimantion, including ophthalmoscopy. ECG, clinical chemistry, hematology, and urinalysis monitoring was conducted before and at regular intervals during the

At the end of the study, all animals were sacrificed and subjected to detailed necropsy. Principal organs were weighed and tissues taken for histopathological examination. Eyes were examined and the lenses removed and weighed; the

ld 67-68-5

Date 17.05.2004

Result

volume of aqueous humour was measured.

There were no marked clinical signs and only one death occurred, after the 4th dose at the intermediate level. This was caused by accidental inspiration of DMSO, leading to a severe pulmonary reaction; the animal was replaced. Occasional isolated bouts of vomiting were seen at 9 ml/kg/day and transitory "head shaking" was temporarily observed during weeks 11 and 12 at this and the 3 ml/kg/day level.

No adverse effects on bodyweight and food intake were recorded. ECG records were normal throughout except for a transient, minimal slowing of the heart rate in recordings made after 4 weeks. Terminal radiology of excised bone showed no evidence of osteoporosis.

Laboratory investigations confirmed the persistence of diuresis in dogs receiving 3 ml/kg and above (increased "overnight" urine volumes, reduced SG and increased water intake during periods of measurement). No renal damage resulted; normal function was demonstrated by the sensitive urine concentration tests and normal plasma urea levels.

The only other change related to red cells: there were persistently increased PCV and haemoglobin levels, and total red cell count, at 9 ml/kg (Table 1). The red cells had normal haemoglobin concentrations (MCHC) and were of normal size (MCV). Bone marrow examination prior to termination revealed no evidence of toxic changes. It seemed possible that the constant diuresis had resulted in a balance with a slightly higher degree of haemoconcentration than is normally found. No increase in serum proteins could be demonstrated to confirm this possibility.

Ocular effects were observed after 5-10 weeks dosing in the dogs receiving 9 ml/kg including central (nuclear) lenticular changes with alteration of the refractive index (myopia) and by the fifth month, transitory equatorial opacities, central (nuclear) opalescence, and changes in the vitreous humour. Similar effects were observed in dogs receiving 1 and 3 ml/kg but they occurred more slowly.

No abnormalities, other than those in the eye, were detected during macroscopic or microscopic examination of organs. The retina was normal. Changes in the lens included an increase in insoluble protein at all dose levels and a reduction of water content in the 3 and 9 ml/kg group. There was a significant reduction of soluble protein and a reduction in glutathione content in the 9 ml/kg group.

Attached document

Noel Table 1.bmp

Noel Table 1 continued bmp

ld 67-68-5 Date 17.05.2004

Works detect Humber High Intermediate	TREAL ANALYSS: LEAST SIGNIFICANT DIFFERENCES LEV Charles L.S.D.	
and not desired of days Doord ND Dread ND	Desired HID Decired HID 590° 1%	
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ource :	Atofina, Paris-la-Défense, France	
teliability :	Atofina Paris La Défense Cedex (3) invalid The blood chemistry parameters and the organs (excepted the eyes) subject to the histopathological examination are not	
6.02.2003	reported.	(11:
ype : Species : Sex : Strain : Coute of admin. :	dog male/female other: pembrokeshire corgis gavage	
exposure period : requency of treatm. :	132 days	
ost exposure period :	not reported 12 days	
loses :	see freetext	
control group :	yes, concurrent no treatment other	
ear :	1967	
ELP : est substance :	no as prescribed by 1.1 - 1.4	
	as prescribed by 1.1 - 1.4	
	3 groups of 10 dogs (young Pembrokeshire Corgis) received a daily oral dose of 4.5 ml/kg for 45 days, then 9 ml/kg; 1.5 ml/kg for 45 days, then 3 ml/kg; or 0.5 ml/kg for 45 days, then 1 ml/kg DMSO for up to 120 days. Because of changes noted in the group receiving the highest dose level, 5 dogs in this group received non DMSO after the 120th days of treatment	
lethod :	daily oral dose of 4.5 ml/kg for 45 days, then 9 ml/kg; 1.5 ml/kg for 45 days, then 3 ml/kg; or 0.5 ml/kg for 45 days, then 1 ml/kg DMSO for up to 120 days. Because of changes noted in the group receiving the highest dose level, 5 dogs in this group received non DMSO after the 120th days of treatment. Lenticular changes were observed in the dogs receiving 4.5 - 9 ml/kg after 68 days. Slight effects were seen in the 0.5 -	
esuit :	daily oral dose of 4.5 ml/kg for 45 days, then 9 ml/kg; 1.5 ml/kg for 45 days, then 3 ml/kg; or 0.5 ml/kg for 45 days, then 1 ml/kg DMSO for up to 120 days. Because of changes noted in the group receiving the highest dose level, 5 dogs in this group received non DMSO after the 120th days of treatment. Lenticular changes were observed in the dogs receiving 4.5 - 9 ml/kg after 68 days. Slight effects were seen in the 0.5 - 1 ml/kg dose group, after 68 days of treatment. Atofina, Paris-la-Défense, France	
dethod :	daily oral dose of 4.5 ml/kg for 45 days, then 9 ml/kg; 1.5 ml/kg for 45 days, then 3 ml/kg; or 0.5 ml/kg for 45 days, then 1 ml/kg DMSO for up to 120 days. Because of changes noted in the group receiving the highest dose level, 5 dags in this group received non DMSO after the 120th days of treatment. Lenticular changes were observed in the dogs receiving 4.5 - 9 ml/kg after 68 days. Slight effects were seen in the 0.5 - 1 ml/kg dose group, after 68 days of treatment.	(126
lethod : esult : ource : eliability :	daily oral dose of 4.5 ml/kg for 45 days, then 9 ml/kg; 1.5 ml/kg for 45 days, then 3 ml/kg; or 0.5 ml/kg for 45 days, then 1 ml/kg DMSO for up to 120 days. Because of changes noted in the group receiving the highest dose level, 5 dogs in this group received non DMSO after the 120th days of treatment. Lenticular changes were observed in the dogs receiving 4.5 - 9 ml/kg after 68 days. Slight effects were seen in the 0.5 - 1 ml/kg dose group, after 68 days of treatment. Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex	(126

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ld 67-68-5 Date 17.05.2004

Species : dog
Sex : male/female
Strain : Beagle
Route of admin. : gavage
Exposure period : 23 weeks

Frequency of treatm. : 5 days a week

Post exposure period : 31 weeks

Doses : 0 - 2500 - 5000 - 10000 - 20000 - 40000 mg/kg/day

Control group : yes, concurrent vehicle
LOAEL : = 2500 mg/kg

 LOAEL
 : = 2500 mg/kg

 Method
 : other: no data

 Year
 : 1967

Year : 19 GLP : no

Test substance : as prescribed by 1.1 - 1.4

Method : Six pairs (male and female) of young adult beagle dogs

received daily oral doses of 0, 2.5, 5, 10, 20 and 40 g/kg/d DMSO, 5 days a week for up to 23 weeks. The 20 and 40 g/kg dose levels were not tolerated by the dose and were reduced. Administration of DMSO was continued for 23 weeks (107 doses) at which time the DMSO was withdrawn from the seven survivors which were then observed for additional 31 weeks.

The dogs were then humanely killes, and histologic

examination as performed.

The dogs were examined by indirect and direct ophthalmoscopy under 0.5% tropicamide-induced mydriasis prior to testing and at a minimum of monthly intervals during dosing and

after withdrawal.

Result : Changes in the lens of the eye were observed in dogs

receiving 5 g/kg after 9 weeks of administration. At lower dose levels by the 18th week all the dogs were affected. Treatment was withheld after 23 weeks and the animals were observed for 31 weeks. The changes persisted after the withdrawal of DMSO but became slightly less pronounced.

Source : Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability : (3) invalid

24.12.2002 (126)

Туре

Species : pig
Sex : no data
Strain : no data
Route of admin. : dermal
Exposure period : 123 days
Frequency of treatm. : twice daily

Post exposure period : none

Doses : 825, 1485, 2475 and 4455 mg/kg wb/d

Control group : yes, concurrent vehicle
NOAEL : = 825 mg/kg bw

Method : = 825 mg/kg t

Year : 1967 GLP : no

Test substance : as prescribed by 1.1 - 1.4

Method : 50 and 90% solutions of DMSO in water at the dose levels of

0, 1.5 and 4.5 ml/kg/d were applied twice daily for up to 123 days to the skin of groups of 8 pigs. Ophthalmoscopic examinations were performed on the 90th, 113th and 123rd day

of DMSO administration.

Result : Dermal application of 4.5 ml 90 % DMSO/kg twice a day caused

lens changes by 90 days of treatment. Lens changes were

ld 67-68-5 Date 17.05.2004

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visible on the 113th day in several of the pigs in the

groups receiving 4.5 ml 50% DMSO/kg and 1.5 ml 90% DMSO/kg.

No changes were visible in the pigs receiving 1.5 ml 50%

DMSO/kg on the 123rd day. Atofina, Paris-la-Défense, France

Source Reliability

24.12.2002

Atofina Paris La Défense Cedex (3) invalid

(126)

GENETIC TOXICITY 'IN VITRO' 5.5

Type

Salmonella typhimurium reverse mutation assay

System of testing Test concentration Cycotoxic concentr. Strains TA97, 98, 100, 102, 104, 1535, 1537, 1538 100, 333, 1000, 3333 and 10000 µg/plate $> 10000 \,\mu\text{g/plate}$

Metabolic activation Result

with and without

Method

negative other: comparable to OECD Guide-line 471

Year **GLP**

1992 no data other TS

Test substance Test substance

Dimethyl sulfoxide, CAS no.: 67-68-5, Source Burdick and Jackson, Purity:

Method

DMSO was tested as a coded chemical in two separate laboratories. Concentrations of DMSO (100, 333, 1000, 3333,

and 10,000 ug), overnight culture of S. typhimurium (0.05-0.10 ml), and S-9 mix or buffer were incubated without shaking for 20 minutes. The top agar was added and the

contents of the tubes were mixed and poured onto the surfaces of petri dishes. His+ (histidine dependent) colonies arising on plates were machine-counted after two

days incubation.

Initial testing was without metabolic activation, with 10% rat liver S-9, or with 10% hamster liver S-9. After a

negative result was obtained, DMSO was retested without S-9

and with 30% S-9 from rat and hamster.

Test condition

Positive controls:

- Without S9: Sodium azide (TA 1535 and TA100), 9-aminoacridine or ICR-

191 (TA 97 and TA1537), 4-nitro-o-phenylenediamine (TA98)

- With S9: 2-aminoanthracene (all strains)

Result

The positive control chemicals induced a significant increase of the revertant frequency in all tester strains, either with or without metabolic

DMSO was negative, in the presence and absence of metabolic

activation, in all tester strains. Atofina, Paris-la-Défense, France

Source Reliability

Atofina Paris La Défense Cedex (1) valid without restriction

Flag

Critical study for SIDS endpoint

06.05.2004

Type System of testing Test concentration

Metabolic activation

Salmonella typhimurium reverse mutation assay Salmonella typhimurium TA 98, 100, 1535, 1537, 1538

up to 500 mg per plate Cycotoxic concentr. Not reported

with and without

ld 67-68-5 Date 17.05.2004

Result negative

Method other: Ames B.N. et al., Mutat. Res., 31, 347-364, (1975)

Year 1975 GLP no

as prescribed by 1.1 - 1.4 Test substance

Method DMSO was tested in the standard plate incorporation assay

using five S. typhimurium tester strains (TA 98, 100, 1535, 1537, 1538) in the presence and absence of metabolic activation. Multiple geometric dilutions were tested in duplicate plates, starting with the maximum non-toxic dose

of 500 mg/plate, with and without rat liver S-9 mix.

Result DMSO was tested to a maximum test dose of 500 mg per plate.

Multiple doses were tested in duplicate.

DMSO was negative, in the presence and absence of metabolic activation,

in all strains tested.

Atofina, Paris-la-Défense, France Source Atofina Paris La Défense Cedex

Reliability (4) not assignable

Documentation insufficient for assessment

06.05.2004 (107)

Salmonella typhimurium reverse mutation assay Type

Salmonella typhimurium TA 98, 100, 1535, 1537, 1538 System of testing Test concentration up to 1.4 mM per plate

Cycotoxic concentr. not reported Metabolic activation with and without

Result negative

other: Ames B.N. et al., Mutat. Res., 31, 347-364, (1975) Method

Year 1981 GLP no data **Test substance** other TS

Source

Dimethyl sulfoxide Test substance

CAS no.: 67-68-5 purity unknown

Method DMSO was tested in the standard plate incorporation assay

using five S. typhimurium tester strains (TA 98, 100, 1535, 1537, 1538) in the presence and absence of metabolic activation. Multiple geometric dilutions were tested in duplicate plates, starting with the maximum non-toxic dose

tested of 1.4 mM, with and without S-9 mix

EXPERIMENTAL CONDITIONS: **Test condition**

Number of replicates: 2

- Metabolic activation: S9 fraction from Aroclor-pretreated rats

- Positive control chemicals: not reported, however among the 106 compound tested by the author, 62 were found to be mutagenic,

demonstrating the sensitivity of the method used.

- Pre-incubation time: none

DESCRIPTION OF FOLLOW-UP REPEAT STUDY: no data

CRITERIA FOR EVALUATING RESULTS: A clearly positive result as indicated by a dose-related and reproducible increase of his+ revertants

over controls (at least a 3-fold increase).

Result DMSO was tested to a maximum test dose of 1.4 mM per plate.

Multiple doses were tested in duplicate.

DMSO was negative, in the presence and absence of metabolic

activation, in all five tester strains. Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

ld 67-68-5 Date 17.05.2004

(39)

Reliability

: (2) valid with restrictions

06.05.2004

Type

Salmonella typhimurium reverse mutation assay

System of testing Test concentration TA98, TA100, TA1535, TA1537 and TA 1538 up to 5 mg/plate

Cycotoxic concentr. Metabolic activation

with and without negative

Result Method

other: Ames et al, Mut. Res., 1975, 31, 347-364.

Year 1975 GLP Test substance other TS

Test substance

Dimethyl sulfoxide CAS no.: 67-68-5 purity unknown

Test condition

EXPERIMENTAL CONDITIONS: - Number of replicates: no data

Metabolic activation: S9 fraction from Aroclor-pretreated rats-

Vehicle: no data

- Positive control chemicals: included in each experiment (no detail available) (among the 71 compound tested by the author, 26 were found to be mutagenic, demonstrating the sensitivity of the method used).

- Pre-incubation time: 2 hours

DESCRIPTION OF FOLLOW-UP REPEAT STUDY: no data

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability (4) not assignable

Documentation insufficient for assessment

06.05.2004

(136)

Cytogenetic assay Type CHO-cells

System of testing **Test concentration** up to 4990 ug/ml Cycotoxic concentr. Metabolic activation Result

> 4990 µg/ml with and without negative

Method OECD Guide-line 473

1990 Year GLP no data **Test substance** other TS

Test substance Dimethyl sulfoxide, CAS no.: 67-68-5, source: Burdick and Jackson

Laboratories Purity: 99.4%

Cell Culture and Medium: CHO cells were obtained from Litton Bionetics Method

(Kensington, MD) at their fifth passage level after cloning, and were designated CHO-LB. A large stock of cells was initially prepared, and vials were stored at -80°C. To ensure karyotypic stability, cells were not used beyond the fifteenth passage after cloning. Cells were tested regularly for mycoplasma contamination using 4,6-diamidino-2-phenylindole (DAPI)

fluorescence and were found to be free of mycoplasma for all

experiments. Growth and treatment conditions were based on procedures described by Galloway et al. (1985). Cells were grown and exposed to

chemicals at 37°C.

Metabolic Activation: The rat liver microsomal fraction was prepared from Aroclor 1254-induced male Sprague-Dawley rats and was combined with

ld 67-68-5 Date 17.05.2004

cofactors and culture medium to form the metabolic activation system.

Controls:Medium and solvent controls were used with each assay. Solvent controls consisted of culture medium with or without S9 and contained the same concentration of solvent as the test cultures (0.5 or 1%). Mitomycin C (MMC; Sigma) was used in the experiments without metabolic activation, and cyclophosphamide (CP; Sigma) was used in the experiments with activation as positive controls.

Test Chemical Dose Selection - Test concentrations were empirically chosen based on toxicity and cell cycle delay. At least five concentrations of the test chemical were selected; the concentrations were spaced using two merged half-log scales, and the highest concentrations analyzed were those yielding a sufficient number of suitable metaphase cells. The concentrations analyzed generally covered a one-log range. Treatmentin the AB trials without S9, the cultures were treated with the test chemical in medium for 8 hr, washed to remove the test chemical, and treated with colcemid for 2-2.5 hr before cell harvest. In the experiments with activation, cultures were exposed to the test chemical in serum free medium with S9 and cofactors for 2 hr, washed to remove the test chemical and S9, and incubated at 37°C with fresh medium for 8 hr. Colcernid was then added, and the cells were harvested 2 hr later. Thus the total durations of the nonactivated and activated AB experiments were 10 hr and 12 hr, respectively, to give 10 hr growth in medium with serum for each experiment.

Staining and Scoring of Slides - Selection of cells for scoring was based on well-spread chromosomes with good morphology and a chromosome number of 21 ± 2. All slides except the high-dose positivecontrol were coded, and a complete experiment was scored by one technician. Slides were stained in 5% Giemsa for 5 min. In early studies, one hundred cells were scored for each of three concentrations: the highest test concentration in which sufficient metaphase cells could be scored and the next two lower concentrations, covering a one-log range. For later studies, 200 cells per dose were scored; however, fewer cells were scored if a test chemical produced a strong positive response or the chemical was toxic. Cells were analyzed for the following categories of chromosomal aberrations: "simple," defined as a chromatid gap, break, fragment, and deletion or chromosome gap, break, or double minutes; "complex," defined as interstitial deletions, triradials, quadriradials, rings, and dicentric chromosomes, and "other" defined as pulverized chromosomes or cells with greater than 10 aberrations. Chromatid and chromosome gaps were recorded but were not used in the analysis. The frequency of polyploid or endoreduplicated cells was noted only when it seemed excessive: however, these categories were not included in the totals or in the statistical analyses.

Analysis of Data - All categories of aberrations (simple, complex, and other) were combined for the statistical analysis, which was based on the percent of total cells with aberrations. The percent of cells with ABs (i.e., percent of aberrant cells) was used for the analysis, rather than the average number of aberrations per cell. The use of the latter could distort the results in cases where there are a high number of aberrations in only one or two cells. A binomial sampling assumption as described by Margolin et al. (1983) was used to examine absolute increases in ABs over solvent control levels at each dose. The P values were adjusted by Dunnett's method to take into account the multiple dose comparisons. Only the "total" percent cells with aberrations were analyzed, and a positive response was designated "positive" if at least two doses gave significantly increased responses.

Result

: DMSO was tested in CHO cells to a maximum concentration of

Source

06.05.2004

ld 67-68-5 Date 17.05.2004

4990 ug/ml. DMSO did not induce cell toxicity or cell cycle delay, and did not induce an increase in the incidence of

chromosomal aberrations.

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions Flag Critical study for SIDS endpoint

(103) (104)

Type Sister chromatid exchange assay

System of testing CHO-cells Test concentration up to 5000 μg/ml Cycotoxic concentr. > 5000 µg/ml Metabolic activation with and without Result negative

Method OECD Guide-line 479

Year 1987 GLP no data Test substance other TS

Test substance

Purity: 99.4%

Dimethyl sulfoxide, CAS no.: 67-68-5, source: Burdick and Jackson

Method

Cell Culture and Medium: CHO cells were obtained from Litton Bionetics (Kensington, MD) at their fifth passage level after cloning, and were designated CHO?LB. A large stock of cells was initially prepared, and vials were stored at -80°C. To ensure karyotypic stability, cells were not used beyond the fifteenth passage after cloning. Cells were tested regularly for mycoplasma contamination using 4,6-diamidino-2-phenylindole (DAPI) fluorescence and were found to be free of mycoplasma for all experiments. Growth and treatment conditions were based on procedures described by Galloway et al. (1985). Cells were grown and exposed to chemicals at 37°C.

Metabolic Activation: The rat liver microsomal fraction was prepared from Aroclor 1254-induced male Sprague? Dawley rats and was combined with cofactors and culture medium to form the metabolic activation system.

Controls: Medium and solvent controls were used with each assay. Solvent controls consisted of culture medium with or without S9 and contained the same concentration of solvent as the test cultures (0.5 or 1%). Mitomycin C (MMC; Sigma) was used in the experiments without metabolic activation, and cyclophosphamide (CP; Sigma) was used in the experiments with activation as positive controls.

Test Chemical Dose Selection: A series of dilutions were made from the stock solution to achieve 10 test concentrations in a half-log series covering a range of five logs. The highest dose used was based on solubility or toxicity, with the highest dose scored being that allowing sufficient M2 cells for analysis at the time of harvest. In the absence of limitations on solubility or toxicity, the maximum test chemical concentration was 5 mg/ml.

Exposure: In tests without metabolicactivation, cell cultures were exposed to DMSO for 24 hr. In tests with metabolic activation, cultures were exposed to DMSO and rat liver S-9 for 2 hr. Cell toxicity wasdetermined by comparing cell monolayers in treated flaskswith control cultures.

Slide preparation: Mitotic cells were harvested, treated with hypotonic buffer, and resuspended in fixative. Slides were stained and 50 seconddivision M2 cells from each of the top three concentrations were scored for

Reliability

ld 67-68-5 Date 17.05.2004

(103) (104)

(20)

(28)

Result DMSO was tested in CHO cells to a maximum concentration of

5000 ug/ml. DMSO did not induce cell toxicity or cell cycle delay, and did not induce an increase in the incidence of

SCEs.

Atofina, Paris-la-Défense, France Source

Atofina Paris La Défense Cedex (2) valid with restrictions

Flag Critical study for SIDS endpoint 06.05.2004

Type Salmonella typhimurium reverse mutation assay

System of testing Salmonella typhimurium TA 97, TA 98, TA 100 100 to 300 mg/plate

Test concentration Cycotoxic concentr.

Metabolic activation with and without

Result negative Method OECD Guide-line 471

Year 1987 GLP no data

as prescribed by 1.1 - 1.4 Test substance

Source Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions 05.06.2003

Salmonella typhimurium reverse mutation assay

System of testing Samonella typhimurium loci 8AG, Fu, ACA

Test concentration

various (solvent of mutagens) Cycotoxic concentr.

Metabolic activation with and without Result negative Method other: no data Year 1983

GLP no data as prescribed by 1.1 - 1.4 Test substance

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions

05.06.2003 (137)

Gene mutation in Saccharomyces cerevisiae Type System of testing Saccharomyces cerevisiae strain D4 and D5

Test concentration 1400 mM

Cycotoxic concentr. Metabolic activation without

Result negative Method other Year 1977 GLP no

Test substance as prescribed by 1.1 - 1.4

Atofina, Paris-la-Défense, France Source

Atofina Paris La Défense Cedex Reliability (2) valid with restrictions

05.06.2003

other: DNA damage + HGPRT locus Type

System of testing V 79 cells Test concentration 2 mMol

Cycotoxic concentr.

ld 67-68-5 Date 17.05.2004

(129)

Metabolic activation

Result Method no data negative other: no data

Year

GLP

no data

Test substance

as prescribed by 1.1 - 1.4

Remark

Four end points were measured:

the relationships between cell killing, mutation induction and DNA double (dsb) and single (ssb) strand breaks have been studied in V79 cells irradiated with X rays under oxic and anoxic conditions in the presence and in the absence of dimethylsulfoxide (DMSO). Curvilinear relationship were found between all pairs of endpoints, except for dsb versus ssb. Statistical analysis of experimental data has shown that in the absence of DMSO there is evidence of good correlations between cell killing, mutation induction and dsb in oxic and anoxic conditions. However, when DMSO was present, no significant correlation was found. In the presence of oxygen DMSO always exerts a protective effect while in anoxia it is generally much less protective and induces a strong sensitization with respect to mutation induction. Possibly DMSO acts not only as a radical scavenger but also as an agent inducing chromatin relaxation and/or under anoxia, forming highly mutagenic short term radicals. The present data suggest that lethal and mutational events are at least partially independent and not proportional to the initial number of DNA breaks. This may imply that either other kinds of lesions are involved in cell lethality and mutability, or dose dependent repair

Source

mechanisms of dsb have to be considered. Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability

(2) valid with restrictions

29.07.2003

Type

Yeast gene mutation assay Schizosaccharomyces pombe (strain ade6-60/rad10-198,h-)

System of testing Test concentration up to 5% (v/v)

Cycotoxic concentr. Metabolic activation Result

with and without negative

Method other: Loprieno, N. et al. 1976. Mutat. Res. 40: 317-324. Year 1980

GLP

Test substance as prescribed by 1.1 - 1.4

Method

Result

DMSO was tested in the S. pombe forward gene mutation assay in the presence and absence of phenobarbitol(PB)-induced mouse liver S-9. Yeast cells were exposed to 0.5, 2.0, 5.0, and 10% (v/v) for 1, 6 and 24 hr.

In a separate experiment, the effect of DMSO on mouse microsomal enzymatic activity was examined in S-10 supernatant from livers of induced and non-induced mice. Yeast cells exposed to DMSO for 24 hr exhibited 100%

mortality; there was no survival at any time interval to cultures exposed to 10% DMSO. Mortality at 1 and 6 hr was

dose-related up to a concentration of 5%

DMSO was consistently negative in the S. pombe forward gene

mutation assay, both in the presence and absence of

phenobarbitol(PB)-induced mouse liver S-9.

DMSO did not affect the basal or PB-induced microsome-

ld 67-68-5 Date 17.05.2004

(1)

(116)

dependent aminopyrine demethylase activity in mouse liver

S-10 supernatant.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 29.07.2003 : (2) valid with restrictions

003

· (2) valid with restrictions

Type

Year

DNA damage and repair assay

System of testing Test concentration UMU gene expression in salmonella typhimurium TA 1535/p SK 1002
 2 to 8 %

Cycotoxic concentr. Metabolic activation Result Method

without
ambiguous
other: no data
1990

GLP

: no data

Test substance

: as prescribed by 1.1 - 1.4

Remark

The level of betagalactosidase activity which shows umu expression gene was approximately 3.5 times as high as the background level with 8 % DMSO. The authors reported that DMSO was inactive in the SOS chromotest, essentially similar to the present umu test system. The membrane permeability of DMSO may differ between S. typhimurium in the umu-test and E. Coli in the SOS chromotest. The authors concluded that in any case, further studies are required to elucidate the mechanism of SOS induction by DMSO, justifying

our characterisation "Ambiguous results" for this non

validated test.

Source

 Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability

24.12.2002

: (2) valid with restrictions

Type System of testing other: Microscreen prophage-induction assayEscherichia coli WP2s(lambda)

Test concentration Cycotoxic concentr.

0.62 to 10.0%

Metabolic activation

with and without positive

Result Method

other: De Marini et al., Env Mol Mut, 1990, 15, 1-9

Method Year GLP

: 1991 : no data

Test substance

Test compound: dimethylsulfoxide

CAS no.: 67-68-5

Source: Burdick and Jackson, Sigma

Batch number: no data Purity: no data

Method

The procedure used permitted the evaluation of each solvent at a maximum concentration of 10%. 2-NF (-S9) and 2-AA (+S9) were the

positive controls.

Briefly, the first well in a dilution series of a 96-well microtiter plate received 250 μ l of supplemented minimal medium and 50 μ l of either the test compound or the medium control. The remaining wells received 150 μ l of the medium, and 2-fold series dilutions of the compounds or controls were made down the columns of each plate. Each well was inoculated with 75 μ l (c.a. 2 x 10e6 cells) of a resuspended log-phase culture of WP2 5(lambda) and 25 μ l of medium or S9 mix (2.5%) from rat-liver homogenate from male Sprague-Dawley rats. After incubation overnight at 37°C, the wells were scored for turbidity, with turbid wells indicating cell growth and clear wells indicating cytotoxicity and/or inhibition of cell growth. The concentration of

Id 67-68-5

Date 17.05.2004

lambda bacteriophage was determined by sampling at least the first 5 turbid wells adjacent to a clear well. A diluted sample was plated onto indicator cells, the plates were incubated overnight at 37°C, and plaques were counted by hand. The dilution tubes were sampled in duplicate, and all experiments were repeated at least twice.

Control values were determined for each experiment (total of 6 Expts.) by calculating the average PFU/plate for 5 wells (2 plates/well), which was the average for 10 plates. Because of the small variability in the assay, identical or similar control values were obtained among several experiments. In addition, because several compounds were tested per experiment (and, thus, shared the same control values), identical control values occur for some compounds.

Previous studies have shown that an induced PFU/plate that reaches the upper limit of the 99% confidence interval represents an approximate 3-fold increase over the background PFU/plate. Consequently, a dose-related increase of induced PFU/plate that reached or exceeded a 3-fold increase in PFU/plate was considered a positive response. If a solvent reached or exceeded this fold increase at only one dose, the result was scored as a weak positive (w +). A summary response was given to each solvent based on the reproducibility of the results from two independent experiments.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 24.12.2002

(2) valid with restrictions

Type

other: DNA polymerase-deficient mutation assay Escherichia coli

System of testing Test concentration Cycotoxic concentr.

50 μ l/plate

Metabolic activation Result

with and without negative

Method Year

other: Slater et al., Cancer Res., 1971, 31, 970.

no

GLP Test substance Method

as prescribed by 1.1 - 1.4

The cultures were obtained in a lyophilized state and after reconstitution stored on agar slants of medium HA in a refrigerator. Fresh stock slants were prepared weekly. All test compounds were applied to a center well in the plate which was prepared for use with experiments involving metabolic activation. The 50 μ I of the test compounds was applied directly to the well which had a total capacity of about 100 pl.

In a typical assay, each test compound was applied to two plates containing the pol A+ organism and two plates containing the pol Aorganism. The plates were then incubated for 16 h and the zones of inhibition were measured. Control plates containing dimethyl sulfate, ampicillin and colistin were run with every assay. Ampicillin and colistin were included to ascertain constancy of strain characteristics. Both cultures are about equally sensitive towards these antibiotics. Dimethyl sulfate served as a positive control since its differential toxicity toward the two cultures is considerable and consistent.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 24.12.2002 (2) valid with restrictions

(66)

(41)

Type

: DNA damage and repair assay

ld 67-68-5 Date 17.05.2004

(155)

System of testing : Escherichia coli PQ37
Test concentration : 7.8 ng/ml to 7.8 mg/ml

Cycotoxic concentr.

Metabolic activation : with and without Result : negative Method : other Year : 1987 : no data

Test substance : Other TS

Test substance : Test compound: Dimethyl sulfoxyde

CAS no.: 67-68-5 Source: Carlo Erba Batch: 2384 M 100

Method

The SOS chromotest was performed with the kits provided by Orgenics, and following exactly the procedure described by the manufacturer.

The bacteria growing time was 3 h at 37°C; the compounds were incubated with bacteria for 2 h at 37°C and the colour development lasted 60 or 90 min at 37°C before optical density readings. For each assay, the colour development was checked with serial dilutions of pure galactosidase provided with the kit; as recommended, 4-nitroquinoline oxide was used as directly mutagenic substance and a viability control of the bacteria was performed at each dilution of the chemicals tested.

The efficacy of the S9-mix provided with the kits (lot number 019270) was not evaluated.

At least 7 dilutions in water were assayed. Multichannel micropipettes were used to deliver the bacterial suspension, the substrate solutions for beta-galactosidase and alkaline phosphatase and the stopping solution; 1 or 10 μ l glass microconstriction pipettes (Pedersen type) were used to sample the various dilutions of chemicals under examination.

The analysis of the genetoxic activity of the tested compounds was carried out quantitatively with a photometer.

Source : Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex Reliability : (2) valid with restrictions

24.12.2002 (20)

Type : other: Mitotic and meiotic chromosome gain in Saccharomyces

cerevisiae

System of testing : Saccharomyces cerevisiae BR1669

Test concentration : 16.3 to 52.4 mg/ml

Cycotoxic concentr.

Metabolic activation

Test substance : as prescribed by 1.1 - 1.4

Source : Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability : (2) valid with restrictions 05.06.2003

Type : Yeast gene mutation assay

System of testing : Neurospora crassa
Test concentration : N/A see remark

Cycotoxic concentr.

ld 67-68-5 Date 17.05.2004

(23)

(67)

Metabolic activation

Result Method without negative other: no data

Year

GLP

no data

Test substance

as prescribed by 1.1 - 1.4

Remark

This paper is a compilation of data found in literature. All results are negative for DMSO.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

other: in vitro micronucleus assay

Reliability

(4) not assignable

SHE cells

no data

no data

negative

05.06.2003

Type System of testing

Test concentration Cycotoxic concentr.

Metabolic activation

Result Method

other: Schmuck et al., Mut. Res., 1988, 203,397-404 Year

GLP no data

Test substance as prescribed by 1.1 - 1.4

Source Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability (3) invalid 05.06.2003

Mammalian cell gene mutation assay Type

System of testing Mouse lymphoma L5178Y cells, tk-/tk+ assay 0.74-2.11 M Test concentration Cycotoxic concentr. >= 1.83 M

Metabolic activation without Result negative Method other: equivalent to OECD Guide-line 476

Year **GLP** no data

Attached document

Test substance other TS

Test substance Dimethyl sulfoxide CAS no.: 67-68-5

purity unknown Amacher Table 1.bmp

ld 67-68-5 Date 17.05.2004

Tru compound or treatment	Constitution	Coll spentral	Mutacia per
	(moienty)	(% of control)	10. 5144
12. Dimethri sulfentile		100	0.14
ICE - 163%1	7.04 x 10 ⁻³	17.0	0.00
	9.46 * 10-1	B4	9.73
	12.00 K 10 ⁻¹	87	0.41
	14.06 H 10*5	7)	9.44
	12.40 K 10 ⁻³	64	0.10
	10.00 X 10 ⁻¹		8.67
	18-31 F 10**	10	1.80
	31.12 × 10*5		1.11

Source

: Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability

(3) invalid

Tested only without metabolic activation

06.05.2004

(3)

(158)

Type

DNA damage and repair assay

System of testing Test concentration primary rat hepatocytes 0.14 M

Cycotoxic concentr.

Metabolic activation

without negative

Result Method

OECD Guide-line 482

Year GLP

no data

Test substance as prescribed by 1.1 - 1.4

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability

(3) invalid

Only one concentration tested

05.06.2003

Type

Salmonella typhimurium reverse mutation assay Salmonella typhimurium TA100, 1535, 98, 1538, 2637, 1537, 102, 104, 97

System of testing Test concentration Cycotoxic concentr.

14 to 40 %

Metabolic activation Result

with and without ambiguous OECD Guide-line 471

Method Year

1993

GLP

no data

Test substance

as prescribed by 1.1 - 1.4

Remark

The concentrations used in this work are unusual. Mutagenicity of DMSO was induced at very high

ld 67-68-5 Date 17.05.2004

(70)

concentrations, where strong cytotoxicity was observed. According the authors themselves, DMSO concentrations used in the routine AMES are usually between 7 and 14%. Therefore they suggest that the routine use of DMSO in bacterial mutagenicity test will not influence the results obtained with test compounds since DMSO concentrations having mutagenic activity (more than about 25 % in strains TA 1537 and TA 2637). These effects are seen with and without S9 mix, after 20 min of incubation.

At a concentration of 33% some lethal toxicity was observed

in some strains, and overt toxicity in all strains was

observed at 37 and 40%.

Additionally Escherichia coli WP2 and WP2UVRA were used, with the same positive results only in the absence of S9 mix, only after 20 min preincubation, only at 33% concentration on WP2UVR A. Lethal effects occured in both

strains at 40% concentration.

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex (3) invalid

Reliability

29.07.2003

Type

Mammalian cell gene mutation assay

System of testing CHO cells, HGPRT assay

Test concentration 2-16% Cycotoxic concentr. >= 10% Metabolic activation without Result negative

Method OECD Guide-line 476

Year 1984 GLP no data Test substance other TS

dimethyl sulfoxide (CAS 67-68-5), purity not specified Test substance

Attached document DMSO10.bmp

(2)

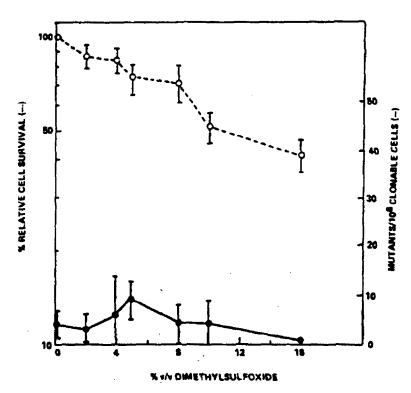


Fig. 2. Mutagenicity and toxicity of dimethyl sulfoxide in CHO cells. Dimethylsulfoxide concentration is indicated as a percent in test medium. Sample means ± 1 S.D. are shown for triplicate trials. The mutant frequency at 5% DMSO was not significantly different than the mutant frequency for solvent controls for $p \le 0.01$.

Source

: Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability

(3) invalid

Only tested without metabolic activation

06.05.2004

Type

Mammalian cell gene mutation assay Mouse lymphoma L5178Y cells, tk-/tk+ assay 0.746-1.55 M

System of testing Test concentration Cycotoxic concentr. Metabolic activation Result

>= 1 M without ambiguous

Method OECD Guide-line 471 Year 1988 GLP

Test substance

no data other TS

ld 67-68-5 Date 17.05.2004

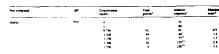
Test substance

dimethyl sulfoxide (CAS 67-68-5), purity not specified

Result

Positive at cytotoxic concentrations (>= 1.0 mol/l).

Attached document Wangenheim table 1.bmp



Source

Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability

(3) invalid

Only tested without metabolic activation

06.05.2004

Method

(153)

GENETIC TOXICITY 'IN VIVO'

Micronucleus assay Type

Species mouse Sex male Strain B6C3F1 Route of admin. i.p. Exposure period single **Doses** 5 ml/kg Result negative

Year 1989 GLP no data

Test substance as prescribed by 1.1 - 1.4

other: no data

Method

DMSO was used as solvent in micronucleus test. 3 groups of 8 male B6C3F1 mice received a single intraperitoneal injection of 5 ml/kg DMSO. Bone marrow samples were taken from a single femur of each animal in groups killed at 24, 48 and 72 hours post-injection. Micronucleated cell frequencies were determined by scoring a minimum of 1000 polychromatic erythrocytes for each animal. The ratio of polychromatic to normochromatic cells was determined and was based on the

first 100 erythrocytic cells encountered.

Source Atofina, Paris-la-Défense, France

ld 67-68-5 Date 17.05.2004

(108)

(62)

(135)

Atofina Paris La Défense Cedex

Reliability : (2) valid with restrictions

Flag : Critical study for SIDS endpoint 29.07.2003

Type : Micronucleus assay

Species : other: larvea from Pleurodeles waitl Sex :

Sex Strain

Route of admin. : other: dissolved in water of the aquariums

Exposure period : 12 days
Doses : 1100 ppm
Result : negative
Method : other
Year : 1993
GLP : no data

Test substance : as prescribed by 1.1 - 1.4

Source : Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability : (2) valid with restrictions

03.06.2003

Type : Sister chromatid exchange assay

Species : mouse
Sex : female
Strain : ICR
Route of admin. : i.p.

Exposure period : i.p. : single on day 13 of gestation

Doses : 2.5, 5.0, 10.0, 20.0 ml/kg
Result : negative

Method : other Year : 1985 GLP : no data

Test substance : as prescribed by 1.1 - 1.4

Method : Sister chromatid exchanges (SCE) and cell replication

kinetics (CRK) after maternal DMSO exposure were studied in mouse dams and fetuses. Pregnant ICR-mice had a 55 milligram

5-bromodeoxyuridine (59143) (BrdU) tablet implanted subcutaneously in the abdomen on gestation day 13. After 30 minutes to 1 hour, animals were treated i.p. with 0, 2.5, 5.0 and 10.0 ml/kg DMSO. About 21 hours after BrdU implantation, dams were injected with 80 micrograms colchicine and killed 2 to 3 hours later. Uterine horns and fetuses were removed.

Fetal livers and maternal bone marrow were prepared for cell scoring. CRK was assessed by classifying fluorescence plus Giernsa stained metaphase cells as M1, M2, or M3 plus, which indicated one, two, or three more rounds of DNA replication since BrdU treatment, respectively. Average generation time (AGT) as a function of test dose was calculated. SCE was scored as a reciprocal exchange between the chromatids of a

chromosome in M2 cells.

Result : There was a significant heterogeneity in relative number of M1, M2, and M3 plus cells among DMSO doses in maternal and

fetal cells. DMSO was classified as negative regarding SCE

induction in maternal bone marrow and fetal liver.

Source : Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability : (2) valid with restrictions

29.07.2003

ld 67-68-5 Date 17.05.2004

(36)(105)

Type : Dominant lethal assay

Species: mouseSex: maleStrain: Swiss

Route of admin. : i.p.

Exposure period : twice at an interval of 20 hr
Doses : 5000 - 7500 - 10000 mg/kg

Result : negative
Method : other: no data
Year : 1975

Year : 1979 GLP : no

Test substance : as prescribed by 1.1 - 1.4

Method : Groups of 15 male mice were injected intraperitoneally with

5, 7.5, and 10 g/kg DMSO twice at an interval of about 20 hours. Control animals received no treatment. Surviving males were paired with untreated virgin females, which were replaced at weekly intervals for five consecutive weeks. Females were killed and examined for implantion sites and dead implants at 10-11 days after separation from males. Pre-implantation loss was evaluated by comparing the number of implantation sites in females mated with DMSO-treated males to the number in females mated with untreated males. The incidence of females with dead implantations was

recorded, and pregnancy rates determined.

Result : Male mice treated with DMSO appeared sedated, and consumed

less food and water than untreated controls. These effects were dose related and were most apparent in mice that received 10 g/kg DMSO. Incidence of mortality was 7, 20, and 73% for the 5, 7.5 and 10 g/kg groups, respectively. During the first week of matings, pregnancy rates were reduced in females paired with 10 g/kg males. Rates increased in subsequent weeks, and were comparable to controls by week 5.

subsequent weeks, and were comparable to controls by week 5 Pregnancy rates of females paired with males given 5 and 7.5 g/kg were similar to controls.

Total implantation rates were reduced in females paired with 7.5 and 10 g/kg males during week 1. There were no significant differences in implantations in subsequent

weeks.

The number of dead implantations in females mated to DMSO-treated males did not differ from that of controls

during the entire test interval.

Source : Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability : (2) valid with restrictions

03.06.2003

Type : other:single-strand breaks in DNA

Species: mouseSex: maleStrain: NMRIRoute of admin.: i.p.

Exposure period : single administration

Doses : 25 to 75 mmol/kg (1950 to 5860 mg/kg)

Result : ambiguous

Method : other: Erixon and Ahnström, Mut res, 1979, 59, 257-271

Year : 1984 GLP : no data

Test substance : as prescribed by 1.1 - 1.4

Method : The method for determination of single-strand breaks (SSB)

in DNA by the technique of alkaline unwinding and

ld 67-68-5

Date 17.05.2004

hydroxylapatite chromatography has been applied for cell nuclei from organs of mice. Male mice were given DMSO by i.p. administration. Cell nuclei were prepared from various organs and then lysed in alkali. The amount of DNA was determined by fluorometry using 4',6-diamidino-2 phenylindole.2HCl. The relative level of SSB in DNA was determined in liver, kidney, lung, spleen, testis or brain,

0.5-24 h after administration of DMSO.

Result

DMSO induced SSB only in DNA of kidney, 0.5 hr after treatment with the high dose of 75 mmol/kg. No effect was observed in kidney at lower dose levels. No effect as well was obseved in the other organs, 0.5, 4 and 24 after the

administration of 0.1 mmol/kg.

Remark

The highest dose level tested (75 mmol/kg = 5460 mg/kg) is largely in exces of the OECD recommended limit dose (2000 mg/kg) for in vivo genotoxicity testing. The effects observed at this high dose are of doubtful significance.

Source

Strain

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 03.06.2003 (2) valid with restrictions

Type

Drosophila SLRL test Drosophila melanogaster

Species Sex male

other: Berlin wild males and Basc females

Route of admin. other: intra-abdominal injection

Exposure period Single dose 0.1, 1.0, 5.0% (v/v) Doses negative Result

Method other Year 1974 **GLP**

Test substance

as prescribed by 1.1 - 1.4

Method

DMSO was injected intraabdominally into 1-2-day-old males at concentrations of 0.1, 1 and 5%; the volume injected was 0.2 µI per fly. Rod-X and ring-X bearing males were used to

for sex-linked recessive lethals and for sex chromosome loss, respectively. Controls consisted of males that were not injected, and males that received saline injections. One day after treatment, each male was individually crossed with three 4-day-old virgin females. In order to collect

postmeiotic and premeiotic germ cell stages separately males were mated every two days to a new set of females. Males were mated five times to obtain broods A to E. Mortality and sterility of treated males were recorded

during the breeding program.

Result

Increased mortality was observed in all injected males; DMSO did not enhance mortality when compared to saline. Intraabdominal injection of DMSO did not induce sex-linked recessive lethals and did not raise the frequency of sex chromosome loss above the spontaneous level. Data from later broods showed lower frequencies of sex chromosome loss than

those from the first brood. This tendency was also observed

in untreated controls.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 29.07.2003 (2) valid with restrictions

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(152)

GLP

ld 67-68-5 Date 17.05.2004

(111)

Somatic mutation assay Type Species Drosophila melanogaster

Sex male/female Strain other Route of admin. oral feed **Exposure period** 2, 8, 24 hr **Doses** 1% (v/v) Result negative Method other Year 1976

Test substance as prescribed by 1.1 - 1.4

no

Method DMSO was tested in the eye mosaic test which detects

genetic recombination in adults as a result of treatment during the larval state. Mutagen-treated Drosophila larvae that are heterozygous for w/wco (white/white-coral) or hemizygous for w/co/Y will display mosaic eye spots as adults. Twin spots represent mitotic recombination, single spots represent somatic mutation as well as mitotic

recombination.

Males and females were mated and eggs collected within 4 hr. Larvae were treated at an age of 44 hr after egg deposition, for 2, 8, or 24 hours, by feeding a yeast suspension containing 1% (v/v) DMSO. Larvae were then allowed to continue development and adults were counted and collected

one to three days after emergence. Eyes were inspected in paraffin oil under a disecting microscope. Numbers and types

of spots were recorded for all mosaic eyes.

Result There was no evidence of genetic recombination in the DNA of

somatic cells of DMSO-treated Drosophila larvae tested in the eye mosaic test. A similar negative effect was seen in

Remark This result agrees with the findings of Mollet that DMSO

does not induce sex-link recessive lethals or chromosome

loss.

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions 29.07.2003

Somatic mutation assay Type **Species** Drosophila melanogaster Sex

male/female Strain other Route of admin. oral feed **Exposure period** 3 days **Doses** 12.8, 128 mM Result negative Method other Year 1993

GLP no data Test substance as prescribed by 1.1 - 1.4

Method DMSO was tested in the w/w+ eye mosaic test which detects

somatic cell recombination in adults as a result of treatment during the larval stage. The w/w+ system monitors

mosaic light spots in the eyes of adult females. Between 12 and 15 pairs of flies were allowed to mate and lay eggs in bottles on food supplemented with 12.8 mM or 128 mM DMSO. Parental flies were discarded and larval feeding with DMSO continued until hatching. Newly hatched females were removed

Result

Source

Method

ld 67-68-5 Date 17.05.2004

(149)

to fresh medium and scored 1-5 days later. Etherized flies were scored under a disecting microscope. Eye spots separated by at least four normal ommatidia were counted as independent events. A minimum of 250 flies were evaluated for each dose tested; at least two separate experiments were

conducted at the same dose levels.

There was no evidence of genetic recombination in the DNA of

somatic cells of DMSO-treated Drosophila larvae tested in the w/w+ eye mosaic test. A similar negative result was also

observed in the parallel control group.

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

(2) valid with restrictions

Reliability 03.06.2003

Type Cytogenetic assay

Species rat Sex male Strain Sprague-Dawley

Route of admin. i.p.

Exposure period 5 administration at 24-hour interval 0.05, 0.5, 2.5, 5 ml/kg Doses

Result positive Method

other Vear GLP no data Test substance other TS

Test substance Lab Grade-Lot # 35359, J.T. Baker Chemical Co.,

Phillipsburg, NJ

Forty albino rats (Sprague-Dawley CD strain from Charles River Breeding Laboratories, Ins., Wilmington, MA) received intraperitoneal (ip) injections of varying concentrations of DMSO for five consecutive days. Each group of 10 rats was administered DMSO at a volume of 5 ml/kg of body weight at

concentrations of 1%, 10%, 50%, or 100% DMSO. The DMSO was diluted in distilled water. A vehicle control group of an additional ten animals was treated with distilled water under the same conditions. All rats were 8-10 week old

On day six, each animal received an ip injection of colchicine at 2 mg/kg of body weight. Two hours later, animals were sacrificed with CO2 anesthesia followed by cervical dislocation. Immediately after sacrifice, bone marrow was aspirated from both femurs of each rat and placed into 5 ml of prewarmed (37°C) Hank's balanced salt solution. The aspirate was centrifuged for five minutes at 100g. The supernatant was removed, 3.0 ml of 0.075 M KCI

was added to each centrifuge tubes, and the tubes were allowed to stand at room temperature for 25 minutes. The cells were centrifuged again for five minutes at 100g. The supernatant was removed and 5 ml ot fixative (3:1 methanol: acetic acid) was added to each precipitate. After 20 minutes at room temperature, the cells were centrifuged as before. The tubes were decanted, and 5 ml of fixative were again added, after which the tubes were sealed and refrigerated overnight at 4°C. Following refrigeration, the cells were centrifuged, decanted, and resuspended in 1-3 ml of fresh

fixative

Three aliquots of the final suspension were dropped onto clean slides which were labeled with the animal number from each respective rat. The slides were allowed to air

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dry, and were stained for 16 minutes in a preparation of 24 ml Giernsa stock solution, 18 ml acetone, and 198 ml Harleco buffer phosphate (pH 6.8).

Dried slides were held in xylene and subsequently mounted with glass cover slips in Cover Bond mounting media. Three slides were prepared per animal.

The animal number on each slide was covered with masking tape, and each slide was assigned a slide code number. The code was not broken until all slides had been analyzed. Fifty metaphases per animal were analyzed for chromosome breaks, chromatid breaks, markers, and severely damaged cells. All data were statistically analyzed by the Wilcoxon nonparametric comparison of group means.

A break is defined as any separation that exceeds the width of the chromatid arm, or a discontinuity associated with an unaligned segment of chromosomal material. Chromatid breaks involve only one arm of the chromosome, while chromosomal breaks involve both arms at identical loci. Markers consist of exchanges, rings and dicentrics. An exchange is a shift of a segment of a chromosome to a new position on a nonhomologous chromosome, resulting in altered chromosomal configuration. A ring chromosome is one in which the ends have joined to form a circle. A dicentric chromosome is one in which two constrictions are symmetrically present on both chromatids. Severely damaged cells are of three types pulverized cells, cells in which the chromosomes are clumped together so as to render individual chromosomes indistinguishable, and cells showing more than 10 aberrations

Result

Two animals treated with 1% DMSO and one animal in both the negative control group and the group treated with 100% DMSO did not present analyzable cells. In addition, three animals in the latter group died prior to termination of the study. These events resulted in less than 500 cells analyzed in these three groups.

The percent of aberrant cells per animal increases at each dose level (Table 1). The Wilcoxon nonparametric comparison of group means was performed on the proportion of aberrant cells per animal. All groups treated with DMSO show a statistically significant elevation in chromosome aberrations when compared to the negative control group. The percent of cells per animal exhibiting each category of chromosome aberration in each group was also compared by the Wilcoxon nonparametric test. The incidence of chromosome breaks is not significantly different between the control and test groups. The level of chromatid breaks is significantly elevated in all test groups except Group 3 (10% DMSO treatment), while the incidence of markers is significantly higher in all test groups when compared to the negative control group. Only in Group 5 is the number of severely damaged cells significantly elevated. The number of severely damaged cells in animals administered 100% DMSO (Group 5) is significantly greater than in all other groups.

Attached document

: Kapp table 1.bmp

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(89)

TABLE I. Results of In Vivo Treatment of Male Rats With Varying Concentrations of DMSO and Resultant Analysis of Bone Marrow Cells

Group no.	Treatment	No. of survivors at sacrifice	No. animals presenting analyzable No. surviving cells at sacrifice	No. cells analyzed	Mean percent aberrant cells per animal	Standard deviation
ı	distilled H ₂ O	10/10	9/10	450	4.0	2.0
2	1% DMSO	10/10	8/10	400	10.0	6.05
3	10% DMSO	10/10	10/10	500	18.0	13.76
4	50% DMSQ	10/10	10/10	500	19.2	6.68
5	100% DMSO	7/10	6/7	300	68.67	21.45

TABLE II. Results of Wilcoxon Nonparametric Comparison of Group Means for Various Chromo Aberrations in Bone Marrow Cells

Groups compared	Total chromosome aberrations	Chromatid breaks	Chromosome breaks	Markers	Severely damaged cells
1 and 2	s	s	NS	S	NS
1 and 3	S	NS	NS	S	NS
l and 4	S	S	NS	S	NS
1 and 5	S	8	NS	S	S

Group 1, vehicle control (distilled H₂O); Group 2, 1% DMSO; Group 3, 10% DMSO; Group 4, 50% DMSO; Group 5, 100% DMSO.

NS, No statistically significant difference between the means of the groups.

S, Statistically significant difference between the means of the groups at the P < 0.05 level. (Increased over negative control means in Group 1.)

Source Reliability : Atofina Paris La Défense Cedex

: (2) valid with restrictions

14.05.2004

other: numerical chromosome aberration study

Type Species Drosophila melanogaster

Sex female Strain other Route of admin. oral feed **Exposure period** 3 days Doses : 2% Result negative

Method other Year 1983 **GLP** no data

Test substance as prescribed by 1.1 - 1.4

Method

DMSO was fed as a 2% solution to virgin y/y female flies for 3 days. Females were then transferred to regular medium and mated with males to determined the incidence of aneuploidy using the conventional or aneuploidy pattern method. Male and female flies were provided with fresh food after 4 days and removed from food after 6 days. In this way, a (4 + 6) brood pattern was obtained in each of the two DMSO experiments and controls.

-Conventional method: y/y females fed DMSO were mated to normal males (Berlin wild). F1 flies with X-chromosome gain (XXY females) and X-chromosome loss (XO males) are visually

distinguished from normal offspring.

-Aneuploidy pattern method: y/y females fed DMSO were mated to C(2L)RM,b;C(2R)RM,vg males. F1 survivors are flies

derived from oocytes that aneuploid with respect to chromosome II, eventually in combination with a gain or loss of chromosome I. The number of flies obtained from each

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(146)

brood is examined, and the pattern compared to that obtained

Both the conventional and aneuploidy pattern methods failed Result

to produce evidence that DMSO induces aneuploidy in oocytes

of Drosophila melanogaster.

DMSO has been consistently negative in Drosophila studies Remark

using male germ cells for sex-linked recessive lethals, dominant lethals, somatic mutations, translocations, and ring-X losses. These results demonstrate that DMSO can be used unhesitatingly as a solvent for chemical agents in

mutagen screening in Drosophila.

Atofina, Paris-la-Défense, France Source

Atofina Paris La Défense Cedex

(2) valid with restrictions Reliability

03.06.2003

other: Micronucleus assay in bone marrow Type

Species mouse male Sex Strain Swiss Route of admin. i.p. Exposure period single

Doses 2 g/kg (as solvent vehicle)

Result negative

Method other: Schmid, 1976

Year 1977 GLP no

Test substance as prescribed by 1.1 - 1.4

Method DMSO was given ip 30 and 6 h to 3 mice before killing. For

each mouse, 4 slides were prepared and an average of 4000

polychromatic erythrocytes and the corresponding normochromatic erythrocytes were scored.

DMSO did not affect the incidence of micronuclei, but Result

reduced the P-N ratio slightly (p>0.1).

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (3) invalid

Only 3 mice treated

(35)03.06.2003

other: Host mediated assay Type

Species other: ex vivo in Rodent with Salmonella typhimurium

Sex no data

Strain

Route of admin. other: N/A see remark

Exposure period see remark

Doses

Result negative Method other: no data

Year 1976 GLP no

Test substance as prescribed by 1.1 - 1.4

Remark This paper is a compilation of data without experimental

details.

Atofina, Paris-la-Défense, France Source Atofina Paris La Défense Cedex

Reliability (4) not assignable

03.06.2003 (96)

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CARCINOGENICITY 5.7

Species

mouse

Sex

female

Strain

other: ICR/Ha swiss S.C.

Route of admin. Exposure period Frequency of treatm.

76 weeks once a week

Post exposure period

none

Doses Result 0.05 ml/mouse (ca. 1.8 g/kg bw)

Control group

negative

Method

other: 2 groups of 30 mice receiving physiologic saline or water and 120 mice without treatment

other 1971

no

Year GLP

Test substance

as prescribed by 1.1 - 1.4

Method

The mice (30) were given weekly sc injection in the left flank of 0.05 ml DMSO. The animals were examined regularly and the finding recorded once a month. The test was continued until there were no survivors. All animals were autopsied, abnormal tissues and tumors were excised and

examined histologically.

Result

Median survival time (>76 weeks) was not affected compared to control groups. No benign or malignant tumor was observed

at the injection site.

Source

Atofina, Paris-le-Défense, France. Atofina Paris La Défense Cedex

Reliability

13.03.2001

(3) invalid

(147)

(139)

Species rat

Sex male/female

Strain other: wistar adrenal Route of admin. short term assays

Exposure period Frequency of treatm. Post exposure period

about number of mitosis

Doses Result 2.5 and 5 ml/rat

Control group Method

other: no data

Year **GLP**

no data

Test substance

as prescribed by 1.1 - 1.4

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 24.12.2002 (3) invalid

Species mouse

Sex male/female Strain other: C3H ventral prostate

Route of admin.

Exposure period Frequency of treatm. Post exposure period

short term assays about number of mitosis

Doses

0.5 %

Result

ld 67-68-5 Date 17.05.2004

(139)

(139)

(139)

Control group

Method

Year GLP

other: no data

Test substance

as prescribed by 1.1 - 1.4

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability

(3) invalid

24.12.2002

Species Sex Strain

hamster male/female other: Chinese V79

Route of admin. Exposure period

short term assays Frequency of treatm. about number of mitosis Post exposure period

Doses

10 mM Result

Control group Method

other: no data

Year GLP

no data

Test substance

as prescribed by 1.1 - 1.4

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

as prescribed by 1.1 - 1.4

Reliability

(3) invalid 24.12.2002

Species Syrian hamster

Sex no data Strain other: syrian (embryo)

Route of admin. **Exposure period** Frequency of treatm.

short term assay

Post exposure period **Doses**

Test substance

Concentration 2 % in cell culture embryo or of sternal hyaline cartilage Result

Control group no data specified Method other: no data

Year GLP no data

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex Reliability (3) invalid

24.12.2002

5.8.1 TOXICITY TO FERTILITY

Type other: reproductive organs toxicity

Species rat

male/female Sex Strain Sprague-Dawley Route of admin. inhalation **Exposure period** 13 weeks

Frequency of treatm. 6 hours/day, 7 days/week

ld 67-68-5 Date 17.05.2004

(52)

(32)

Premating exposure period

Male

Female

Duration of test 13 weeks

No. of generation

studies Doses

0.310, 0.964 and 2.783 mg/l yes, concurrent vehicle

Control group **NOAEL** parental = 2.783 mg/l

Method other: OECD guide-line 413 Year 2000

GLP yes

as prescribed by 1.1 - 1.4 Test substance

Method During the 90-day inhalation toxicity study reported in

section 5.4, the oestrus cycle of female rats was monitored, male rats were subjected to sperm investigations (count, motility and morphology) and the reproductive organs of both sexes were examined histologically.

No treatment related effects were observed. Result

Atofina, Paris-le-Défense, France. Source Atofina Paris La Défense Cedex

Reliability (1) valid without restriction Critical study for SIDS endpoint Flag

29.07.2003

Type Fertility **Species** rat male/female Sex

Strain Sprague-Dawley Route of admin.

gavage 4 days before coïtus **Exposure period**

Frequency of treatm. daily

Premating exposure period Male 4 days

Female 4 days **Duration of test** until weaning

No. of generation

studies

Strain

Doses 5 g/kg

Control group no data specified **NOAEL** parental = 5000 mg/kg bw NOAEL F1 offspring = 5000 mg/kg bw Method other

Year 1964 GLP no

Test substance as prescribed by 1.1 - 1.4

Result Oral doses of 5 g/kg DMSO administered to male and female

rats for 4 days before coïtus did not affect fertility.

Female rats treated orally with DMSO throughout gestation delivered normally and the offspring developed normally.

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex Source

Reliability (3) invalid

24.12.2002

Fertility Type Species mouse Sex female

Swiss Route of admin. other: intraperitoneal or oral (gavage)

ld 67-68-5 Date 17.05.2004

Exposure period: Different periods of administration during pregnancy (see method)

Frequency of treatm. : see method

Premating exposure period

Male

Female : none

Duration of test

Control group

Doses

No. of generation studies

: 0.5 ml/kg/g : no

NOAEL parental : = 275 mg/kg bw

 Method
 : other

 Year
 : 1982

 GLP
 : no data

Test substance : other TS: 50/50 mixture of DMSO with 95% ethanol

Method : Adult Swiss female mice (25-30 g) were mated with proven Swiss male mice. 0.5 ml/kg/d of a mixture DMSO/95% ethanol

50/50 (v/v) was administered i.p. or p.o. to several groups of pregnant mice as follow:

1. from day 2 through 5 of pregnancy. The laparotomy was performed on day 7 of pregnancy.

2. from day 8 through 12 of pregnancy. The laparotomy was

performed on day 18 of pregnancy.

Result : 1. No inhibition of implantation was observed in mice given

an i.p. or p.o dose of 0.5 ml/kg/d of the mixture from GD2

to GD5.

2. No abortifacient effect was observed in pregnant mice given an i.p. or p.o dose of 0.5 ml/kg/d of the mixture from

GD8 to GD12.

Source : Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability : (3) invalid

29.07.2003 (122)

5.8.2 DEVELOPMENTAL TOXICITY/TERATOGENICITY

Species : rat Sex : female

Strain : Sprague-Dawley

Route of admin. : gavage

Exposure period : 10 days, days 6-15 of gestation Frequency of treatm. : daily

Duration of test

Doses

Control group

NOAEL maternal tox.

NOAEL teratogen.

Sacrifice on day 20 of gestation

200 - 1000 - 5000 mg/kg

yes, concurrent vehicle

1 = 1000 mg/kg bw

1 = 1000 mg/kg bw

Method : OECD Guide-line 414 "Teratogenicity"

Year : 1997
GLP : yes
Test substance : other TS

Test substance : dimethylsulfoxide, CAS no.: 67-68-5

Source: Elf Aquitaine Production

Purity: 99.89%

Method : Three groups of 25 mated female rats received DMSO by gavage

ld 67-68-5

Date 17.05.2004

(55)

at the dose levels of 200, 1000, and 5000 mg/kg/day as a solution in purified water. DMSO was administered each day from day 6 to day 15 of gestation. A control group of 25 mated females was given the vehicle alone. Day 0 of pregnancy was designated as the day of confirmed mating. Clinical signs including mortality and evidence of abortion were checked daily. Food consumption and body weight were recorded at designated intervals during pregnancy. On day 20 of pregnancy, females were killed. The gravid uterus was weighed and fetuses removed by hysterectormy. Females were examined macroscopically. Litter parameters were recorded: number of corpora lutea, implantation sites, early and late responptions, and dead and live fetuses. Fetuses were weighed, sexed, and submitted to external examination and then to soft tissue or skeletal examinations

Result

Maternal data: There were no clinical signs observed in treated or controlgroups. No maternal deaths or abortions occurred in any group. Lower food consumption (-11% compared to control) was noted in females of the 5000 mg/kg group during the treatment period. The body weight gain of the gravid females given 5000 mg/kg/d was slightly lower that of controls, during the treatment period: 41 g vs. 60 g (i.e. -32%, p<0.001); this resulted in mean absolute body weight on day 20 of pregnancy lower by -4% (365g vs. 382g) compared to controls. No macroscopic findings were noted at necropsy in any of the females.

Litter data

Pre- and post-implantation losses were similar in all groups. No treatment-related effects were observed on the number of fetuses or the sex ratio. In the 5000 mg/kg/day group, fetal body weights were slightly lower than that of controls (3.54g vs 3.79 g (-7%, p< 0.05), an indirect consequence, at least in part, of decreased maternal food consumption and body weight gain.

Fetal examination:

No external malformations or anomalies were observed in fetuses from any group. An increased incidence of two soft tissues anomalies were observed: dilated renal pelvis for fetuses in all treated groups, which was associated at 5000 mg/kg/day with an increased incidence of dilated ureter(s). No treatment-related soft tissue malformations were observed. There were no treatment-related skeletal variations or malformations in any group. An increased incidence of reduced or delayed ossification of ribs was observed in fetuses of the 5000 mg/kg group. This skeletal anomaly is considered to be a consequence of the lowered fetal body weights observed for this group. No treatment-related microscopic changes were noted in the kidneys of fetuses with dilated renal pelvis. Dilation of the renal pelvis may be related to the diuretic properties of DMSO

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability Flag : (1) valid without restriction: Critical study for SIDS endpoint

14.05.2004

Species: ratSex: femaleStrain: Sprague-Dawley

Route of admin. Exposure period

gavage10 days, days 6-15 of gestation

ld 67-68-5 Date 17.05.2004

Frequency of treatm.

Duration of test Doses

Control group
NOAEL maternal tox.

NOAEL maternal tox. NOAEL teratogen.

Method Year

GLP Test substance daily

sacrifice on day 20 of gestation 1000 - 5000 - 10,000 mg/kg yes, concurrent vehicle = 1000 mg/kg bw

: = 1000 mg/kg bw

other: range-finding study according to OECD guideline 414

: 1996 : ves

st substance : other TS

Test substance

Result

dimethyl sulfoxide, CAS no.: 67-68-5

Source: Elf Aquitaine Production

Purity: 99.89%

Method : Three groups of seven mated female rats received DMSO by

gavage at the dose levels of 1000, 5000, and 10,000 mg/kg/day as a solution in purified water. A constant volume dosage of 10 ml/kg was used for each group. DMSO was administered each day from day 6 to day 15 of gestation. A control group of seven mated females was given the vehicle alone. Day 0 of pregnancy was designated as the day of

confirmed mating.

Clinical signs including mortality and evidence of abortion were checked daily. Food consumption and body weight were

recorded at designated intervals during pregnancy.
On day 20 of pregnancy, females were killed, examined macroscopically and fetuses removed by Caesarean section. Litter parameters were recorded: number of corpora lutea, implantation sites, resorptions, and dead and live fetuses. Fetuses were weighed, sexed, and submitted to external examination.

examination. : Maternal data:

There were no clinical signs observed in treated or control groups. No maternal deaths or abortions occured in any

group.

Lower food consumption (-14%, -21%) and body weight gain (-32%, -50%) were noted during treatment (day 6-15 of pregnancy) in females of the 5000 mg/kg group, and throughout the gestation period in females administered 10,000 mg/kg DMSO, respectively. No macroscopic findings were noted at necropsy in any of the females of the 1000, 5000, or 10,000

mg/kg DMSO groups.

Litter data:

The mean number of corpora lutea and implantation sites per animal showed some variations between control and treated groups; these differences were not dose related and could not be ascribed to treatment with DMSO. No late resorptions or dead fetuses were noted in any group. Higher rates of early resorptions per animal, and higher total post implantation loss were observed in the 5000 and 10000 mg/kg groups. The mean number of live fetuses per animal presented variations between control and treated groups; these differences were not dose-related and could not be ascribed to treatment with DMSO. A treatment-related decrease in the rate of live fetuses was slightly lower in the 5000 and 10000 mg/kg groups. Slight to moderately lower fetal body weights were noted in the 5000 and 10000 mg/kg groups, in line with the treatment-related effect on maternal food consumption and body weight gain. The sex ratio was similar in control and treated groups

Fetal examination:

ld 67-68-5 Date 17.05.2004

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No external anomalies or malformations were observed in

fetuses from any group.

Atofina, Paris-la-Défense, France Source

Atofina Paris La Défense Cedex

Reliability 07.05.2004 (2) valid with restrictions

Species rat Sex female Strain Wistar Route of admin. gavage

d6 to d12 of gestation Exposure period

Frequency of treatm. daily

Duration of test Sacrifice 1 to 3 days before parturition 5000 and 10000 mg/kg (as a 50% solution) yes, concurrent vehicle **Doses**

Control group NOAEL teratogen. = 10000 mg/kg bw Method other: no data

Year 1967 GLP no

Test substance as prescribed by 1.1 - 1.4

Method Pregnant rats were administered oral doses of 5 and 10

g/kg/d DMSO on days 6 to 12 of gestation. They were sacrificed 1 to 3 days before parturition. When obtained, the fetuses were weighted, and examined grossly before and

after fixing in Bouin's solution.

Result No effects was observed on the number of females going full

term, the number of aborted embryos and the number of

malformations.

Atofina, Paris-la-Défense, France Source

Atofina Paris La Défense Cedex

(3) invalid

This study was performed in the early 60'. Protocols of teratogenic studies were not validated. Insufficient number of animal, without S.P.F. status, doubtful breeding (feeding - environmental status) were used. In these experimental conditions, it is difficult to find out the precise results in the papers, and it is impossible to interpretate (no statistical evaluation on unformal findings). These studies

are not valid to assess the teratogenic potential of DMSO.

29.07.2003

Reliability

(31)

Species rat Sex female Strain no data Route of admin.

6-15 post coitus **Exposure period** Frequency of treatm. daily no data

Duration of test

Doses 2 ml/kg for GD 6 to 15; 2, 4 or 8 ml/kg once or 3 times on GD 5, 7, 9, 11,

13, 15, or 17 no data specified

Control group NOAEL maternal tox. < 2000 mg/kg bw Method other: no data 1977 Year

no

Test substance as prescribed by 1.1 - 1.4

Result When DMSO was administered i.p. to pregnant rats at daily

dose of 2 ml/kg for days 6 to 15 of gestation, 2 of 9 dams died and all the fetuses were resorbed completely in the

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(31)

others. When DMSO (2, 4 or 8 ml/kg was administered i.p. either once or 3 times on day 5, 7, 9, 11, 13, 15, or 17 of gestation, increased resorption of foetuses was observed following a dose of 8 ml/kg on day 17, or when DMSO was

administered three times on days 7, 11, or 15. Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

(3) invalid Reliability

This study was performed in the 70'. Protocols of teratogenic studies were not validated. Insufficient number of animal, without S.P.F. status, doubtful breeding (feeding - environmental status) were used. In these experimental conditions, it is difficult to find out the precise results in the papers, and it is impossible to interpretate (no statistical evaluation on unformal findings). These studies

are not valid to assess the teratogenic potential of DMSO.

29.07.2003

(151)

Species rat female Sex Strain Wistar Route of admin. i.p.

Exposure period d6 to d12 of gestation

Frequency of treatm.

Duration of test daily

sacrifice 1 to 3 days before parturition

Doses 5, 8 and 10 g/kg Control group yes, concurrent vehicle

Method other: no data Year 1967

GLP no

as prescribed by 1.1 - 1.4 Test substance

A dose of 10 g/kg DMSO caused the death of 4 of 14 parent Result

rats and the intraperitoneal injection of 8 to 10 g/kg caused an increase in the number of aborted foetuses, and a reduction in the weight of live foetuses. Eleven out of 729 live foetuses from the rats receiving intraperitoneal doses of DMSO exhibited malformations (of the abdominal wall, CNS,

limbs, jaw and tail), in comparison with 1 out of 558

control foetuses. Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability (3) invalid

This study was performed in the early 60'. Protocols of teratogenic studies were not validated. Insufficient number of animal, without S.P.F. status, doubtful breeding (feeding - environmental status) were used. In these experimental conditions, it is difficult to find out the precise results in the papers, and it is impossible to interpretate (no statistical evaluation on unformal findings). These studies

are not valid to assess the teratogenic potential of DMSO.

24.12.2002

Species mouse female Sex Strain ICR Route of admin. dermal **Exposure period** D9

Frequency of treatm. single day 9

Duration of test Day 10 or complete gestation Doses concentration in distilled water: 0.04 - 0.4 - 4 %

Control group yes, concurrent vehicle 121 / 157

Remark

ld 67-68-5 Date 17.05.2004

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(31)

Method other: no data Year 1988 GLP no data Test substance

Result

High frequencies of embryo damages were observed in all DMSO

treated group on day 10 of gestation. Litter size

determined at birth decreased in the DMSO treated mice. The dermal application used in this study is not the one

recommanded by current guidelines. The lower right appendage

of the mice was dipped for 20 seconds into a solution of

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability

(3) invalid 29.07.2003

Species mouse female Sex Strain **Swiss**

Route of admin. i.p. d6 to d12 gestation **Exposure period**

Frequency of treatm.

daily

Duration of test sacrifice 1 to 3 days before parturition Doses 5, 8, 10 and 12 g/kg

Control group yes, concurrent vehicle other: no data

Method 1967 Year

GLP no

Test substance as prescribed by 1.1 - 1.4

Pregnant mice were administered daily with intraperitoneal Remark

doses of 5 to 12 g/kg DMSO on days 6 to 12 of gestation. Following intraperitoneal administration, 7 out of 100 foetuses showed malformations (4% malformed limbs, 2% anencephalia, 1 % celosomia), compared with 4 malformations

out of 1768 control foetuses.

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (3) invalid

This study was performed in the early 60'. Protocols of teratogenic studies were not validated. Insufficient number of animal, without S.P.F. status, doubtful breeding (feeding - environmental status) were used. In these experimental conditions, it is difficult to find out the precise results in the papers, and it is impossible to interpretate (no statistical evaluation on unformal findings). These studies

are not valid to assess the teratogenic potential of DMSO.

24.12.2002

Species rat Sex female

Strain Sprague-Dawley

Route of admin.

Exposure period d8, d8 to d9; d8 to d10 of gestation

Frequency of treatm. **Duration of test**

1 - 2 or 3 days 10250 mg/kg Doses

yes, concurrent no treatment < 10250 mg/kg bw Control group NOAEL teratogen.

Method other: no data

1967

ld 67-68-5 Date 17.05.2004

GLP

no

Test substance

as prescribed by 1.1 - 1.4

Remark

Subcutaneous injection of 31 rats with 10.25 g DMSO/kg/d on day 8, 8 and 9, or 8-10 of gestation did not significantly influence the body weight gain of the mothers during pregnancy or of the live young obtained on day 19. DMSO administered on days 8 to 10 of gestation reduced the average number of fetuses per litter and increased the number of resorptions. No gross or skeletal malformations were observed in any of the live fetuses obtained from the

DMSO-treated rats.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability

(3) invalid

This study was performed in the early 60'. Protocols of teratogenic studies were not validated. Insufficient number of animal, without S.P.F. status, doubtful breeding (feeding - environmental status) were used. In these experimental conditions, it is difficult to find out the precise results in the papers, and it is impossible to interpretate (no statistical evaluation on unformal findings). These studies are not valid to assess the teratogenic potential of DMSO.

29.07.2003

(88)

Species mouse Sex female Strain **Swiss** Route of admin. gavage

Exposure period

d6 to d12 of gestation daily

Frequency of treatm. **Duration of test**

Sacrifice 1 to 3 days before parturition

Doses 5 - 8 - 10 - 12 g/kg Control group yes, concurrent vehicle NOAEL teratogen. = 12000 mg/kg bw Method other: no data Year

1967 GLP

Test substance as prescribed by 1.1 - 1.4

Method Pregnant mice were administered oral doses of 5, 8, 10 and

12 g/kg/d DMSO on days 6 to 12 of gestation, respectively. They were sacrificed 1 to 3 days before parturition. When obtained, the foetuses were weighted, and examined grossly

before and after fixing in Bouin's solution.

Result No effects was observed on the number of females going full

term, the number of aborted embryos and the number of

malformations.

Remark Due to many uncertainties on the protocol and the results

the absence of statistical analysis and the use of non relevant route of administration and dose levels, this study is not valid to assess the teratogenic potential of

DMŚO.

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (3) invalid

This study was performed in the early 60'. Protocols of teratogenic studies were not validated. Insufficient number of animal, without S.P.F. status, doubtful breeding (feeding - environmental status) were used. In these experimental conditions, it is difficult to find out the precise results in the papers, and it is impossible to interpretate (no

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statistical evaluation on unformal findings). These studies are not valid to assess the teratogenic potential of DMSO.

24.12.2002

(31)

Species hamster Sex female

Strain other: golden Route of admin. i.p. **Exposure period** d8 of gestation

Frequency of treatm. single **Duration of test** 1 d

Doses 0.5 ml per animal (undiluted)

Control group no data specified NOAEL maternal tox. < 5500 mg/kg bw NOAEL teratogen. < 5500 mg/kg bw Method other: no data

Year 1966 GLP no

Test substance as prescribed by 1.1 - 1.4

Result

Pregnant hamsters were administered a single intraperitoneal injection of 0.5 ml undiluted DMSO on day 8 of gestation and the embryos were examined 1 to 3 days later. DMSO was found to be embryotoxic, the embryocidal effect being most marked

in litters from mothers weighing less than 110 g at the time of injection. Various degrees of

excencephaly and anencephaly were found in those embryos

surviving up to 3 days after the injection. Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (3) invalid

This study was performed in the early 60'. Protocols of teratogenic studies were not validated. Insufficient number of animal, without S.P.F. status, doubtful breeding (feeding - environmental status) were used. In these experimental conditions, it is difficult to find out the precise results in the papers, and it is impossible to interpretate (no statistical evaluation on unformal findings). These studies

are not valid to assess the teratogenic potential of DMSO.

29.07.2003

(61)

Species : hamster Strain no data

other: intravenous at doses of 50 to 5500 mg/kg or intraperitoneal at doses Route of admin.

of 5500 and 8250 mg/kg on day 8 of gestation

Exposure period Frequency of treatm. single administration **Duration of test** 3 days

Doses 50 to 8250 mg/kg Control group yes, historical

NOAEL maternal tox. = 2500 mg/kg bw NOAEL teratogen. = 2500 mg/kg bw Method other

Year 1966 GLP no

Test substance as prescribed by 1.1 - 1.4

Method Groups of 5-6 pregnant golden hamsters were injected with

dilutions of DMSO ranging from 50 to 5500 mg/kg iv or 5500

and 8250 mg/kg ip on the eighth day of gestation. Result

Examination of the embryos 3 days later revealed that no

Source

Id 67-68-5 Date 17.05.2004

(60)

embryocidal or teratogenic effects were noted until levels of 2500 mg/kg were reached. At higher levels, malformations, including exencephaly, rib fusions, microphthalmia, limb abnormalities and cleft lip were found. There was no appreciable effect of DMSO on maternal weight gain or

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

(3) invalid

Reliability 24.12.2002

Species rabbit Sex female Strain other Route of admin. S.C.

Exposure period 6 to 14d of gestation

Frequency of treatm. daily

Duration of test GD 20 to GD 24

Doses 4 g/kg

Control group yes, concurrent vehicle NOAEL teratogen. = 4000 mg/kg bw Method other: no data Year 1967

GLP

Test substance

Remark Five pregnant rabbits received a daily subcutaneous dose of

g/kg DMSO on days 6 to 14 of gestation. There were no

adverse effects upon fetal weight or numbers.

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (3) invalid

This study was performed in the early 60'. Protocols of teratogenic studies were not validated. Insufficient number of animal, without S.P.F. status, doubtful breeding (feeding - environmental status) were used. In these experimental conditions, it is difficult to find out the precise results in the papers, and it is impossible to interpretate (no statistical evaluation on unformal findings). These studies

are not valid to assess the teratogenic potential of DMSO.

29.07.2003

(31)

Species rabbit Sex female Strain other Route of admin. gavage

Exposure period day 6 to 14 of gestation

Frequency of treatm. daily

Duration of test sacrifice on GD 20 to GD 24 **Doses**

5 g/kg

Control group yes, concurrent vehicle NOAEL teratogen. = 5000 mg/kg bw

Method other: no data Year **GLP** no data

Test substance as prescribed by 1.1 - 1.4

Method 10 pregnant rabbits received a daily oral dose of 5 g/kg

(as a 50% solution) on days 6 to 14 of gestation. the

rabbits were sacrificed on GD20 to 24.

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(31)

Result There were no adverse effects upon fetal weight, numbers or

malformations.

Remark Due to many uncertainties on the protocol and the results

this study is not valid to assess the teratogenic potential

of DMSO.

Source Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (3) invalid

> This study was performed in the early 60'. Protocols of teratogenic studies were not validated. Insufficient number of animal, without S.P.F. status, doubtful breeding (feeding - environmental status) were used. In these experimental conditions, it is difficult to find out the precise results in the papers, and it is impossible to interpretate (no

statistical evaluation on unformal findings). These studies are not valid to assess the teratogenic potential of DMSO.

29.07.2003

Species hen

Sex Strain Leghorn Route of admin. other: injection **Exposure period** day 3 of incubation

Frequency of treatm. single between stages 17 and 23

Duration of test eggs are allowed to grow to day 10 of incubation

Doses 1µ1 90 % DMSO yes, concurrent no treatment

Control group Method other: no data Year 1987 GLP no data

Test substance as prescribed by 1.1 - 1.4

Result Mortality was not significantly different between

DMSO-treated and untreated embryos. Statistical analysis and histological data suggest that scapular and vertebral

defects were caused by DMSO induced damage to somites. Source

Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex (3) invalid Reliability

This positive result is questionable due to the non

validated model used.

24.12.2002 (94)

Species hen Sex no data

Strain other: Rhode island Route of admin. other: injection **Exposure** period days 3 or 4 of incubation single

Frequency of treatm.

Duration of test Doses

solutions containing 50 % DMSO

Control group yes, concurrent vehicle

Method other Year 1965 GLP no

Test substance as prescribed by 1.1 - 1.4

Positive result is questionnable due to the non validated Result

model and the high concentration used. Source

Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (3) invalid

Id 67-68-5

Date 17.05.2004

This study was performed in the early 60'. Protocols of teratogenic studies were not validated. Insufficient number of animal, without S.P.F. status, doubtful breeding (feeding - environmental status) were used. In these experimental conditions, it is difficult to find out the precise results in the papers, and it is impossible to interpretate (no statistical evaluation on unformal findings). These studies are not valid to assess the teratogenic potential of DMSO.

24.12.2002

(33)

(33)

Species rabbit Sex female Strain other Route of admin. S.C.

Exposure period whole pregnancy

Frequency of treatm. daily

Duration of test

Doses 2000 and 4000 mg/kg Control group NOAEL teratogen. yes, concurrent no treatment

= 4000 mg/kg bw Method other: no data

1965 GLP

Test substance as prescribed by 1.1 - 1.4

Atofina, Paris-la-Défense, France Source Atofina Paris La Défense Cedex

Reliability (3) invalid

This study was performed in the early 60'. Protocols of teratogenic studies were not validated. Insufficient number of animal, without S.P.F. status, doubtful breeding (feeding - environmental status) were used. In these experimental conditions, it is difficult to find out the precise results in the papers, and it is impossible to interpretate (no statistical evaluation on unformal findings). These studies

are not valid to assess the teratogenic potential of DMSO. 24.12.2002

other: Xenopus laevis (South African clawed frog)

Species Sex male/female

Strain other: FETAX assay Route of admin. other: immersion

Exposure period 96 hr Frequency of treatm. continuous **Duration of test** 96 hr

.25, .5, .75. 1.0, 1.25. 1.5, 1.75, 2.0 % (v/v) Doses

Control group yes, concurrent no treatment

NOAEL teratogen.

Method other: FETAX assay Year 1992

GLP no data

as prescribed by 1.1 - 1.4 Test substance

Method The Frog Embryo Teratogenesis Assay-Xenopus (FETAX) assay

was used to assess the teratogenic potential of DMSO. Embryos of the South African clawed frog were exposed for 96 hr to DMSO at dose levels of 0.25, 0.5, 0.75, 1.0, 1.25, 1.5, and 1.75% (v/v). Exposure groups of embryos were maintained using a static renewal system in which exposure media was changed at 24-hr intervals. Control embryos were reared in FETAX solution under identical environmental

conditions. A total of 200 embryos were tested per

Result

Remark

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concentration. Survival was monitored at 24-hr intervals. Length, as an indicator of growth effects, and developmental malformations were determined at the end of the 96-hr assay. The 96hr LC50, 96 hr EC50 for malformations, and the NOEL

for mortality, malformation, and growth were also

determined.

There was a marked increase in embryo mortality at the top dose concentration of 2.0%. The LC50 for replicate experiments was 1.92%. A dose-dependent increase in malformations was observed. The EC50 for malformations was 1.57%. Malformations observed included skeletal anomalies,

ocular abnormalities, and gut abnormalities. The frequency of these malformations increased with increasing

concentrations. Abnormal swimming behavior, characterized by

swimming in a spiral, and decreased growth, were also

observed at concentrations at or greater than 1.0%.

Xenopus embryos were able to survive and develop normally in relatively high concentrations of DMSO. The estimated NOELs

are 1.0% for malformations, 1.50 - 1.75% for mortality, and 1.0 - 1.25% for growth. The estimated TIs (teratogenic index) for DMSO of 1.20 - 1.24 indicate that DMSO should be designated as having a low teratogenic potential in Xenopus. This classification is comparable to mammalian and avian

studies which concluded that DMSO is generally

non-teratogenic.

Atofina, Paris-la-Défense, France

Atofina Paris La Défense Cedex

Reliability (2) valid with restrictions

24.12.2002

Source

(48)

5.8.3 TOXICITY TO REPRODUCTION, OTHER STUDIES

5.9 SPECIFIC INVESTIGATIONS

EXPOSURE EXPERIENCE

Type of experience

: Human

Method

The absorption, excretion and metabolism of DMSO have been studied in man by gas chromatography and radiometric techniques. DMSO was administered orally at a dose of 1g/kg as a 70% aqueous solution; a dose of 1 g/kg was administered

Remark

DMSO was readily absorbed when administered dermally, peak serum levels occurring after 4-8 hr. Orally administered DMSO was rapidly absorbed, reaching a peak serum level in 4 hr. Serum levels of DMSO were undetectable after 120 hr. Both unchanged DMSO and a metabolite, dimethylsulfone (DMSO2) were isolated from urine. Dimethylsulfone appeared in serum after about 48 hr and persisted for as long as 400 hr. Urinary excretion of DMSO after dermal and oral administration amounted to approximately 13% and 30-68% of the dose, respectively. Excretion of DMSO2 was about 5 to 10% and 21 to 23%, respectively. Data for subjects given DMSO chronically are also presented in this publication, as is evidence that the fraction of DMSO excreted is entirely

accounted for by unchanged DMSO and DMSO2.

ld 67-68-5 5. Toxicity Date 17.05.2004 Atofina, Paris-la-Défense, France Source Atofina Paris La Défense Cedex Reliability (2) valid with restrictions 27.12.2002 (80)Type of experience : Human : In man, DMSO is oxidized into dimethylsulfone DMSO2, Remark metabolite excreted by urine (17-22 %). DMSO is reduced into dimethylsulfide, DMS, a volatile metabolite, responsible for garlic odour of exhaled air (1 %). About 85 % is excreted unchanged, both by urine (50 %) and feces (50 %). The half time for elimination in volunteers given 1000 mg/kg intravenously is 4 days. By oral route, the same dose is excreted about 51 % as DMSO and about 10 % DMSO2 in urine within 120 hrs. Source Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex Reliability (4) not assignable (123)24.12.2002 Type of experience Human Method Plasma concentrations of DMSO, dimethylsulfone (DMSO2), and dimethylsufide (DMS) were assessed in 10 patients who underwent autologous transplants with stem cells, cryopreserved in 10% DMSO (vol/vol). Blood was sampled at multiple times after the stem-cell infusion. Urine was pooled during the 24 hours postinfusion. DMSO, DMSO2, and DMS were assayed simultaneously by gas chromatography. A one-compartment model with saturable elimination proved most suitable for fitting plasma DMSO concentration versus time Result Stem-cell volumes infused ranged between 180 and 585 ml. (254 to 824 mmol DMSO). Infusions lasted between 20 and 120 minutes. Peak plasma DMSO concentrations were 19.1 ± 6.3 mmol/L. Pharmacokinetic parameters for volume of the central compartment (Vc), maximum velocity (Vmax), and Michaels-Menten constant (Km) were 37.3 \pm 17 L, 0.99 \pm 0.57 mmol/L/h, and 5.2 ± 5.0 mmol/L, respectively. Plasma DMS02 concentrations increased during the first 24 hours, plateaued at 4.4 ± 1.2 mmol/L, and remained there until 48 hours (the last sample). DMS concentrations were at steady-state by 5 minutes and remained between 3 and 5 mmol/L for 48 hours. Urinary excretion of DMSO and DMS02 accounted for 44% ± 4% and 4% ± 1%, respectively, of the administered DMSO dose. Renal clearance of DMSO was 14.1± 3.4 ml/min. Source Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex Reliability (2) valid with restrictions 29.07.2003 (50)Type of experience : Human Adverse effect: dermal and ocular application Remark In humans, topical and intradermal application of DMSO produced garlic breath, mast cell degranulation, an increase in polymorphonuclear leukocytes and perivascular eosinophils, itching, and histamine mediated and non histamine dependent whealing and erythematous flare. Two drops of >50% DMSO in the eye caused a temporary burning

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sensation and vasodilatation; concentrations of <50%

exhibited no effects.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 24.12.2002 (4) not assignable

(157)

Type of experience

: Human

Remark

Adverse effect: skin irritation

Dermal exposure to DMSO causes skin reactions, erythema and pruritis, which appear immediately after contact with the undiluted substance; 70% solutions are usually tolerated without symptoms. In very sensitive individuals, however, reactions have been seen after contact with 10% solutions. The skin reaction to the undiluted substance is ascribed to the hygroscopic properties of DMSO, on the one hand, and to

the exothermic dissolving process on the other.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 24.12.2002

(4) not assignable

(125)

Type of experience

Human - Medical Data

Remark

Adverse effect: intravenous administration Yellowlees et al. and Greenfield reported a toxic reaction in two elderly people receiving DMSO intravenously for treatment of arthritis (three daily doses of 100 g of 20 % DMSO). In one patient there was serious illness including oliguria, hemolysis, tremor, and loss of consciousness. The second patient did not become ill. Both patients had changes in blood aspartate transaminase, hydroxy-butyrate

dehydrogenase, and creatine kinase, and elevation of blood creatinine and urea nitrogen. Prothrombin and partial thromboplastin times were significantly shorter.

Knott and van Rijswijk, who had treated patients similarly to Yellowlees et al but without toxic effect, suggest that the toxic reactions seen in the latter's patients were caused by the action of DMSO as a drug potentiator, and that the reaction may have been due to an enhancement of the

toxicity of quinine sulfate, indomethacin, or phenothiazine,

which patients were receiving at the time.

Another patient receiving intravenously cryopreserved autologous marrow blood (to which DMSO was added) for treatment of myeloblastic leukemia suffered a reaction to the administration. In addition to a DMOP in hemoglobin, the patient became agitated, pyrexic, hypotensive, and

developed tachycardia. Recovery occured but the patient died ten days later. O'Dommell et al believed the mixture was toxic (the patient received 35g of DMSO as a 10% solution), and suggested that DMSO interactions may be significant and

potentially dangerous. These doses are considerable.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 29.07.2003 (4) not assignable

(125)

Type of experience

: Human - Medical Data

Remark

: A 43-y-old Caucasian female applied DMSO to her lower

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abdomen for treatment of interstitial cystitis. She used 2 treatments separated by 1 1/2 h consisting of applications of 2 ounces of DMSO mixed with 2 ounces of distilled water on a clean white washcloth over a 6" x 12" skin area, for a total application of 132 g or 1.8 g/kg. The first application was from a bottle obtained from a family member and had been used by the family member with no reported ill effects. The bottle came from a store that sold undiluted DMSO. After the first application the patient experienced a garlic taste in her mouth, but had no improvement in her abdominal pain. The second application was from a previously unopened bottle from the same store; a pulverized 200 mg ibuprofen tablet was mixed into the solution. She again noted a garlic taste but no symptomatic improvement.

Within 24 h the patient developed fatigue and cyanosis, as well as dyspnea with mild exertion which did not worsen over time. She sought medical attention for her symptoms 10 d after the DMSO application and presented to the doctor's office with generalized cyanosis and a room air pulse oximetry of 42-47%. She was mildly dyspneic but fully alert. She was transferred to the emergency department where her oximetry increased to 54% on oxygen per nasal cannula. On admission she reported her current medications as docusate calcium, ibuprofen, amitriptyline, pseudoephedrine, valaciclovir, loratadine, gualfenesin, azelastine nasal spray, and "Yeast Fighters", in over-the-counter preparation containing lactobacillus acidophilus bacteria, all of which she had been taking regularly for several months prior to admission. In addition, she had taken lansoprazole for approximately 2 w, starting 11 d prior to onset of the cyanosis, alprazolam for a longer period on an as needed basis, and 200 mg phenazopyridine tid starting 3 d after the onset of cyanosis.

Her initial laboratory tests were remarkable for a 47% methemoglobin level on an Instrumentation Laboratory carbon monoxide-oximeter model 282, a hemoglobin of 9.4 g/dl, a hematocrit of 27.7%, and a reticulocyte count of 5.9%. She received 2 treatments with 1 mg methylene blue/kg iv without significant improvement in either her cyanosis or methemoglobin level. Repeat analysis on a blood sample drawn the day following admission using a Radiometer OSM-3 carbon monoxide-oximeter demonstrated a sulfhemoglobin level of 6.2% and a methemoglobin level of < 0.1%. During hospitalization the patient was transfused with 2 units of packed red blood cells. She was discharged on the third hospital day still on oxygen but with continued symptomatic improvement. Her G6PD level was 280 U/ trillion RBC (normal range 146-376).

Analysis of the DMSO solution from the second bottle did not reveal any contaminants. The first bottle of DMSO had been discarded and was not available for analysis. Adverse effect: Sulhemoglobinemia after dermal exposure

Source

Method

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

24.12.2002

: Human - Medical Data

Type of experience

: Male volunteer subjects, ages 21-55, were treated with DMSO,

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(27)

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applied as an 80% gel, to the skin in a single daily dose of 1 g/kg body weight. DMSO was applied daily for 12 weeks; a total of 38 subjects completed the entire study. A control group of 18 males, not exposed to DMSO, were studied in a similar manner. Physical and laboratory examinations were performed prior to the start of the study. Blood and urine were obtained from all subjects at 1,2,4,6,8,and 13 weeks. All subjects received a physical examination at the end of the study. All subjects were given ophthalmological examinations prior to initiation of the study, at weekly intervals during the study, and at intervals up to 18 months after treatment was terminated.

Result

Subjects treated with DMSO exhibited a characteristic respiratory odor, previously identified as dimethyl sulfide. a metabolite. A variable degree of skin reaction was observed, characterized by wheal and erythema, drying and scaling. Other side effects included some sedation, and

occasional bouts of insomnia and nausea. With the exception of eosinophilia, no significant

abnormalities were observed in blood chemistry, hematology, and urine analysis. DMSO had no significant effect on

pulmonary function, EKG or EEG.

Subjective eye complaints consisted of mild photophobia and foreign body sensations. Less frequently, tearing, blurring of vision, disturbances in peripheral vision, and tiring of the eyes were mentioned. These symptoms could not be substantiated by ophthalmological examination or testing and did not persist after DMSO was discontinued. There were no differences in accommodation, near point of convergence, ocular tension, cycloplegic retinoscopy, and cycloplegic refraction. Slit lamp examination revealed no abnormal corneal or lens changes. Visual function, and pupillary mydriasis to 1% tropicamide were not altered.

Remark Source Clinical study: dermal exposure and effects on eye

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 24.12.2002 (2) valid with restrictions

(22)(81)

Type of experience

Human - Medical Data

Remark

Clinical study: skin sensitization

No skin sensitization reaction was observed in 23 subjects after five 48-hour induction exposures to 75% DMSO solution (each one preceded by a 24-hour pre-treatment with 5% sodium lauryl sulphate) and challenge with 25% DMSO solution.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 24.12.2002 (2) valid with restrictions

(92)

Type of experience

: Human - Medical Data

Remark

Clinical study: effects on the renal function DMSO administered intravenously can protect experimental animals with massive stroke and brain swelling from mortality and neurologic impairment. Studies in patients suffering from cerebral trauma also suggest considerable efficacy. Intravenous DMSO was used to treat seven patients with stable spinal cord injuries. Because of drug-associated hemoglobinemia and hemoglobinuria, the patients were studied for subtle evidence of renal tubular dysfunction by serial

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measurements of urinary beta-2-microglobin excretion. No increases in tubular protein excretion or decreases in glomerular filtration rate were observed following short-term infusions of 10-40% DMSO. No significant short-term nephrotoxicity was observed from intravenous

DMSO.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 24.12.2002 (2) valid with restrictions

· (2) valid Will 1030

(10)

5.11 ADDITIONAL REMARKS

Type

: Distribution

Remark

A. Membrane penetration

In man radioactivity of 35S DMSO appeared in blood 5 min after cutaneous application. One hour later, radioactivity could detected in bones. This readily crossing depends on a reversible configuration change of the skin proteinic barrier occurring when DMSO substitutes for water.

B. Membrane transport

DMSO carry a wide range of substances through living membranes, vegetal or animal. Concentration of active

substance in DMSO are 0.5 to 90 %. Concentrations above 10 %

can be IRRITATING for skin by repeated application.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability

(4) not assignable

(83)

27.12.2002

: Biochemical or cellular interactions

Type Remark

Dimethyl sulphoxide (DMSO), at concentrations of 1-2%, induces terminal differentiation in several different cell types in vitro and enhances the growth of newborn mouse epidermal cells in primary culture under conditions that also permit terminal differentiation. We have found that DMSO concentrations approaching 4 % reversibly inhibited (with little quest toxicity) beginned differentiation of

DMSO concentrations approaching 4 % reversibly inhibited (with little overt toxicity) terminal differentiation of normal epidermal cells from newborn SENCAR mice. Cells cultured in medium containing 4 % DMSO and calcium in excess of 1 mM did not stratify extensively or slough large numbers of squamous cells or keratin bundles, as revealed by light and electron microscopy. The number of detergent insoluble cornified envelopes was similarly reduced. Long term growth of epidermal colonies in secondary culture was optimum in 1%

DMSO, this concentration also permitting normal terminal differentiation of these cells. Since DMSO had these effects on epidermal cells in vitro, it may also affect epidermal cell proliferation and terminal differentiation in vivo, an important consideration should DMSO ever be approved for terifolium in the Idea.

topical use in the US.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 12.09.2000 (2) valid with restrictions

(109)

Туре

Biochemical or cellular interactions

ld 67-68-5

Date 17.05.2004

(99)

Remark

Dimethyl sulfoxide (DMSO) can protect the liver from injury produced by a variety of hepatotoxicants when administered prior to or concomitant with the toxicants. This protective action has previously been attributed to DMSO-induced inhibition of bioactivation of the compounds to toxic intermediates. In these studies, the ability of DMSO to provide protection when administered 10 hr after a toxicant was evaluated in several animal models of xenobiotic-induced liver and kidney injury.

In the guinea pig model of halothane-associated hepatotoxicity, male outbred Hartley guinea pigs received 2 ml/kg DMSO 10 hr after an inhalation exposure to 1.0% halothane, 40% O2 for 4 hr. DMSO decreased the extent of liver necrosis as indicated by a threefold decrease in plasma alanine aminotransferase activity 48 hr after exposure and a reduction in the incidence and extent of zone 3 necrosis. These results do not appear to be due to alterations in halothane biotransformation since DMSO administered at 10 hr after halothane had no affect on plasma concentrations of the halothane metabolite trifluoroacetic acid or covalent binding by reactive halothane biotransformation intermediates to hepatic protein. In addition, administration of the structurally analogous biotransformation inhibitor diallyl sulfide at 10 hr after halothane also had no effect on biotransformation or covalent binding but provided no protection from liver injury. Hepatic glutathione concentrations in the guinea pigs 24 hr after halothane exposure were also unaffected by late treatment with DMSO. Further studies in male Sprague-Dawley rats demonstrated the ability of DMSO to decrease the hepatic injury resulting from oral administration of 1.0 ml/kg chloroform or 0.5 ml/kg bromobenzene when administered 10 hr after either toxicant. The chloroform-treated rats also developed renal tubular necrosis with large increases in plasma creatinine and urea nitrogen, which were completely ameliorated by DMSO.

Elucidating the mechanism(s) of this protective action of late DMSO administration should provide insight into the cascade of events that lead to liver and kidney injury from toxicants and, hopefully, therapeutic modalities for individuals suffering from acute, progressing, xenobiotic-induced hepatitis.

Source

: Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 29.07.2003 : (2) valid with restrictions

Type

Biochemical or cellular interactions

Method

DMSO has previously been shown to attenuate chloroform (CHCl3) and bromobenzene (BB) induced hepatotoxicity in the rat when a dose of 2.0 ml/kg is given 24 hr after the toxicants (Lind and Gandolfi, 1997). However, the optimal dose of DMSO and the latest time at which DMSO can be administered and still provide effective protection have not been determined. In order to determine the latest time at which DMSO can interrupt the development of necrosis, male Sprague-Dawley rats received either 0.75 ml/kg CHCl3 or 0.5 ml/kg BB, 20 % in corn oil, po, followed by single dose of 2 ml/kg DMSO, 50 % in saline, ip, at 24, 26, 28 or 30 hr

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later. Positive control groups received either CHCl3 or BB and then 4.0 ml/kg saline, ip, 24 hr later. All of the animals were then killed 48 hr after toxicant dosing. The extent of liver injury present when DMSO was administered was examined by killing animals at 24, 26, 28 or 30 hr after toxicant dosing. The optimal dose of DMSO for providing protection was estimated by administering either 0, 1.0, 2.0, 3.0 or 4.0 ml/kg DMSO at 24 hr after toxicant dosing and then killing the animals at 48 hr.

Result

Delaying DMSO administration to times later than 24 hr after toxicant dosing led to a loss of protection as indicated by both plasma ALT activity and the light microscopic appearance of liver tissue. The distinctive liver lesions present at 24 hr after CHCl3 or BB dosing rapidly expanded from being limited around central veins to bridging between centrilobular areas in only a few hours. This was accompanied by large increases in plasma ALT. With both toxicants, doses of DMSO greater than 2 ml/kg did not enhance its protective action while the lower dose of 1 ml/kg DMSO was not as effective. The loss of DMSO's antidotal action when given at times later than 24 hr after the toxicants indicates irreversible changes were underway as the centrilobular lesions progressed from being limited to more bridging in nature.

Source

: Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 12.09.2000

: (2) valid with restrictions

(100)

Type

: Immunotoxicity

Method

Male C3H/HeJ mice weighing between 20-30 g were used. In experiments requiring a comparison between two groups of animals (i.e., DMSO-treated vs. untreated), care was taken to match all groups relative to their whole body weight prior to the start of DMSO treatment and/or antigen injection. DMSO was mixed with tap water and mice were allowed to drink ad libitum. In a separate study, intraperitoneally administered DMSO was given in undiluted daily injections of 0.1 or 0.2ml. Immunization of mice was accomplished by a single intraperitoneal injection (0.1 ml) of 10% sheep red blood cells suspended in sterile saline.

The serum levels of IgM, IgG1, IgG2a, IgG2e, and IgA were

determined by radial diffusion assays using commercially, prepared assay kits and reference.

The number of direct (IgM) and facilitated (IgG1, IgG2b, and

The number of direct (IgM) and facilitated (IgG1, IgG2b, and IgA) plaques were evaluated for spleen cell suspensions using the microtechnique. Each serum was titrated for

hemagglutination. Total body weights and spleen weights were measured and the percent of the spleen weight to whole body weight

was calculated.

Result

The general effects of oral DMSO were first evaluated in normal uninjected animals. Age- and weight-matched mice were separated into four groups of 35 each (5 mice/cage) and given 1%, 2.5%, 5%, or 10% DMSO (V/V) in their drinking water. A fifth group served as controls and received ordinary tap water. All animals were allowed to drink ad libitum. The amount of food and water consumed were measured daily for six weeks. Mice refused to drink 10% DMSO and were eliminated from the study. By the 4th week, mice drinking 5% and 2.5% DMSO had increased daily fluid intake from 5.5

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Date 17.05.2004

ml/day/mouse (controls) to 23.5 ml/day/mouse and 10 ml/day/mouse, respectively. No difference was observed for the amount of food consumed when mice drinking 5% DMSO were compared with controls. During the six-week period, mice drinking 5% DMSO experienced a significant weight loss for both total body weight and spleen weight. The percent of spleen weight to total body weight also dropped significantly. No histological differences between controls and DMSO-treated mice were noted when stained paraffin sections of spleen, lung, kidney, liver, gut, and lymph nodes from the two groups were compared. Total serum volume, determined from hematocrit values, had dropped by 20% (P > .001) in the fourth week of 5% DMSO treatment. By the sixth week of treatment, however, the difference in serum volume was only 10% (P > 0.01).

Although an occasional test group revealed slightly elevated values for individual immunoglobulin classes, differences between treated and untreated animals were not significant. In a second study, the effect of intraperitoneal DMSO on serum immunoglobulin levels was evaluated for matched groups of 10 animals given daily intraperitoneal injections of 0.1 or 0.2 ml of undiluted DMSO. Alter three injections of 0.2 ml, only six animais survived and immunoglobulin concentrations were determined on the fourth day. Animals given 0.1 ml received 7 daily injections and their serum immunoglobulin levels were assayed on the 8th day. Surviving animals given 0.2 ml DMSO (8.8g/kg) demonstrated a 60-80% drop in IgG subclasses, a 64% drop in IgA, and a 50% drop in IgM. Animals given 0.1 ml (4.4g/kg) experienced a 30% drop for IgG subclasses and a 21% drop for IgA. IgM levels remained unchanged.

The primary humoral immune response was evaluated in two groups of 25 mice allowed to drink 5% DMSO ad libitum for 16 days or 8 weeks prior to (and during) the 7-day immunization period. The results were compared with an untreated control group of 25 similarly immunized mice. Five animals from each group were evaluated for PFCs, immunoglobulin concentrations, and hemagglutination titers on days 3-7 after SRBC injection. An additional 30 mice were included in both the 16 day pre-DMSO groups and controls for comparison of hemagglutination titers at 10-14 and 38 days after immunization. All three groups demonstrated an increase in spleen size measured on days 3, 4, and 5 after antigen injection. Spleens from DMSO-treated animals, however, remained significantly smaller than those of the untreated controls. On the basis of plaque-forming cells measured 3-7 days after antigen injection, DMSO-treated mice experienced inhibition of IgM on days 3, 4, and 5, IgG(s) on days 4, 5, and 6, and IgA on day 4 (and day 5 for 8-week DMSO group). This difference was further reflected in the hemagglutination titers, which were significantly lower in the treated population. Immunoglobulin levels were assayed daily 3-7 days after injection. Except for serum IgG, differences in immunoglobulin levels were not significant. When the two DMSO-treated groups were compared, plaque-forming cells and hemagglutination titers were significantly lower in mice having ingested DMSO for 8 weeks prior to immunization. The increased inhibition was also reflected by a 24-hr delay in the peak responsiveness of plaque-forming cells. No significant differences between the two groups were observed for serum concentrations of individual immunoglobulins.

ld 67-68-5 5. Toxicity Date 17.05.2004 The possibility that DMSO might influence the antigenicity of SRBC was studied by pretreating red cells with 10% DMSO for 24 hr at room temperature. Washed, treated cells were used to immunize normal mice and the results were compared with a group of mice similarly injected with untreated red cells. No significant differences were noted between the two mouse groups for plaque-forming cells or hemagglutination titers Remark The effects observed at high dose of DMSO, 5% in the drinking water (equivalent to a daily intake of 27 g/kg during the first week to 52 g/kg by the end of the third week), or 0.1 and 0.2 ml/animal by ip injection (equivalent to 5 and 10 g/kg, respectively) should be interpreted with caution taken into account the huge amount of DMSO administered. Atofina, Paris-la-Défense, France Source Atofina Paris La Défense Cedex Reliability (2) valid with restrictions (117)29.07.2003 : Immunotoxicity Type Method Autoimmune strain MRL/lpr, C3H/lpr, and male BXSB mice were placed on a continuous treatment regimen with 3% DMSO or 3% DMS02 in the drinking water, ad libitum, commencing at 1 to 2 months of age, before spontaneous autoimmune lymphoproliferative disease development could be detected. This represented doses of 8-10 g/kg/day of DMSO and 6-8 g/kg/day of DMS02. Plasma antinuclear antibodies were analyzed employing an indirect immunofluorescence assay with chicken erythrocyte nuclei as substrate. Serum IgG was measured by radial immunodiffusion utilizing a quantitative immunodiffusion kit. Direct antibody plaque formation was measured using spleen cells from C3H/lpr mice that had been injected ip with 5 X 10e8 washed sheep erythrocytes (SRBC) 5 days previously. Result Both compounds were observed to extend the mean life span of MRL/lpr mice from 5.5 months to over 10 months of age. All strains showed decreased antinuclear antibody responses and significant diminution of lymphadenopathy, splenomegaly, and anemia development. Serum IgG levels and spleen IgM antibody plaque formation, however, did not differ from control values. There was no indication of involvement of systemic immunosuppressive or antiproliferative effects, and treated animais were observed to remain healthy and vigorous with no signs of toxicity. These results demonstrate that high doses of both DMSO and its major in vivo metabolite, DMSO2, provide significant protection against the development of murine autoimmune lymphoproliferative disease. Source Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex Reliability (2) valid with restrictions 12.09.2000 (114)

Type

: Immunotoxicity

Method

Two groups of 8 female SW mice were injected i.p. daily with 2.5 g/kg 100% DMSO for one week, 1.3 g/kg every other day for the next week (because they appeared weak) and 2.5 g/kg daily for the following 3 weeks. Control mice received identical amounts of sterile saline by the same route. All

i. Toxicity	ld 67-68-5 Date 17.05.2004	1
	mice were immunized sc with 0.05 ml 5% sheep red blood cells on days 13 and 24, and bled twice by caudal incision on days 20 and 29. Anti-SRBC hemagglutination titers were determined by doubling dilutions; leukocyte counts, hematocrits, and organ weights (liver, lungs, spleen, thymus, kidneys, and heart) were measured by standard methods. The experiment	
Result	ended after 36 days of treatment. In DMSO-treated mice, haematocrits were significantly decreased (p<=0.002) but still within the normal range. The primary and secondary antibody response to sheep red blood cells, leukocytes counts, body weight, and the size of the heart, lungs, spleen, thymus, and kidneys were not affected. DMSO treatment resulted in significant liver enlargement (p=0.02). It is concluded that DMSO is not deleterious to the humoral immune response in mice responding to a new	
Source	antigen: Atofina, Paris-la-Défense, France	
	Atofina Paris La Défense Cedex	
Reliability 12.09.2000	: (2) valid with restrictions	(29)
Туре	: other: initiation/promotion study	
Remark Source Reliability	: The effects of DMSO on the tumorigenic activity of dimethylbenz[a]anthracene (DMBA) was investigated in rats. Two groups of 50 male Sprague-Dawley rats were given 20 mg DMBA by gavage. DMSO (50 ppm the drinking water) was started 3 days before or 3 days after DMBA administration and administered for 18 months. A third group received no DMSO and served as untreated controls. DMSO had no beneficial or deleterious effect on the latency of the tumours induced by DMBA nor on the tumour frequency. Rats receiving DMSO weighed more and had fewer tumors than did the controls at the end of the 18-month study period. This was suggestive that DMSO decreased the total number of tumors, although the difference between treated and control rats did not reach statistical significance: Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex (2) valid with restrictions	
24.12.2002	,,	(65)
Туре	: other: initiation/promotion study	
Remark	: In 20 ICR/Ha Swiss mice, dermal application of 0.1 ml DMSO, 3 times weekly over a period of 400 days, after a primary treatment with DMBA (applied once only, 20μg in 0.1 ml acetone), induced no skin tumours.	
Source	: Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex	
Reliability 24.12.2002	: (2) valid with restrictions	(102)
Туре	: other: initiation/promotion study	()
Remark	: The role of dimethyl sulfoxide ((DMSO) CAS: 67-68-5) in experimental tumorigenesis was investigated because of conflicting reports in the literature ranging from inhibition to no effect to enhancement. With the use of numbers of skin tumors produced on the back of the mouse following topical applications of carcinogenic agents as the variable and with acetone serving as the control solvent,	

ld 67-68-5

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the following results were obtained: When DMSO was the solvent for benzo(a)pyrene (CAS: 50-32-8) in the single-stage model (C3H mice), tumor numbers doubled. When DMSO was the solvent for 7,12-dimethylbenz(a)anthracene (CAS: 57-97-6) serving as initiator in the two-stage model (CD-1 mice), tumor numbers were unaffected. In the two-stage model, when DMSO was the solvent for the potent promoter phorbol-12-myristate-13-acetate ((PMA) CAS: 16561-29-8) or was applied to skin at the initiation site (the back) before PMA, tumor numbers were reduced to one-third of control. However, when DMSO was applied before PMA to the abdomen, a site remote from initiation, tumor numbers doubled. Enhancement of PMA appears to be unique. Recognition that diverse effects can occur depending on the method of application of DMSO may help to decipher the conflicting literature on its relation to tumorigenesis, could be of value in probing the mechanism of tumor promotion, and might signal further caution in its clinical use.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 24.12.2002 (2) valid with restrictions

Type

other: initiation/promotion study

Remark

The effect of intravesical instillation of dimethylsulfoxide (DMSO) on bladder carcinogenesis was examined in mice. Experiment 1: Fifty- five female C3H/He mice were administered 0.05% N-butyl-N-(4-hydroxy-butyl) nitrosamine (BBN) in their drinking water for 8 weeks. In week 9 they were divided into two groups consisting of 25 mice each. Then, under nembutal anesthesia the first group was given weekly intravesical inatillations of 0.1 ml DMSO (minimum 99.0%) for 10 weeks. The second group received no treatment except anesthesia. All mice were killed 30 weeks after the begining of the experiment and their urinary bladder resected for histological examination. The incidence of bladder carcinoma was 93.7% (15/16) and 27.7% (6/22) in groups 1 and 2, respectively. These incidences differed significantly between the two groups. Experiment 2: One hundred and twenty female C3H/He mice were divided into two groups. The first group was given 0.05% BBN in their drinking water for 5 weeks and then tap water. The second group was not given BBN. In week 6, the first group was divided again into three groups (1, 2 and 3) consisting of 28, 26, and 27 mice, respectively. The second group was divided into groups 4 and 5 consisting of 21 and 18 mice, respectively. Under nembutar anaesthesia groups 1 and 4 received weekly intravesical instillation of 0.05 ml DMSO (minimum 99.0%) from weeks 6 to 13, Group 2 received weekly intravesical instillation of 0.05 ml distilled water from weeks 6 to 13. Groups 3 and 5 received no treatment except anesthesia. 25% of the group 1 mice developed bladder carcinomas compared to 0% in the controls.

Source

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

Reliability 29.07.2003 (4) not assignable

(120)

(85)

Type

other: General pharmacology

Remark

: GENERAL PHARMACOLOGY

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DMSO has stimulated speculation and imagination but few useful and incontrovertible data have been generated. DMSO's affinity for water, its superiority as a solvent, and its ability to trap oxygen related free radicals are firmly established. DMSO's effectiveness as a penetrant, carrier, cryoprotectant, radioprotectant and as an antiischemic, antiinflammatory and analgesic agent are well established, but the mechanisms are incompletely understood. DMSO's systemic toxicity and teratogenicity are considered low. DMSO's local toxicity may be significant, depending on dose, route, species and individual variation. Combinations of DMSO with other agents may be dangerous. Potential complications should be considered, and investigated or avoided. Side effects of DMSO, such as mast cell degranulation, diuresis and volume depletion may exacerbate patient's preexisting problems, and should be considered before treatment is initiated.

- Only veterinary preparations of DMSO should be used. The industrial solvents may contain impurities that can endanger health
- 2) DMSO should be kept in an airtight bottle, and the bottle should be tightly closed when not in use. Exposed DMSO is rapidly diluted by water in the air, and becomes less effective.
- 3) DMSO should only be applied to a clean, dry, unmedicated skin surface. DMSO can carry unwanted substances through skin, into the body. DMSO's reaction with excess water on damp skin can produce more heat and discomfort than is necessary.
- 4) DMSO should be applied with sterile or clean cotton in order to minimize contamination with potentially dangerous substances and to minimize human exposure. Wearing rubber gloves will also help reduce human exposure to contaminants and to DMSO itself, but some compounds in solution with DMSO will even penetrate rubber gloves.
- It is unfortunate when a potentially useful drug cannot be added to the approved medical arsenal because sufficient information is not available. Similarly, ineffective or dangerous drug applications should be exposed by well documented case reports and by well designed investigations. The medical professions, the patients and the drug deserve more thorough investigation of DMSO's therapeutic potential,

and its interactions with other drugs and disease states. Current evidence from in vitro situations and from laboratory animals suggests that DMSO may prove to be especially valuable in improving outcomes of intestinal surgeries, and in treatment of transient ischemic insults to organs or systems. Other areas that may be worthy of investigation by and for the veterinary profession include: post operative patient comfort and wound healing; ulcerative wound healing; acute swelling due to injury in other than equine species; reduced metabolism and damage in ethylene glycol intoxication

because of alcohol dehydrogenase inhibition by DMSO; therapeutic combinations of DMSO with other drugs - antimicrobials, cytotoxic drugs in cancer chemotherapy, steroids; dangerous combinations of DMSO with other drugs; dangerous interactions of DMSO with some disease states (e.g. mastocytoma).

Atofina, Paris-la-Défense, France Atofina Paris La Défense Cedex

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Source

5. Toxicity ld 67-68-5 Date 17.05.2004 Reliability : (4) not assignable 29.07.2003 Remark A. MEMBRANE PENETRATION - TRANSPORT DMSO readily crosses most tissue membranes of lower animals and man. This property depends on a reversible configuration change of the protein occuring when DMSO subsitutes for water. DMSO in combination with electrolytes reduced the electrical resistance of the skin by facilitating the absorption of these electrolytes which it was itself being absorbed. B. EFFECT ON COLLAGENE After immersion in DMSO, the collagen fraction extractable with neutral salt solution was significantly decreased. C. ANTIINFLAMMATION Some antiinflammatory effects were demonstrated with intraarticular DMSO in rabbits following the creation of experimental (oil) arthritis. D. ANALYSIS DMSO produces analgesia by acting both locally and systemically, with a longer duration of action than morphine 6 hr vs 2 hr respectively. E. BACTERIOSTASIS Bacteriostasis from DMSO occures due to a loss of RNA conformational structure required for proteins synthesis. F. DIURESIS With the important increase in urine volume, there was an increase in Na and K excretion. G. ENHANCEMENT DMSO increase the effectiveness of Griseofulvin and potentiates the action of Digoxin.

H. CHOLINESTERASE INHIBITION In vitro assays 0.8 - 8 % DMSO inhibits bovine erythrocyte cholinesterase 16 - 18 %.

I. VASODILATATION DMSO possess potent histamine liberating properties. J. MUSCLE RELAXATION

DMSO applied topically to the skin of patients produces electromyographic evidence of muscle relaxation after 1 hr. K. ANTAGONISM TO PLATELET AGGREGATION

DMSO is a good antagonist in vitro and in vivo.

L. ENHANCEMENT OF CELL DIFFERENTIAITON AND FUNCTION DMSO stimulates cyclic AMP accumulation and lipolysis and decreases insulin stimulated glucose oxydation in free white fat cells of rat

M. RADIOPROTECTIVE AND CRYOPROTECTIVE ACTION DMSO possess such properties widely used.

N. PROTECTION AGAINST ISCHEMIC INJURY

Studies have reported that DMSO can increase the synthesis of PGE1, a moderate vasodilator. It also inhibits the calcium-induced release of nor adrenaline in nerve terminals. These actions are likely to be involved in its

ability to protect against ischemic injury.

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(4) not assignable

Source

Reliability 29.07.2003

(83)

(21)

6. Analyt. Meth. for Detection and Identification	•	67-68-5 17.05.2004	
6.1 ANALYTICAL METHODS			
6.2 DETECTION AND IDENTIFICATION			

/. Eπ.	Against Target Org. and Intended Uses		67-68-5
		Date	17.05.2004
7.1 F	UNCTION		
7.2 E	FFECTS ON ORGANISMS TO BE CONTROLLED		
7.3	ORGANISMS TO BE PROTECTED		
7.4 l	USER		
7.5 F	RESISTANCE		
7.5 F	IESIS I AIVE		
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8.	Meas.	Nec.	to	Prot.	Man.	Animals.	Environment
•					man,	milliais,	THE STREET

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- 8.1 METHODS HANDLING AND STORING
- 8.2 FIRE GUIDANCE
- 8.3 EMERGENCY MEASURES
- 8.4 POSSIB. OF RENDERING SUBST. HARMLESS
- 8.5 WASTE MANAGEMENT
- 8.6 SIDE-EFFECTS DETECTION
- 8.7 SUBSTANCE REGISTERED AS DANGEROUS FOR GROUND WATER
- 8.8 REACTIVITY TOWARDS CONTAINER MATERIAL

9. References ld 67-68-5 Date 17.05.2004 (1) Abbondandolo, A., Bonatti, S., Corsi, C., Corti, C., Fiorio, R., Leporini, C., Mazzaccaro, A. and Nieri, R. (1980) the use of organic solvents in mutagenicity testing, Mutation Res., 79, 141-150. (2) Amacher, D. E. and Zelljadt, I. (1984) Mutagenic activity of some clastogenic chemicals at the hypoxanthine guanine phosphoribosyl transferase locus of Chinese hamster ovary cells, Mutation Res., 136, 137-145. Amacher, D. E., Paillet, S.C., Turner, G.N., ray, V.A. and (3) Salburg, D.S. (1980) Point mutations at the thymidine kinase locus in L5178Y mouse lymphoma cells. II. test validation and interpretation. Mutation Res., 72, 447-474. (4) ANDREAE, M.O., 1980. Anal. Chem, 52, 150-3. ANDREAE, M.O., 1980.Anl. Chem., 52, 150-3. (5) Anton O, Rouxhet PG; Clay Miner 25: 259-63 (1977). (6) ATKINSON, R., 1989. Kinetics and mechanisms of the gas-phase (7)reactions of the hydroxy radical with organic compounds. Journal of physical and chemical reference data. Monograph N°1. BARBOSA,I.R., MARTINS, R.M., SA E MELO M.L. and SOARESA.M.V.M., 2003. Acute (8) and chronic toxicity of Dimethylsulfoxide to Daphnia magna. BULL ENVIRON CONTAM TOXICOL, 70, 1264-1. BAROHANA-GOMARIZ, M.V. et al, 1994. Acute toxicity of organic solvents on Artemia (9) salina.Bull. Environ. Contam. Toxicol., 52(5), 766. Bennett WM and Muther RS (1981) Lack of nephrotoxicity of (10)intravenous dimethylsulfoxide. Clinical Toxicol. 18, 615-618. BENVILLE, P., 1974. Acute toxicity of nine solvents to (11)rainbow trout fingerling (unpublished, transmitted from Tiburon Laboratory, NOAA, july 10. (12)BENVILLE, P.E. et al, 1968. Some toxic effects of dimethyl sulfoxide in salmon and trout. Toxicol. Appl. Pharmacol. 22: (13)Biodegradation and bioaccumulation data of existing chemicals based on the CSCL (Chemical Substances Control Law). Edited by Chemicals Inspection and Testing Institute, Japan, October 1992. (14)Biodegradation and bioaccumulation data of existing chemicals based on the CSCL (Chemical Substances Control Law). Edited by Chemicals Inspection and Testing Institute, Japan, october 1992.

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